

The Brazilian Journal of INFECTIOUS DISEASES



www.elsevier.com/locate/bjid

Original Article

Phase I safety and immunogenicity study of a Brazilian serogroup B vaccine



R. Menezes Martins a,1 , A.R.S. Périssé b , L.A.B. Camacho b , M.L. Leal c , M.L.S. Maia a , A. Homma d , E. Jessouroun b c,*

- ^a Clinical Advisory Unit, Bio-Manguinhos/Fiocruz, Rio de Janeiro, RJ, Brazil
- ^b National School of Public Health, Fiocruz, Rio de Janeiro, RJ, Brazil
- ^c Bacterial Technology Laboratory, Bio-Manguinhos/Fiocruz, Rio de Janeiro, RJ, Brazil

ARTICLE INFO

Article history: Received 12 July 2021 Accepted 21 October 2021 Available online 16 November 2021

Keywords:

Meningococcal vaccine
Outer membrane vesicles

ABSTRACT

Meningococcal disease by serogroup B has been a public health problem in Brazil in the last decades. The Brazilian Oswaldo Cruz Foundation has been working to develop a vaccine with detergent-treated outer membrane vesicles (OMV) and detoxified endotoxin (dLOS) from Neisseria meningitidis serogroup B prevalent strains. A phase I study, enrolling 26 adults (18-44 years of age) was performed using experimental vaccines combining B components and aluminum hydroxide as adjuvant. It was a dose escalation study testing vaccines made of 25, 50, and 100 μg OMV protein/mL (sum of both strains) and dLOS in half amount of total protein concentration, with three doses given two months apart. Adverse events were mild/moderate with frequency increasing with the amount of antigens. Pain in the site of injection was the most frequent reaction in all doses, reported in more than the 85% across vaccine groups. Considering all injections, cephalea was the most common systemic adverse event, detected in 11.1%, 17.2% and 32.1%, respectively with doses of 12.5 μ g, 25 μ g and 50 μ g. High titers of total IgG (ELISA) were observed for the vaccine components before vaccination. Protective levels of bactericidal antibodies (titer ≥1:4) for both vaccine strains were also present. Considering a 4-fold increase of IgG titers compared to pre-immune values (seroconversion), 50%-70% of those who received intermediate and highest doses of antigens presented satisfactory response for OMV of N44/89 strain. The lowest dose vaccine induced no seroconversion for strain N44/89, and 11% for strain N603/95. For the three vaccines doses, 25% of seroconversion, in total IgG against LOS, was observed. Increased antibody bactericidal activity was observed for both strains in higher antigen concentrations. For IgG against LOS, all vaccine formulations showed 25% of seroconversion. In conclusion, MenB-Bio experimental vaccines were well tolerated and immunogenic, thus allowing phase II studies.

© 2021 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

E-mail address: ellen@bio.fiocruz.br (E. Jessouroun).

^d Bio-Manguinhos/Fiocruz, Rio de Janeiro, RJ, Brazil

^{*} Corresponding author.

¹ in memoriam.

Introduction

Meningococcal disease has been an important public health issue in Brazil, with endemic occurrence and recurrent epidemics and outbreaks since 1920. 1-4 Serogroups B, C, Y and W of the Neisseria meningitidis have been the most frequently identified in meningococcal invasive disease in Brazil in the last two decades (http://www.saude.gov.br/ boletins-epidemiológicos). Following the introduction of the meningococcal C vaccine in the Brazilian basic schedule of immunization in 2010, the proportion of cases of serogroup C decreased from 40% to 27% in 2018, whereas serogroup B increased from 7% to 15%, with the caveat that the proportion of cases with missing serogroup remained approximately 50%.5-7 Serogroup B was initially the most important, with the phenotype, B:4,7:P1.19,15; L3,7,9 responsible for an epidemic in 1988 in São Paulo. This serosubtype still represented the predominant B phenotype from 2000 to 2011 and seems to remain the main strain nowadays.8,9

A universal meningococcal vaccine has been pursued over the years due to the severity and high mortality of these infections. For serogroups A, C, W and Y conjugated polysaccharide vaccines have been successfully used not only for the induction of protective bactericidal antibody titers but also for reducing the pathogens circulation by inducing herd immunity. 10-12 For serogroup B, capsular polysaccharidebased vaccines have not been the strategy pursued because of the similarity of the polysaccharide structure to human cell sugars, which represents a potential risk for autoimmune response induction.¹³ Subcapsular antigens and, more recently, proteins with important roles in the onset of the disease, obtained by reverse vaccination, have been studied as new vaccine targets. 14,15 Outer membrane vesicle-based vaccines (OMVs) have been evaluated in efficacy trials in Cuba, Brazil, New Zealand, and Norway. Efficacy has been demonstrated for adolescents and children aged more than four years. In a Brazilian study, protection against meningococcal disease caused by serogroup B was not elicited in younger children. The protection provided by OMV vaccines is associated with serum bactericidal antibodies which appear to be immunodependent on Por A surface protein. This characteristic is responsible for the specific serosubtype response induced by these vaccines. 16-19

Following the epidemiological situation of meningococcal disease in Brazil since late 1990's three Brazilian research institutions (Bio-Manguinhos/Fiocruz, Instituto Butantan and Instituto Adolfo Lutz) have collaborated to develop a group B meningococcal vaccine consisting of detergent treated OMV and detoxified endotoxin (dLOS) from Brazilian prevalent strains. From the early stages of development to the production process standardization, the two most prevalent meningococcal B serosubtypes in Brazil were B:4,7:P1.19,15:L1,3,7,9 (62%) and B:4,7:P1.7,1:L3,7 (9%), accounting for approximately 70% of all circulating serosubtypes. In order to develop a tailor-made vaccine, Bio-Manguinhos has developed a bivalent vaccine candidate (MenB-Bio) including OMV and dLOS from these prevalent strains, and dLOS with aluminum hydroxide as adjuvant. Good immunogenicity and protection against challenges in mice were demonstrated.²⁰ The results

supported initiation of Phase I clinical studies with MenB-Bio.²¹ The detoxified endotoxin was extracted from vaccine strain B:4,7:P1.19,15. Herein, we present the results of a phase I clinical study with this vaccine candidate to assess safety and to obtain preliminary information on immunogenicity of MenB-Bio.

Materials and methods

Vaccine components

The bacteria growth and biomass separation were performed following Good Manufactoring Practice (GMP). Vaccine strains N44/89 (B:4,7:P1.19,15,L3,7,9) and N603/95 (B:4,7:P1.7.1,L3,7) were provided by Adolfo Lutz Institute, São Paulo, Brazil. The production process included cultivation of vaccine strains in 150 L bioreactor (B Braun Model Biostat UD 100) in Catlin MC.6 with 20 $\mu\mathrm{M}$ of Fe⁺³ and 42 $\mu\mathrm{M}$ of Ethylenediamine-di – (Ohydroxy-phenylacetic acid) (EDDHA) to obtain outer membrane vesicles (OMV) and without EDDHA for lipooligosaccharide (LOS) production.²² In both processes bacteria growth was inactivated by heat (56°C/30 min) and the biomass separation was performed by continuous flow centrifugation (LAPX 4045GP-31G, Alfa Laval). The LOS depleted OMVs from vaccine strains were DOC-extracted from concentrate supernatant by tangential ultrafiltration in Centrasette stainless steel cassette holder with 0.9 m² polyethersulphone Centrasette cassette and a molecular weight cutoff of 100 kDa (Pall Corporation). After the concentration step, OMV was treated with 2% (w/v) sodium deoxycholate (DOC) and harvested by ultracentrifugation (100,000 g for 2 h). The concentrate bulk was sterile-filtered using 0.22 μm Millipak®-200 with membrane polyvinylidene fluoride (PVDF- Millipore). The raw LOS was extracted from total biomass culture from N44/89 strain after hexadecyl trimethyl ammonium bromide (Cetavlon) treatment.²³ The extracted LOS was further purified by gel filtration chromatography using 20 mM Tris-HCl, pH 8.5, containing 0.5% DOC and 5 mM Ethylenediaminetetraacetic acid (EDTA) as mobile phase on a single Sephacryl HR S-300 column (ID: 5.0X L: 100.0 cm). The LOS sample solution applied to the column was limited to 4% of bed volume and a LOS mass not higher than 600 mg. The purified LOS was detoxified (dLOS) with 0.2N NaOH in a water bath at 60°C/150 min.²⁰

Experimental vaccines

Three formulations of the candidate vaccine used in the phase I study were prepared as follows. Vaccine 1: 25 μ g of OMV protein (12.5 μ g from N44/89 and 12.5 μ g from N603/95) plus 12.5 μ g of dLOS (N44/89) per mL; Vaccine 2: 50 μ g of OMV protein (25 μ g from N44/89 and 25 μ g from N603/95) plus 25 μ g of dLOS (N44/89) per mL; Vaccine 3: 100 μ g of OMV protein (50 μ g from N44/89 and 50 μ g from N603/95) plus 50 μ g of dLOS (N44/89) per mL. Those formulations had been tested in preclinical studies and were analogous to a commercially available vaccine. The vaccine candidates were lyophilized and reconstituted with diluent Al (OH)₃ (2mg/mL) at the time of administration. Vaccine production and "in process" quality control were carried out in a GMP pilot plant production by

the Quality Control Department from Bio-Manguinhos/Fiocruz.

Study design, safety assessment and monitoring

Vaccines were administered in three 0.5 mL intramuscular injections in the deltoid region at 8-week intervals (6-10 weeks acceptable). A sequential protocol was used in order to maximize safety for the volunteers. The first 10 volunteers received an injection containing 6.25 μg of each of the two strains of meningococcal B plus 6.25 μ g of dLOS per dose, and were followed up for one month (group 1) after each dose to ascertain adverse events. Only after the first concentration was considered safe for human use after 30 days of follow-up, the second group of 10 volunteers was vaccinated with injections containing 12.5 μ g of each of the two strains of meningococcal B plus 12.5 μg of dLOS (group 2). Likewise, only after 30 days of observation of this group disclosed no severe adverse events vaccination of the third group of 10 volunteers (25 μ g of each of the two strains of meningococcal B plus 25 μ g of dLOS - group 3) was started. After vaccination, study subjects stayed under observation at the Unit during two hours. Health professionals were trained for medical assistance in case of need, especially for anaphylactic reactions. Diary cards, telephone contacts, and visits to the trial unit were used to check for local and systemic adverse events. Diary cards were supplied to all volunteers so that they could record any adverse event for seven days after each injection. They were asked to bring the diary when they returned for clinical assessment on day 7. Another diary was then given for the period between day 7 and day 60. All subjects were given thermometers and oriented on how to measure and record the axillary temperature six and 12 h after vaccination and daily for seven days. Local and systemic adverse events (any sign or symptom) were evaluated in site by physical examination and interview on days 2 and 7 after each vaccine dose, and on day 30 after the last shot. Adverse events were also ascertained by telephone at days 1, 3, 15 and 30 after each immunization (except 30 days after the last shot when volunteers returned to the trial unit for their last study interview). Abnormal laboratory test results were evaluated by comparing baseline data collected prior to the first immunization to those obtained from blood samples taken 48 h after each shot. Blood samples were analyzed for full blood count, coagulation studies, platelet concentration, liver and renal function tests, and C-reactive protein. Autoantibodies were tested at screening and 30 days after the last vaccine dose. The severity of adverse events was scored as 0 (absent); 1 (mild, easily tolerated); 2 (moderate discomfort, could interfere with some activities); 3 (intense, unable to perform usual activity), and 4 (life-threatening events). Fever was defined as axillary temperature ≥37.0°C. Hospital facilities were made available to manage severe adverse events. The study was approved by the Research Ethics Committee of the Instituto de Pesquisa Clínica Evandro Chagas (CAAE-0047.0.009.000-05) and was registered at the International Standard Randomized Controlled Trial Number (ISRCTN 75538667). An independent External Monitoring Committee, constituted by medical experts in clinical studies and meningococcal vaccines evaluated and followed up all

study procedures to ensure the safety of participants and scientific integrity of the study.

Serum bactericidal assay (SBA)

The strains N44/89 and N603/95 were used as the targets in SBA. Bacteria suspension from work seed lots prepared in Bio-Manguinhos, was streaked on a Columbia blood agar (Merck 1.10455.0500) with 5% horse blood (CBA) and incubated overnight at 37°C with 5% CO₂. Afterwards, it was subcultured onto another CBA plate and incubated for 4 h at 37°C with 5% CO₂. After 4 h growth the bacteria concentration was adjusted to 2 imes 10 organisms/mL. Equal volumes (10 μ L) of the bacterial suspension and human complement were added to 20 μ L heat-inactivated test serum serially diluted twofold in bactericidal buffer in 96-well U-bottom microtiter plates (Greiner, Frickenhausen, Germany). The SBA were performed on serum samples from volunteers before immunization schedule (T0), before the second dose (T60) and one month after the third dose (T90) of immunization. Plasma selected from a suitable donor from Hemorio Hospital, Rio de Janeiro, Brazil) was used as the complement source at a 25% final concentration after 60 min of incubation. The number of CFU was determined at time zero and after 60 min of incubation, by allowing 10 μ L of the reaction mixture to flow 8-10 cm, in lanes, down a CBA plate (the tilt method). After overnight incubation at 37°C with 5% CO₂ colonies were counted. SBA titers were expressed as the reciprocal of the final serum dilution step giving 50% killing at 60 min compared to the number of CFU at time zero. The assay used three in-house serum controls, with low, media and high titers for each target strain. 24,25

Enzyme-linked immunosorbent assay (ELISA)

To detect anti-OMV total IgG, 96-well plates (ref. no. 3590; Corning-Costar, Corning, NY, USA) were coated with 100 μ L/ well of OMV (4 μ g/mL) from N44/89 or N603/95 strains. Serum samples in two-fold serial dilutions in TBS with 0.05% Tween 20 (Merck, Schuchardt, Germany) and 5% FCS (Sigma-Aldrich, St. Louis, MO, USA) were incubated overnight at 4°C. The plates were incubated with anti-mouse IgG conjugated with alkaline phosphatase (whole molecule) (Sigma-Aldrich A-3688) diluted 1:2000. All sera were titrated in duplicate, and the titers were determined at an absorbance of 405 nm using a VERSA max tunable microplate reader (Molecular Devices, Sunnyvale, CA, USA). As the antibody standard, a positive post-vaccination serum was used in all experiments. The observed optical density was transformed to arbitrary units per milliliter by a sigmoidal standard curve (logit-log transformation) calculated from the values (1000 EU/mL) of the reference serum. To detect anti LOS total IgG, microtiter plates (Immulux ref 1000; Dynex Technologies, Chantilly, VA, USA) were coated overnight at room temperature with 100 μ L/well of 10 μ g/mL LOS-DOC micelles dissolved in PBS with 0.1% DOC. TBS with 5% bovine serum albumin (BSA) (Sigma-Aldrich ref A9418) was used as the blocking buffer for 2 h at 37°C. Sera samples were titrated in duplicate in TBS with 5% BSA and incubated for 3 h at 37°C. Conjugated antibodies (Sigma-Aldrich A-3688) were diluted 1:1000 and incubated for 2h at 37°C. The reaction was developed for 20 min with a

phosphatase substrate (Sigma-Aldrich S0942). All sera were titrated in duplicate and titers were determined with absorbance at 405 nm in VERSA max tunable microplate reader – Molecular Devices. The total IgG antibody responses (EU/mL) against OMVs and LOS, were presented as geometric mean concentrations (GMC) of five serum samples (pool of five mice) by using a 4-parameter logistic curve-fitting analysis with SoftMaxPro software and documentation - Molecular Devices, against the standard curve generated by in-house serum with 1000 EU/mL of arbitrary units. 26,27

Statistical analysis

- Univariate analysis described the frequency distribution of sociodemographic characteristics of the participants.
- Safety. The frequency of local and systemic adverse events were reported by group, dose, severity, and timing of occurrence.
- Immunogenicity. The proportion of participants with SBA ≥8 and seroconversion (ELISA) to each meningococcal strain was calculated, by vaccine dose. Criteria for seroconversion were bactericidal titer <1/8 before vaccination and ≥1/8 after the third dose or ≥4 fold increase on bactericidal titers. The geometric mean of the reciprocal dilution (bactericidal titers) and Elisa units were estimated.

Statistical analyses were performed with PASW Statistics 18 (Chicago: SPSS Inc.)

Subjects and ethical aspects

The study was a Phase I, unblinded, ICH/GCP compliant trial in 30 healthy adult volunteers aged 18-44 years, conducted from January 2006 to January 2007, at a special unit for immunobiological studies, which is an annex of an infectious

disease hospital at the Fiocruz campus in Rio de Janeiro. Subjects were Fiocruz employees, not involved directly in the production of the vaccine. After reading, understanding and agreeing with an Informed Consent Form, potential volunteers were screened for their health status through their medical history, medical examination, routine biochemical and hematological tests, chest X-Rays and electrocardiogram. Before each dose of the vaccine, women were screened for pregnancy and HIV seropositivity, which was also an exclusion criterion. The study was conducted in accordance with the Declaration of Helsinki²⁸ (World Medical Association Declaration of Helsinki 2001) and the European Clinical Trials Directive (European Clinical Trials Directive 2001)²⁹ and the Brazilian legal and regulatory requirements (http://bvsms. saude.gov.br/1996/res0196_10_10_1996.html).30 The trial was approved by a research ethics committee (registration number CAAE-0047.0.009.000-05 at Institute of Infectology Evandro Chagas) and the Brazilian Regulatory Agency (Proc. ANVISA number: 25351.175427/05-24). The research protocol was registered at the International Standard Randomized Controlled Trial Number (ISRCTN no.: 75538667).

Results

From January 2006 to January 2007, 150 individuals were contacted, 57 agreed to participate, 30 were eligible and were assigned to one of the three groups (Fig. 1). In the 12.5 μg group eight volunteers received all three doses, one received one dose and one received two doses. In the 25 μg group, nine volunteers received all three doses, and one received only two doses. In the 50 μg group nine volunteers received all three doses and one received only one dose. Twenty six volunteers were adherent to the protocol. Study subjects were mostly young adults in all vaccine groups. Distribution by sex

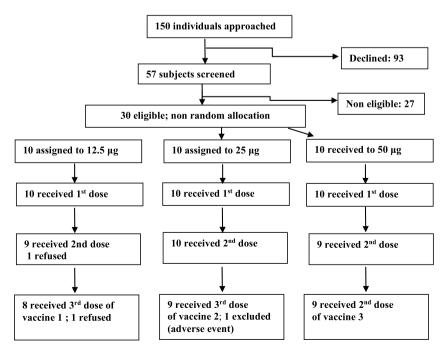


Fig. 1-Flow chart of participants in meningococcal B candidate vaccine trial.

Table 1 – Age and sex distribution of participants by vaccine group.

Group	Se	ex.	Age, y	Age, years		
	M	F	Median	Range		
12.5 μg	6	4	28	22-36		
$25~\mu \mathrm{g}$	5	5	36	24-43		
$50~\mu \mathrm{g}$	10	-	29	25-44		
All	21	9	30	22-44		

was balanced in all but the 50 μ g group, which included only men (Table 1).

There were no serious adverse events. Of the four volunteers who left the study before completing procedures, only one was due to an adverse event, in the 25 μ g group, after the second dose, who presented local pain grade 3, local and thoracic exanthema, and moderate fever.

Safety

All solicited local and systemic adverse events occurred in the first three days after vaccination and were of low or moderate intensity. Local pain in the site of injection was the most frequent reaction in all doses and vaccine groups. In the 50 μg group it occurred with similar frequency after all doses, whereas in the other vaccine groups the frequency decreased substantially in the second and third doses (Table 2). There was an increasing trend for local adverse events after larger doses, except for induration.

There was also an increasing trend for more intense adverse events after larger doses. Moderate or intense adverse events made up 13.6%, 17.3% and 30.3% of adverse events after 12.5 μ g, 25 μ g, and 50 μ g doses, respectively (Table 4). Pooling together injections for each vaccine group, cephalea was the most common systemic adverse event, detected in 11.1%, 17.2% and 32.1%, respectively in groups 12.5 μ g, 25 μ g, and 50 μ g. Axillary temperature \geq 37.0°C was also frequent especially after the first dose of the 50 μ g group (Table 3). Axillary temperatures ≥37.5°C occurred in 3.7%, 10.3% and 17.9% of all vaccinations, respectively in groups 12.5 μ g, 25 μ g and 50 μ g (data not shown). Malaise and myalgia were also more frequent following the 50 μg vaccine (Table 4). The highest axillary temperatures were 37.5°C, 38.0° C and 38.5°C for the 12.5 μ g, 25 μ g and 50 μ g groups, respectively. There was a trend for sooner occurrence and longer duration of adverse events with larger doses.

Vaccine group	Dose	Local pain	Erythema	Induration	Edema
	1st (n = 10)	10	1	2	0
12.5μg	2nd(n = 9)	7	0	1	0
_	3rd(n = 8)	5	0	1	1
	Total*	22/27 (81.5%)	1/27 (3.7%)	4/27 (14.8%)	1/27 (3.7%)
	1st (n = 10)	10	0	0	0
25μg	2nd(n = 10)	9	1	1	1
	3rd (n = 9)	6	0	2	1
	Total*	25/29 (86.2%)	1/29 (3.4%)	3/29 (10.3%)	2/29 (6.9%)
	1st (n = 10)	10	0	1	1
50μg	2nd(n = 9)	9	2	0	2
, 3	3rd (n = 9)	9	1	1	1
	Total*	28/28 (100%)	3/28 (10.7%)	2/28 (7.1%)	4/28 (14.3%

Vaccine group	Adverse Event	Nausea	Malaise	Myalgia	Cephalea	Ax. Temp.≥37°0
	1st dose (n = 10)	1	1	0	3	1
12.5μg	2nd dose $(n = 9)$	1	1	0	0	4
	3rd dose (n = 8)	0	1	1	0	2
	Total (%)*	7.4	11.1	3.7	11.1	26.0
	1st dose (n = 10)	2	0	0	4	3
25μg	2nd dose $(n = 10)$	1	1	1	1	3
_	3rd dose (n = 9)	0	1	0	0	4
	Total (%)*	10.3	6.9	3.4	17.2	34.5
	1st dose (n = 10)	1	1	0	3	5
50μg	2nd dose $(n = 9)$	0	2	1	5	2
	3rd dose (n = 9)	1	2	2	1	3
	Total (%)*	7.1	17.9	10.7	32.1	35.7

Table 4 – Adverse events according to intensity and vaccine group.								
		.5 μg = 44	$25 \mu g$ N = 52		50 μg N = 66			
Intensity of adverse events	N	%	N	%	N	%		
Mild Moderate Intense	38 6 0	86.4 13.6 0	43 8 1	82.7 15.4 1.9	46 20 0	69.7 30.3 0		

Table 5 – Number of participants with hematological laboratory results below reference values, according to time

* N = total number of adverse events reported after all 3 doses per group.

of follow-up after administration of meningococcus B candidate vaccine.

Time of follow-up	Red cells	Hemoglobin	Hematocrit
Day 0 (screening)	2	3	6
Day 2 (48 h after 1st dose)	7	10	9
Day 62 (48 h after 2nd dose)	4	8	9
Day 122 (48 h after 3rd dose)	2	7	5

Laboratory test abnormalities were apparent in hematological parameters, with increased number (compared to baseline data) of participants presenting levels below the reference values (Table 5). Those lower values were observed predominantly in women, even though they comprised 1/3 of the study subjects. In fact, among women the geometric mean of hemoglobin decreased 4–8% approximately, after the first dose in all three vaccine groups, whereas in men there was a 3% reduction in groups 12.5 μ g and 50 μ g and 0.7

increase in group 25 μ g. A 15% increase in the geometric mean of leucocyte count was seen after the first dose of the 50 μ g group whereas in the other vaccine groups it was less than 3%. There was also an increase in the percentage and number of neutrophils after all dose groups, but more evident in the 50 μ g group, in which the increase in C-reactive protein was also more marked, consistent with an inflammatory response. One participant had laboratory test result consistent with auto-immunity (cardiolipin IgM of 20) after the third dose in the 50 μ g group. No other clinically relevant laboratory test abnormality was detected.

Immunogenicity

All participants had bacteridal antibody titers equal to or higher than 1:4 before and after vaccination (data not shown).

The three groups that received the experimental vaccines showed protective bactericidal antibody activity in pre-immune sera for both tested strains (Table 6). Conversely, seroconversion based on ELISA units against OMVs from N. meningitidis was more frequent for higher doses of N44/89 strains compared to the N603/95 strain (Table 7).

Table 6 – Geometric mean titers of bactericidal antibodies against N. meningitidis serogroup B strains N44/89 and N603/95 in serum samples, according to vaccine formulation.

			G		% of seroconversion ^b		
Vaccine formulation $(\mu g \text{ of OMV protein } +$		Pre-vac	Pre-vaccination Post-vaccination				ccination
μg dLOS/dose)	N	N44/89	N603/95	N44/89	N603/95	N44/89	N603/95
12.5 + 6.25	8	25.0	19.0	35.0	32.0	25.0	50.0
25 + 12.25	9	7.3	8.3	19.4	15.4	66.7	55.6
50 + 25	9	7.2	8.3	19.4	18.0	100.0	55.6

^a GMT: geometric mean bactericidal titer (reciprocal dilution).

Table 7 – Geometric mean of ELISA units against OMVs from N. meningitidis strains N44/89 and N603/95 in serum samples from vaccines, and proportion of seroconversion, according to vaccine formulation.

			. 1				
Vaccine formulation μ g OMV protein +		Pre-vaccination Post-		Post-va	ccination	% Seroconversion ^b	
μg dLOS/dose)	N	N44/89	N603/95	N44/89	N603/95	N44/89	N603/95
12.5 + 6.25	8	401.4	381.9	968.4	810.9	0	11
25 + 12.25	9	159.6	191.1	713.6	765.9	67	34
50 + 25	9	138.1	176.5	409.1	516.1	56	34

^a GMT: geometric mean of ELISA units for OMV from each strain.

b Bactericidal titer <1/8 before vaccination and ≥1/8 after the 3rd dose or ≥4 fold increase.

b ≥4 fold increase after the 3rd dose.

Table 8 – Geometric mean of ELISA units against LOS from N. meningitidis strain N44/89 in serum samples from vacinated volunteers and proportion of seroconversion, according to vaccine formulation.

Vaccine formulation		GMT ^a (El	h	
(μ g OMV protein + μ g dLOS/dose	N	Pre-vaccination	Post-vaccination	% Seroconversion ^b
12.5 + 6.25	8	78.6	229.1	25
25 + 12.25	9	109.0	174.7	25
50 + 25	8	81.1	118.9	25

^a GMT: geometric mean of ELISA units for LOS from N44/89 strain.

Total IgG antibody against OMVs from both strains presented high levels before vaccination (Table 7). For the intermediate and highest doses a fourfold increase was observed in 67% and 56% of vaccinated volunteers, respectively, for the N44/89 strain. Only the group that received a 12.5 μ g dose did not show any seroconversion. The post-vaccination values for N603/95 strain were 11%, 34% and 34% for 12.5 μ g; 25 μ g and 50 μ g/dose, respectively. Total IgG antibodies against LOS had a fourfold increase in 25% of the three vaccinated groups (Table 8).

Discussion

This was the first clinical trial of a new vaccine against serogroup B meningococcus totally produced in Brazil and based on the concept of a tailor-made vaccine, that is, a vaccine made based on the Brazilian epidemiological conditions. A dose-escalating design was used with the objective of finding the best balance between reactogenicity and immunogenicity.

The reactogenicity profile found in the current study was expected for this kind of vaccine and was similar to those obtained in other studies with higher frequency of adverse events after larger doses. Tappero et al. in 1999^{31} found that the reactogenicity of the Cuban vaccine (50 μ g) was higher than the Norwegian vaccine (25 μ g). Studies comparing the Norwegian vaccine to the New Zealand vaccine, both with 25 μ g OMV, have shown similar frequency of adverse events. ²² A phase I/II study comparing two antigenic concentrations of the New Zealand vaccine, 25 μ g and 50 μ g/ dose, showed that the higher concentration induced more adverse events, without additional benefit in immunogenicity. ^{32,33}

In the current study, adverse events were mild or moderate, and no serious adverse event was observed. The most frequent local and systemic adverse events were local pain and low grade fever, respectively. In line with the studies mentioned above, we observed that adverse events increased with higher antigen concentrations, chiefly local adverse events. Similarly, as antigen concentration increased, there was a trend for sooner occurrence and longer duration of adverse events.

Clinical laboratory tests, in general, remained within normal limits, but reflected an inflammatory response expected for this kind of vaccine.

One caveat is the lack of an unvaccinated control group which would allow to distinguish systemic nonspecific adverse events that could have had other causes unrelated to the candidate vaccine. It is also important to consider that the sample size was not conceived to detect nor to compare less frequent adverse events, which will be addressed in phase II trials.

Immunological response was very good for strain B:4: P1.14.15 (N44/89) and moderate for strain B:4:P1.7.1 (N603/95), although the high level of antibodies before vaccination have impaired analysis of immunogenicity.

We used a very strict criterion for seroconversion, considering seroprotected individuals with bactericidal titers \geq 1:8 or fourfold increase in titers. The most recent recommendations consider seroprotected individuals with \geq 1:4 bactericidal titers or fourfold increase in titers. ^{34–36} However, in the current study, adoption of this last recommendation would lead to the same results.

In summary, the Brazilian vaccine against meningococcus B has shown an acceptable safety profile and promising immunogenicity, which shall be further verified in a phase II study in seronegative children or with low level of antibodies before vaccination.

Conflicts of interest

At the time of the study, all authors, except LABC, were employed at Bio-Manguinhos/Fiocruz, a Brazilian state-owned producer of vaccines.

Acknowledgements

We thank Dr Carl Frash and Dr Margaret Bash for helpful inputs during the design of the study.

We are also grateful to the Data Monitoring Committee: José Cassio de Moraes. Gabriel Wolf Oselka (University of São Paulo) and Francisco José Roma Paumgartten (Fiocruz).

Sponsorship

The study was sponsored by Financiadora de Estudos e Projetos (FINEP). Ministério da Ciência e Tecnologia, Brazil, as part of the Inovacina's grant.

b ≥4 fold increase after the 3rd dose.

REFERENCES

- Fonseca C, Moraes JC, Barata RB. O Livro da Meningite. São Paulo: Segmento Farma; 2004.
- Moraes JC, Barata RB. A doença meningocócica em São Paulo, Brasil, no século XX: características epidemiológicas. Cad Saude Publica. 2005;21:1458–71.
- Sáfadi MAP, Farhat CK. Vacinas meningocócicas. In: Farhat CK, Weckx LY, Carvalho LHFR, Succi RCM, eds. Imunizações, 5th. ed; 2008. p. 387–402.
- 4. Azevedo N. Bio-Manguinhos na origem: um capítulo da história da auto-suficiência tecnológica em saúde no Brasil. In: Azevedo N, Gadelha CAG, Ponte CF, Trindade C, Hamilton W, eds. Rio de Janeiro, Fiocruz: Inovação em Saúde; 2007. p. 82.
- 5. Coordenação-Geral Do Programa Nacional De Imunizações. Doença Meningocócica. Bol Epidemiol. [Internet]set [1 de outubro de 2019];21-22(Número especial: Vigilância em Saúde no Brasil 2003|2009: da criação da Secretaria de Vigilância em Saúde aos dias atuais). Disponível em http://www.saude.gov.br/boletins-epidemiologicos.
- de Filippis I, Lemos APS, Hostetler JB, et al. Molecular Epidemiology of Neisseria meningitides Serogroup B in Brazil. PLoS ONE. 2012;7:e33016 www.plosone.org.
- 7. SINAN, http://portalsinan.saude.gov.br/meningite 2021
- Sacchi CT, Zanella RC, Caugant DA, et al. Emergence of a new clone of serogroup C Neisseria meningitides in São Paulo, Brazil. J Clin Microbiol. 1992;30:1282–6.
- Milagres LG, Ramos SR, Sacchi CT. Immune response of Brazilian children to a Neisseria meningitides serogroup B outer membrane protein vaccine: comparison with Efficacy 1994. Intection Immunity. 1994;62:4419–24.
- Halperina SA, Bettinger JA, Greenwoodc B, et al. The changing and dynamic epidemiology of meningococcal disease. Vaccine. 2012;30S:B26–36.
- 11. Sáfadi MAP, Cintra OAL. Epidemiology of meningococcal disease in Latin America: current situation and opportunities for prevention. Neurol Res. 2010;32:265–71.
- Borrow R, Alarcón P, Carlos J, et al. The Global Meningococcal Initiative: global epidemiology, the impact of vaccines on meningococcal disease and the importance of herd protection. Expert Review Vaccines. 2017;16:313–28.
- Michael W. Shea the long road to an effective vaccine for meningococcus group B (MenB). Annal Med Surg. 2013;2:53–6.
- Gorringe AR, Pajon R. Bexsero: a multicomponent vaccine for prevention of meningococcal disease. Hum Vac Immunoth. 2012;8:174–83.
- Azzari C, Bonanni P. A new meningococcal B vaccine for adolescents and adults: characteristics and methods of use. J. Prev. Med. Hyg. 2018;59:E257–60.
- 16. Boutriau D, Poolman J, Borrow R, et al. Immunogenicity and safety of three doses of a bivalent (B:4:P1.19,15 and B:4:P1.7-2,4) meningococcal outer membrane vesicle vaccine in healthy adolescents. Clin Vaccine Immunol. 2007;14:65–73.
- Noronha CP, Struchiner CJ, Halloran ME. Assessment of the direct effectiveness of BC meningococcal vaccine in Rio de Janeiro, Brazil: a case-control study. Int J Epidemiol. 1995;24:1050–7.
- Kelly C, Arnold R, Galloway Y, O'Hallahan J. A prospective study of the effectiveness of the New Zealand meningococcal B vaccine. Am J Epidemiol. 2007;166:817–23.
- Jackson C, Lennon DR, Sotutu VTK. Phase II meningococcal B vesicle vaccine trial in New Zealand infants. Arch Dis Child. 2009;94:745–51.
- 20. Jessouroun E, da Silveira IF, Larangeira AP, et al. Outer membrane vesicles (OMVs) and detoxified lipooligosaccharide

- (dLOS) obtained from Brazilian prevalent N. meningitidis serogroup B strains protect mice against homologous and heterologous meningococcal infection and septic shock. Vaccine. 2004;22:2617–25.
- 21. Pizza MV, Scarlato V, Masignani V, et al. Identification of vaccine candidates against serogroup B meningococcus by whole-genome sequencing. Science. 2000;287:1816–20.
- 22. Fu J, Bailey FJ, King JJ, et al. Recent advances in the large scale fermentation of Neisseria meningitidis group B for the production of an outer membrane protein complex. Biotechnology. 1995;13:170–4.
- 23. Gotschlich EC, Liu TY, Artenstein MS. Human immunity to the meningococcus. 3. Preparation and immunochemical properties of the group A, group B, and group C meningococcal polysaccharides. J Exp Med. 1969;129:1349–65.
- 24. Borrow R, Aaberge IS, Santos GF, et al. Interlaboratory standardization of the measurement of serum bactericidal activity by using human complement against meningococcal serogroup b, strain 44/76-SL, before and after vaccination with the Norwegian MenBvac outer membrane vesicle vaccine. Clin Diagn Lab Immunol. 2005;12:970–6.
- Borrow R, Carlone GM, Rosenstein N, et al. Neisseria meningitidis group B correlates of protection and assay standardization—international meeting report Emory University, Atlanta, Georgia, United States, 16-17 March 2005. Vaccine. 2006;24:5093–107.
- Rosenqvist E, Kayhty H, Pollard AJ. Determination of antibody responses to meningococcal antigens by ELISA. Methods Mol Med. 2001;66:255–73.
- Estabrook MM, Jarvis GA, McLeod Griffiss J. Affinity-purified human immunoglobulin G that binds a lacto-N-neotetraosedependent lipooligosaccharide structure is bactericidal for serogroup B Neisseria meningitidis. Infect Immun. 2007;75:1025– 33.
- 28. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. J Bull World Health Organ. 2001;79:373–4.
- **29.** European Clinical Trials Directive. 2001/20/EG of the European Parliament and Concil. 34.
- Brasil. Ministério da Saúde. Conselho Nacional de Saúde. RESOLUÇÃO N° 196, DE 10 DE OUTUBRO DE. Available from http://bvsms.saude.gov.br/bvs/saudelegis/cns/1996/ res0196_10_10_1996.html.
- Tappero JW, Lagos R, Ballesteros AM, et al. Immunogenicity of 2 serogroup outer-membrane protein meningococcal vaccines: a randomized controlled trial in Chile. JAMA. 1999;281:1541–3.
- 32. Nokleby H, Aavitsland P, O'Hallahan J, Feiring B, Tilman S, Oster P. Safety review: two outer membrane vesicle (OMV) vaccines against systemic Neisseria meningitidis serogroup B disease. Vaccine. 2007;25:3080–4.
- 33. Thornton V, Lennon D, Rasanathan K, et al. Safety and immunogenicity of New Zealand strain meningococcal serogroup B, OMV vaccine in healthy adults: Beginning of epidemic control. Vaccine. 2006;24:1395–400.
- 34. Meeting report 2006. Neisseria Meningitidis Group B Correlates of Protection and Assay Standardization International Meeting Report Emory University, 24; 2006. Atlanta, Georgia, United States, 16-17 March 2005. Vaccinep. 5093–107.
- 35. Findlow J, Taylor S, Aase A, et al. Comparison and correlation of Neisseria meningitidis serogroup B immunologic assay results and human antibody responses following three doses of the Norwegian meningococcal outer membrane vesicle vaccine MenBVac. Infect Immun. 2006;74:4557–65.
- Borrow R, Balmer P, Miller E. Meningococcal surrogates of protection - serum bactericidal antibody activity. Vaccine. 2005;23:2222–7.