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## Original Article

# **In vitro activity of ceftazidime-avibactam against Enterobacterales and *Pseudomonas aeruginosa* isolates collected in Latin America as part of the ATLAS global surveillance program, 2017–2019**

James A. Karlowsky<sup>a</sup>, Krystyna M. Kazmierczak<sup>b,\*</sup>,  
Maria Lavínea Novis de Figueiredo Valente<sup>c</sup>, Elkin Lemos Luengas<sup>d</sup>,  
Monique Baudrit<sup>e</sup>, Alvaro Quintana<sup>f</sup>, Paurus Irani<sup>g</sup>, Gregory G. Stone<sup>h</sup>,  
Daniel F. Sahn<sup>b</sup>

<sup>a</sup> University of Manitoba, Max Rady College of Medicine, Department of Medical Microbiology and Infectious Diseases, Winnipeg, Canada

<sup>b</sup> IHMA, Schaumburg, IL, USA

<sup>c</sup> Pfizer Brazil, São Paulo, SP, Brazil

<sup>d</sup> Pfizer Colombia, Bogota, Colombia

<sup>e</sup> Pfizer Central America and Caribbean, Cartago, Costa Rica

<sup>f</sup> Pfizer Inc., New York, NY, USA

<sup>g</sup> Pfizer UK Ltd, Walton Oaks, Tadworth, Surrey, UK

<sup>h</sup> Pfizer Inc., Groton, CT, USA

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## ABSTRACT

The Antimicrobial Testing Leadership and Surveillance (ATLAS) global surveillance program collected clinical isolates of Enterobacterales ( $n = 8416$ ) and *Pseudomonas aeruginosa* ( $n = 2521$ ) from 41 medical centers in 10 Latin American countries from 2017 to 2019. *In vitro* activities of ceftazidime-avibactam and comparators were determined using the Clinical and Laboratory Standards Institute (CLSI) broth microdilution method. Overall, 98.1% of Enterobacterales and 86.9% of *P. aeruginosa* isolates were susceptible to ceftazidime-avibactam. When isolates were analyzed by country of origin, susceptibility to ceftazidime-avibactam for Enterobacterales ranged from 97.8% to 100% for nine of 10 countries (except Guatemala, 86.3% susceptible) and from 75.9% to 98.4% for *P. aeruginosa* in all 10 countries. For Enterobacterales, 100% of AmpC-positive, ESBL- and AmpC-positive, GES-type carbapenemase-positive, and OXA-48-like-positive isolates were ceftazidime-avibactam-susceptible as were 99.8%, 91.8%, and 74.7% of ESBL-positive, multidrug-resistant (MDR), and meropenem-nonsusceptible isolates. Among meropenem-nonsusceptible isolates of Enterobacterales, 24.4% (139/570) carried a metallo- $\beta$ -lactamase (MBL); 83.3% of the remaining meropenem-nonsusceptible isolates carried another class of carbapenemase and 99.4% of

\* Corresponding author.

E-mail address: [kkazmierczak@ihmainc.com](mailto:kkazmierczak@ihmainc.com)

(K.M. Kazmierczak).

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those isolates were ceftazidime-avibactam-susceptible. Among meropenem-non-susceptible isolates of *P. aeruginosa* ( $n = 835$ ), 25.6% carried MBLs; no acquired  $\beta$ -lactamase was identified in the majority of isolates (64.8%; 87.2% of those isolates were ceftazidime-avibactam-susceptible). Overall, clinical isolates of Enterobacterales collected in Latin America from 2017 to 2019 were highly susceptible to ceftazidime-avibactam, including isolates carrying ESBLs, AmpCs, and KPCs. Country-specific variation in susceptibility to ceftazidime-avibactam was more common among isolates of *P. aeruginosa* than Enterobacterales. The frequency of MBL-producers among Enterobacterales from Latin America was low (1.7% of all isolates; 146/8,416), but higher than reported in previous surveillance studies.

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## Introduction

Ceftazidime-avibactam combines ceftazidime, an established third-generation cephalosporin, with avibactam, a diazabicyclooctanone non- $\beta$ -lactam  $\beta$ -lactamase inhibitor, and is indicated in the treatment of patients with complicated urinary tract infections, including pyelonephritis, complicated intra-abdominal infections, and hospital-acquired and ventilator-associated bacterial pneumonia caused by gram-negative bacilli, including species of Enterobacterales and *Pseudomonas aeruginosa*.<sup>1</sup> *In vitro*, avibactam has consistently demonstrated the ability to restore the potency of ceftazidime against isolates of Gram-negative bacilli carrying Ambler class A  $\beta$ -lactamases (including extended spectrum  $\beta$ -lactamases [ESBLs] and KPC carbapenemases), class C  $\beta$ -lactamases (AmpC cephalosporinases), and some class D  $\beta$ -lactamases (OXA-48-like carbapenemases), including isolates harboring ESBL and AmpC enzymes in combination with impaired permeability due to porin mutation or loss.<sup>2–6</sup> Ceftazidime-avibactam is not active against isolates of Enterobacterales and *P. aeruginosa* carrying class B metallo- $\beta$ -lactamases (MBLs) (e.g., NDM, IMP, VIM) and rare isolates that possess specific protein sequence mutations in target enzymes (AmpC, KPC, PBP3) or overexpress certain efflux pumps due to mutation.<sup>2–4,5–18</sup>

Carbapenemase-producing gram-negative bacilli are of increasing global concern as carbapenems are recommended in the treatment of patients with severe infections due to ESBL- and AmpC-producing Gram-negative bacilli. Carbapenemase-producing Gram-negative bacilli frequently demonstrate a multidrug-resistant (MDR) phenotype leaving few therapeutic options and are associated with elevated patient morbidity and mortality.<sup>2,3</sup> Ceftazidime-avibactam may be an appropriate agent for treatment of some patients with infections caused by Enterobacterales or *P. aeruginosa* when treatment options are limited.

The intent of the current study was to provide data on the *in vitro* activities of ceftazidime-avibactam and comparators tested against clinical isolates of Enterobacterales and *P. aeruginosa* collected from hospitalized patients in 10 Latin American countries over a recent three-year time period (2017–2019). In addition, we sought to analyze the activity of ceftazidime-avibactam against carbapenem-nonsusceptible, MDR, and molecularly characterized  $\beta$ -lactamase-producing isolate

subsets. These data were collected as part of the Antimicrobial Testing Leadership and Surveillance (ATLAS) global surveillance program, which succeeded the International Network for Optimal Resistance Monitoring (INFORM) global surveillance program and continues its framework and mission. The INFORM program was established in 2012 to benchmark and then monitor the *in vitro* activity of ceftazidime-avibactam and comparative agents against clinical isolates of  $\beta$ -lactamase-producing Enterobacterales and non-fermentative Gram-negative bacilli, including *P. aeruginosa*. The current study extends earlier publications on isolates collected in Latin America from 2012 to 2017 under the INFORM program.<sup>2,3</sup> Only a limited number of other, now outdated, surveillance studies to determine rates of antimicrobial resistance in clinical isolates from patients in Latin American countries have been published.<sup>19–23</sup> To date, the majority of other published studies have not included  $\beta$ -lactamase characterization of resistant isolates from the Latin America region and generally have not provided country-specific ceftazidime-avibactam susceptibility data for Gram-negative bacilli isolated from patients in as many Latin America countries as the current study of ATLAS surveillance program data.<sup>2,3,19–23</sup>

## Materials and methods

### Clinical isolates of Enterobacterales and *P. aeruginosa*

The ATLAS global surveillance program collected 10937 non-duplicate clinical isolates of Gram-negative bacilli (8416 isolates of Enterobacterales and 2521 isolates of *P. aeruginosa*) from 41 medical center laboratories in 10 countries in Latin America from 2017 to 2019. The ATLAS program annually requests that each participating medical center laboratory collect pre-defined quotas of selected bacterial pathogens isolated from patients with specific types of infection.<sup>2,3</sup> Collection was limited to one isolate per patient. All isolates were determined to be clinically significant by participating laboratory algorithms and were collected irrespective of antimicrobial susceptibility profile.<sup>2,3</sup> The demographic information associated with the 10937 isolates is summarized in [supplementary Table S1](#). All isolates were transported to IHMA (Schaumburg, IL, USA) which served as the central

testing laboratory for the ATLAS program. IHMA confirmed the identity of each isolate using MALDI-TOF mass spectrometry (Bruker Daltonics, Billerica, MA, USA) prior to antimicrobial susceptibility testing.

### Antimicrobial susceptibility testing

Antimicrobial susceptibility testing was performed following the Clinical and Laboratory Standards Institute (CLSI) standard method using custom 96-well broth microdilution panels prepared by Trek (Thermo Fisher Scientific, Oakwood Village, OH, USA; 2017) or IHMA (2018–2019).<sup>24,25</sup> Ceftazidime-avibactam was tested using a fixed concentration of 4  $\mu\text{g}/\text{ml}$  of avibactam.<sup>24</sup> MICs were interpreted using 2020 CLSI breakpoints<sup>24</sup> with the exception of tigecycline which used current US FDA MIC interpretative breakpoints.<sup>26</sup>

An MDR phenotype was defined as resistance to sentinel agents from three or more antimicrobial agent classes, including cephalosporins (cefepime), monobactams (aztreonam),  $\beta$ -lactam- $\beta$ -lactamase inhibitor combinations (piperacillin-tazobactam), carbapenems (meropenem), fluoroquinolones (levofloxacin), aminoglycosides (amikacin), and polymyxins (colistin).

### Screening of clinical isolates of Enterobacterales and *P. aeruginosa* for $\beta$ -lactamase genes

All isolates of Enterobacterales testing with MICs to meropenem of  $\geq 2 \mu\text{g}/\text{ml}$  and all *E. coli*, *K. pneumoniae*, *K. oxytoca* and *P. mirabilis* with MICs to ceftazidime or aztreonam of  $\geq 2 \mu\text{g}/\text{ml}$  were screened for  $\beta$ -lactamase content using published multiplex PCR assays.<sup>27</sup> These assays detected genes encoding carbapenemases (KPC, GES, NDM, IMP, VIM, SPM, GIM, OXA-48-like), ESBLs (TEM, SHV, CTX-M, VEB, PER, GES), original (narrow)-spectrum  $\beta$ -lactamases (TEM and SHV enzymes lacking substitutions at amino acid positions 104, 164, or 238 in TEM or 146, 238, or 240 in SHV that are associated with ESBL activity),<sup>28</sup> and plasmid-mediated AmpC  $\beta$ -lactamases (ACC, ACT, CMY, DHA, FOX, MIR, MOX) as previously described.<sup>27</sup> All isolates of *P. aeruginosa* testing with MICs to meropenem of  $\geq 4 \mu\text{g}/\text{ml}$  were screened for genes encoding the  $\beta$ -lactamases listed above and OXA-24/40-like carbapenemases, as described previously.<sup>3</sup> Enzyme variants were identified by amplification of full-length  $\beta$ -lactamase genes followed by DNA sequencing and comparison of the sequences generated to the National Center for Biotechnology Information database ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)).

## Results

Of the 8416 isolates of Enterobacterales tested, 98.1% were susceptible to ceftazidime-avibactam (MIC<sub>90</sub>, 0.5  $\mu\text{g}/\text{ml}$ ) (Table 1). Percent susceptible values for all other agents tested were comparable (tigecycline, amikacin) or lower than for ceftazidime-avibactam. Among the 6.8% of isolates that tested as meropenem-nonsusceptible, which included 139 MBL-positive isolates, 74.7% were susceptible to ceftazidime-avibactam. When MBL-positive isolates were removed from the subset of meropenem-nonsusceptible isolates, susceptibility

to ceftazidime-avibactam increased to 99.4% among isolates that were serine carbapenemase-positive and 95.8% among isolates that were meropenem-nonsusceptible and carbapenemase-negative. Of the subset of isolates identified with an MDR phenotype, which included MBL-positive isolates, 91.8% were susceptible to ceftazidime-avibactam.

Percent susceptible values for ceftazidime-avibactam for all Enterobacterales isolates collected from nine of the 10 countries surveyed ranged from 97.8% (Venezuela) to 100% (Chile, Dominican Republic) (Table 2). Enterobacterales from Guatemala were exceptional in that only 86.3% of isolates were susceptible to ceftazidime-avibactam. The percentages of isolates that tested as non-susceptible to meropenem differed by almost 15% among the 10 Latin American countries surveyed, ranging from 0% in Dominican Republic and 0.9% in Panama to 14.8% in Guatemala. Among meropenem-nonsusceptible isolates, susceptibility to ceftazidime-avibactam ranged from 100% (Chile) to 0% (Costa Rica,  $n = 3$ ) and was  $>87\%$  for four countries,  $\sim 50\text{--}67\%$  for three countries, and  $<10\%$  for two countries. When MBL-positive isolates were removed from the dataset, susceptibility to ceftazidime-avibactam increased to  $\geq 98.8\%$ , exceeding susceptibility to all other agents tested, for meropenem-nonsusceptible isolates collected in all Latin American countries except Guatemala (66.7% susceptible,  $n = 6$ ) and Mexico (92.9% susceptible among meropenem-nonsusceptible, carbapenemase-negative isolates). Country-specific MDR rates among Enterobacterales isolates ranged from 8.1% (Costa Rica) to 26.9% (Guatemala), with  $\geq 93.5\%$  of MDR isolates collected in seven countries testing as susceptible to ceftazidime-avibactam; susceptibilities of MDR isolates were lower in Venezuela (88.0%), Costa Rica (80.0%), and Guatemala (49.0%).

One or more  $\beta$ -lactamase genes were detected in 2512 of 2578 Enterobacterales isolates screened for  $\beta$ -lactamase content (Methods). A total of 2254 ESBLs were detected singly or in combination with other  $\beta$ -lactamases in 2161 isolates, with CTX-M-15 identified in  $>50\%$  of ESBL-positive isolates in each Latin American country and in 71.5% (1545/2161) of ESBL-positive isolates overall (Fig. 1). Similarly, 519 genes encoding carbapenemases were identified in 497 of 569 meropenem-nonsusceptible Enterobacterales isolates (one meropenem-nonsusceptible isolate collected in Guatemala was not molecularly characterized for  $\beta$ -lactamase genes). KPC was the most common carbapenemase identified in meropenem-nonsusceptible molecularly characterized isolates overall, accounting for 61.5% (350/569) of isolates, and isolates carrying KPC as the sole carbapenemase composed the majority in Argentina (89.7%), Brazil (81.7%), Colombia (77.6%), and Panama (66.7%) (Fig. 2). NDM (an MBL) was the second most common carbapenemase identified in the region and was found alone or with serine carbapenemases in 100%, 86.8%, 48.5%, and 47.5% of meropenem-nonsusceptible isolates collected in Costa Rica ( $n = 3$ ), Guatemala, Venezuela, and Mexico, respectively.

Table 3 depicts the *in vitro* activity of ceftazidime-avibactam and comparator agents against isolates of Enterobacterales molecularly characterized for  $\beta$ -lactamase gene content. Ceftazidime-avibactam inhibited 100% of AmpC-positive, ESBL- and AmpC-positive, GES-type carbapenemase-positive, and OXA-48-like-positive isolates of Enterobacterales as well

**Table 1 – In vitro activity of ceftazidime-avibactam and comparator agents against Enterobacterales isolates collected in the Latin American region as part of the ATLAS global surveillance program from 2017 to 2019.**

Organism, phenotype/genotype (no. of isolates)	Antimicrobial agent	MIC ( $\mu\text{g/ml}$ )		Interpretation (CLSI)		
		MIC <sub>50</sub>	MIC <sub>90</sub>	% Susceptible	% Intermediate	% Resistant
Latin America, All Enterobacterales (8416) <sup>a</sup>	Ceftazidime-avibactam	0.12	0.5	98.1	NA	1.9
	Ceftazidime	0.25	64	67.8	3.6	28.5
	Cefepime	$\leq 0.12$	>16	68.5	6.7	24.7
	Piperacillin-tazobactam	2	>64	82.7	5.4	11.9
	Meropenem	$\leq 0.06$	0.12	93.2	0.8	6.0
	Levofloxacin (n = 8415) <sup>d</sup>	$\leq 0.25$	>8	61.0	5.6	33.4
	Amikacin	2	8	96.2	1.5	2.3
	Colistin	0.5	>8	NA	81.1	18.9
	Tigecycline	0.5	1	97.2	2.4	0.4
	Meropenem-NS (570) <sup>b</sup>	Ceftazidime-avibactam	1	>128	74.7	NA
Ceftazidime		128	>128	5.4	3.9	90.7
Cefepime		>16	>16	3.7	7.7	88.6
Piperacillin-tazobactam		>64	>64	1.9	5.6	92.5
Meropenem		>8	>8	0	11.6	88.4
Levofloxacin		>8	>8	16.0	8.4	75.6
Amikacin		8	>32	70.2	10.4	19.5
Colistin		0.5	>8	NA	78.2	21.8
Tigecycline		0.5	2	95.6	4.0	0.4
Meropenem-NS, MBL-, carbapenemase+ (358)		Ceftazidime-avibactam	1	2	99.4	NA
	Ceftazidime	64	>128	5.3	5.6	89.1
	Cefepime	>16	>16	4.5	8.4	87.2
	Piperacillin-tazobactam	>64	>64	0	3.4	96.6
	Meropenem	>8	>8	0	7.0	93.0
	Levofloxacin	>8	>8	14.2	5.0	80.7
	Amikacin	4	>32	76.5	13.1	10.3
	Colistin	0.5	>8	NA	74.9	25.1
	Tigecycline	0.5	2	98.0	1.7	0.3
	Meropenem-NS, MBL-, carbapenemase- (72)	Ceftazidime-avibactam	1	4	95.8	NA
Ceftazidime		64	>128	16.7	2.8	80.6
Cefepime		>16	>16	6.9	8.3	84.7
Piperacillin-tazobactam		>64	>64	11.1	15.3	73.6
Meropenem		2	>8	0	52.8	47.2
Levofloxacin		>8	>8	19.4	9.7	70.8
Amikacin		4	>32	81.9	2.8	15.3
Colistin		0.5	>8	NA	87.5	12.5
Tigecycline		0.5	2	91.7	8.3	0
Multidrug-resistant (1773) <sup>c</sup>		Ceftazidime-avibactam	0.5	4	91.8	NA
	Ceftazidime	64	>128	5.4	4.7	89.9
	Cefepime	>16	>16	1.6	3.6	94.8
	Piperacillin-tazobactam	32	>64	44.3	11.1	44.6
	Meropenem	$\leq 0.06$	>8	69.4	2.6	28.0
	Levofloxacin (n = 1772) <sup>d</sup>	>8	>8	7.1	4.3	88.5
	Amikacin	4	>32	84.2	5.5	10.3
	Colistin	0.5	>8	NA	85.4	14.6
	Tigecycline	0.5	2	95.8	3.5	0.7

Abbreviations: NS, non-susceptible; MBL-, no gene encoding a metallo- $\beta$ -lactamase was detected by PCR; carbapenemase+/-, a gene encoding a serine carbapenemase was (+) or was not (-) detected by PCR; multidrug-resistant, isolates resistant to three or more sentinel agents from different antimicrobial classes; NA, no breakpoint available.

<sup>a</sup> All Enterobacterales were composed of *Citrobacter amalonaticus* (n = 10), *Citrobacter braakii* (n = 20), *Citrobacter farmeri* (n = 7), *Citrobacter freundii* (n = 251), *Citrobacter koseri* (n = 119), *Citrobacter sedlakii* (n = 6), *Citrobacter youngae* (n = 1), *Citrobacter* sp. (n = 4), *Enterobacter asburiae* (n = 87), *Enterobacter bugandensis* (n = 8), *Enterobacter cloacae* (n = 703), *Enterobacter cloacae* complex (n = 25), *Enterobacter kobei* (n = 27), *Enterobacter ludwigii* (n = 4), *Enterobacter xiangfangensis* (n = 14), *Enterobacter* sp. (n = 32), *Escherichia coli* (n = 2747), *Klebsiella aerogenes* (n = 221), *Klebsiella oxytoca* (n = 213), *Klebsiella pneumoniae* (n = 2441), *Klebsiella variicola* (n = 98), *Morganella morganii* (n = 263), *Pantoea agglomerans* (n = 1), *Pantoea septica* (n = 1), *Proteus hauseri* (n = 48), *Proteus mirabilis* (n = 436), *Proteus penneri* (n = 8), *Proteus vulgaris* (n = 58), *Proteus* sp. (n = 2), *Providencia alcalifaciens* (n = 4), *Providencia rettgeri* (n = 51), *Providencia stuartii* (n = 83), *Providencia* sp. (n = 2), *Raoultella ornithinolytica* (n = 17), *Raoultella planticola* (n = 2), *Raoultella terrigena* (n = 1), *Salmonella* sp. (n = 1), *Serratia liquefaciens* (n = 3), *Serratia marcescens* (n = 392), *Serratia rubidaea* (n = 1), *Serratia ureilytica* (n = 2), and *Serratia* sp. (n = 2).

<sup>b</sup> One meropenem-nonsusceptible isolate collected in Guatemala was not molecularly characterized for  $\beta$ -lactamase genes.

<sup>c</sup> Multidrug resistant Enterobacterales were composed of *Citrobacter freundii* (n = 15), *Citrobacter koseri* (n = 2), *Enterobacter asburiae* (n = 11), *Enterobacter cloacae* (n = 123), *Enterobacter cloacae* complex (n = 2), *Enterobacter xiangfangensis* (n = 1), *Enterobacter* sp. (n = 2), *Escherichia coli* (n = 605), *Klebsiella aerogenes* (n = 12), *Klebsiella oxytoca* (n = 14), *Klebsiella pneumoniae* (n = 845), *Klebsiella variicola* (n = 5), *Morganella morganii* (n = 18), *Proteus mirabilis* (n = 39), *Providencia rettgeri* (n = 7), *Providencia stuartii* (n = 17), *Raoultella ornithinolytica* (n = 1), *Salmonella* sp. (n = 1), and *Serratia marcescens* (n = 53).

<sup>d</sup> Excluded one isolate of *Salmonella* sp.

**Table 2 – Percentages of Enterobacterales isolates collected in ten Latin American countries as part of the ATLAS global surveillance program from 2017 to 2019 that were susceptible to ceftazidime-avibactam and comparator agents.**

Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)					
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-	Multidrug-resistant
Argentina		(1039)	(87)	(81) <sup>a</sup>			(212)
	Ceftazidime-avibactam	98.8	92.0	98.8			96.7
	Ceftazidime	71.7	2.3	2.5			6.6
	Cefepime	73.1	2.3	2.5			1.4
	Piperacillin-tazobactam	79.1	1.1	1.2			23.1
	Meropenem	91.6	0	0			59.9
	Levofloxacin	61.0	9.2	9.9			7.1
	Amikacin	94.8	65.5	69.1			76.9
	Colistin	NA	NA	NA			NA
Tigecycline	96.8	97.7	98.8			96.2	
Brazil		(1646)	(202)		(164)	(14)	(400)
	Ceftazidime-avibactam	98.4	87.6		99.4	100	93.5
	Ceftazidime	71.7	5.0		4.9	14.3	9.0
	Cefepime	69.3	1.5		1.8	0	2.0
	Piperacillin-tazobactam	78.0	0.5		0	7.1	24.3
	Meropenem	87.7	0		0	0	50.2
	Levofloxacin	61.8	10.9		9.8	14.3	6.8
	Amikacin	96.5	79.7		80.5	78.6	86.5
	Colistin	NA	NA		NA	NA	NA
Tigecycline	97.5	97.0		97.6	92.9	95.5	
Chile		(805)	(22)	(22) <sup>b</sup>			(157)
	Ceftazidime-avibactam	100	100	100			100
	Ceftazidime	67.2	0	0			10.2
	Cefepime	69.3	0	0			1.9
	Piperacillin-tazobactam	84.5	4.5	4.5			52.9
	Meropenem	97.3	0	0			86.0
	Levofloxacin	67.2	0	0			3.2
	Amikacin	96.8	95.5	95.5			86.0
	Colistin	NA	NA	NA			NA
Tigecycline	97.4	90.9	90.9			95.5	
Colombia		(1252)	(107)		(83)	(12)	(209)
	Ceftazidime-avibactam	98.9	89.7		100	100	94.7
	Ceftazidime	72.1	13.1		12.0	33.3	5.3
	Cefepime	71.7	10.3		10.8	16.7	2.9
	Piperacillin-tazobactam	80.8	0.9		0	8.3	34.0
	Meropenem	91.5	0		0	0	55.0
Levofloxacin	67.3	31.8		27.7	58.3	17.2	

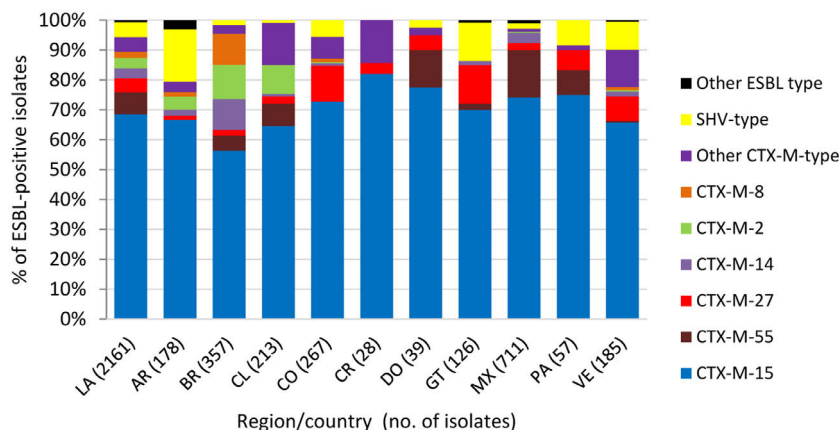
Table 2 (continued)							
Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)					
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-	Multidrug-resistant
Costa Rica	Amikacin	96.2	71.0		71.1	83.3	81.8
	Colistin	NA	NA		NA	NA	NA
	Tigecycline	97.0	95.3		97.6	75.0	95.2
		(185)	(3) <sup>c</sup>				(15)
	Ceftazidime-avibactam	98.4	0				80.0
	Ceftazidime	75.7	0				6.7
	Cefepime	85.4	0				13.3
	Piperacillin-tazobactam	88.6	0				53.3
	Meropenem	98.4	0				80.0
	Levofloxacin	83.2	0				6.7
Dominican Republic	Amikacin	98.9	33.3				86.7
	Colistin	NA	NA				NA
	Tigecycline	99.5	100				100
		(193) <sup>d</sup>					(28)
	Ceftazidime-avibactam	100					100
	Ceftazidime	75.1					14.3
	Cefepime	75.6					0.0
	Piperacillin-tazobactam	95.3					78.6
	Meropenem	100					100
	Levofloxacin	51.3					0.0
Guatemala	Amikacin	96.4					85.7
	Colistin	NA					NA
	Tigecycline	97.9					96.4
		(364)	(54) <sup>e</sup>	(6) <sup>f</sup>			(98)
	Ceftazidime-avibactam	86.3	7.4	66.7			49.0
	Ceftazidime	58.2	1.9	16.7			2.0
	Cefepime	58.2	5.6	50.0			1.0
	Piperacillin-tazobactam	73.4	1.9	0.0			33.7
	Meropenem	85.2	0.0	0.0			49.0
	Levofloxacin	54.9	22.2	50.0			10.2
Mexico	Amikacin	90.7	46.3	66.7			69.4
	Colistin	NA	NA	NA			NA
	Tigecycline	99.2	98.1	100			99.0
		(1802)	(59)		(16)	(14)	(460)
	Ceftazidime-avibactam	98.2	49.2		100	92.9	93.7
	Ceftazidime	55.3	6.8		6.3	21.4	1.3
	Cefepime	57.8	3.4		6.3	7.1	0.7

**Table 2 (continued)**

Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)					Multidrug-resistant
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-	
	Piperacillin-tazobactam	86.2	10.2		0	28.6	67.8
	Meropenem	96.7	0		0	0	88.0
	Levofloxacin	55.3	18.6		18.8	14.3	3.9
	Amikacin	96.7	67.8		100	85.7	88.9
	Colistin	NA	NA		NA	NA	NA
	Tigecycline	96.6	86.4		100	100	95.0
	Panama	(316)	(3)	(2) <sup>g</sup>			(44)
	Ceftazidime-avibactam	99.7	66.7	100			97.7
	Ceftazidime	76.6	0	0			0
	Cefepime	79.7	0	0			2.3
Piperacillin-tazobactam	91.5	0	0			63.6	
Meropenem	99.1	0	0			93.2	
Levofloxacin	55.4	33.3	50.0			4.5	
Amikacin	99.7	100	100			100	
Colistin	NA	NA	NA			NA	
Tigecycline	96.2	100	100			93.2	
Venezuela	(814)	(33)	(16) <sup>h</sup>			(150)	
Ceftazidime-avibactam	97.8	48.5	100			88.0	
Ceftazidime	74.4	0	0			4.0	
Cefepime	74.2	0	0			0.7	
Piperacillin-tazobactam	87.1	0	0			55.3	
Meropenem	95.9	0	0			78.7	
Levofloxacin (n = 813) <sup>i</sup>	58.8	9.1	0			8.1	
Amikacin	95.9	48.5	62.5			80.0	
Colistin	NA	NA	NA			NA	
Tigecycline	97.2	97.0	100			98.0	

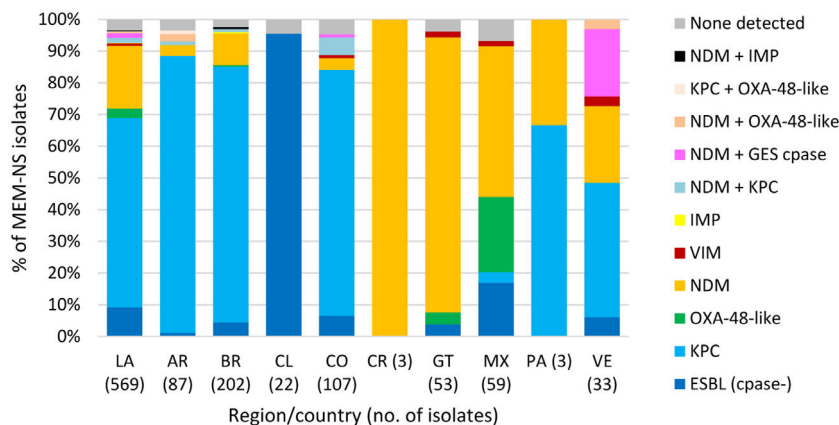
Abbreviations: NS, non-susceptible; MBL-, no gene encoding a metallo-β-lactamase was detected by PCR; carbapenemase+/-, a gene encoding a serine carbapenemase was (+) or was not (-) detected by PCR; multidrug-resistant, isolates resistant to three or more sentinel agents from different drug classes; NA, no breakpoint available.

<sup>a</sup> Composed of 77 carbapenemase-positive isolates and 4 carbapenemase-negative isolates.  
<sup>b</sup> Composed of 22 carbapenemase-negative isolates.  
<sup>c</sup> Composed of 3 MBL-positive isolates; no meropenem-NS, MBL-negative isolates were identified during the surveyed time period.  
<sup>d</sup> No meropenem-NS isolates were identified during the surveyed time period.  
<sup>e</sup> One meropenem-nonsusceptible isolate collected in Guatemala was not molecularly characterized for β-lactamase genes.  
<sup>f</sup> Composed of 2 carbapenemase-positive isolates and 4 carbapenemase-negative isolates.  
<sup>g</sup> Composed of 2 carbapenemase-positive isolates.  
<sup>h</sup> Composed of 14 carbapenemase-positive and 2 carbapenemase-negative isolates.  
<sup>i</sup> Excludes one isolate of *Salmonella* sp.



**Fig. 1 – Extended-spectrum  $\beta$ -lactamases identified in Enterobacterales isolates collected in ten Latin American countries as part of the ATLAS global surveillance program from 2017 to 2019**

LA, Latin America; AR, Argentina; BR, Brazil; CL, Chile; CO, Colombia; CR, Costa Rica; DO, Dominican Republic; GT, Guatemala; MX, Mexico; PA, Panama; VE, Venezuela; ESBL-positive, one or more genes encoding an extended-spectrum  $\beta$ -lactamase was detected by PCR. ESBL-positive isolates included meropenem-susceptible and meropenem-nonsusceptible isolates, some of which carried multiple ESBLs or additional  $\beta$ -lactamases (e.g. original (narrow)-spectrum  $\beta$ -lactamases, AmpC  $\beta$ -lactamases, serine carbapenemases, and/or metallo- $\beta$ -lactamases). Isolates carrying multiple ESBLs were counted for each individual ESBL type.



**Fig. 2 –  $\beta$ -lactamases identified in meropenem-nonsusceptible Enterobacterales isolates collected in ten Latin American countries as part of the ATLAS global surveillance program from 2017 to 2019**

LA, Latin America; AR, Argentina; BR, Brazil; CL, Chile; CO, Colombia; CR, Costa Rica; GT, Guatemala; MX, Mexico; PA, Panama; VE, Venezuela; Cpase, carbapenemase; ESBL, extended-spectrum  $\beta$ -lactamase; None detected, no gene encoding an acquired  $\beta$ -lactamase was detected by PCR. ESBL (cpase-) included isolates carrying CTX-M-type [CTX-M-15 ( $n = 28$ ), CTX-M-2 ( $n = 8$ ), CTX-M-15 and CTX-M-2 or TEM-type ESBL ( $n = 7$ ), CTX-M-1 group ( $n = 7$ ), CTX-M-8 ( $n = 1$ )] and SHV-type ( $n = 1$ ) ESBLs that were assumed to harbor permeability defects. One meropenem-nonsusceptible (MEM-NS) isolate collected in Guatemala was not molecularly characterized for  $\beta$ -lactamase genes. No MEM-NS Enterobacterales isolates were collected in the Dominican Republic during the surveyed time period.

as 99.8% of ESBL-positive isolates. Percentages of susceptibility to ceftazidime-avibactam were similar to or greater than observed for other  $\beta$ -lactams, including meropenem (95.7–100% susceptible) and piperacillin tazobactam (56.5–85.7% susceptible), among ESBL-positive, AmpC-positive, and ESBL- and AmpC-positive isolates. The activities of all  $\beta$ -lactams tested were significantly reduced compared to ceftazidime-avibactam against KPC-positive (2.5–9.1% susceptible) and

OXA-48-like-positive (0–37.0% susceptible) isolates. As anticipated, ceftazidime-avibactam, similar to all other  $\beta$ -lactams, was poorly active against isolates carrying MBLs; only tigecycline retained *in vitro* activity against >50% of MBL-positive isolates.

Of the 2521 *P. aeruginosa* isolates collected in Latin America from 2017 to 2019, 86.9% were susceptible to ceftazidime-avibactam (MIC<sub>90</sub>, 32  $\mu$ g/ml) (Table 4). Percent susceptibilities to



**Table 3 – In vitro activity of ceftazidime-avibactam and comparator agents against  $\beta$ -lactamase-positive Enterobacterales isolates collected in the Latin American region as part of the ATLAS global surveillance program from 2017-2019.**

Organism, phenotype/genotype (no. of isolates)	Antimicrobial agent	MIC ( $\mu\text{g/ml}$ )		Interpretation (CLSI)		
		MIC <sub>50</sub>	MIC <sub>90</sub>	% Susceptible	% Intermediate	% Resistant
ESBL-positive (1816) <sup>a</sup>	Ceftazidime-avibactam	0.25	0.5	99.8	NA	0.2
	Ceftazidime	32	128	13.1	11.2	75.7
	Cefepime	>16	>16	4.4	20.2	75.4
	Piperacillin-tazobactam	8	>64	76.0	11.7	12.3
	Meropenem	$\leq 0.06$	$\leq 0.06$	97.3	1.8	0.9
	Levofloxacin	>8	>8	18.9	10.2	70.8
	Amikacin	4	8	95.1	1.9	3.0
	Colistin	0.5	1	NA	97.2	2.8
	Tigecycline	0.25	1	97.7	1.9	0.4
	AmpC-positive (49) <sup>b</sup>	Ceftazidime-avibactam	0.12	0.25	100	NA
Ceftazidime		32	128	6.1	16.3	77.6
Cefepime		0.25	2	91.8	8.2	0.0
Piperacillin-tazobactam		4	>64	85.7	2.0	12.2
Meropenem		$\leq 0.06$	0.12	100	0	0
Levofloxacin		8	>8	28.6	8.2	63.3
Amikacin		2	4	100	0	0
Colistin		0.25	>8	NA	85.7	14.3
Tigecycline		0.25	1	95.9	2.0	2.0
ESBL-positive + AmpC-positive (23) <sup>c</sup>		Ceftazidime-avibactam	0.25	2	100	NA
	Ceftazidime	64	>128	0	8.7	91.3
	Cefepime	>16	>16	8.7	13.0	78.3
	Piperacillin-tazobactam	16	>64	56.5	17.4	26.1
	Meropenem	$\leq 0.06$	0.25	95.7	0	4.3
	Levofloxacin	>8	>8	21.7	8.7	69.6
	Amikacin	2	8	91.3	0	8.7
	Colistin	0.25	0.5	NA	95.7	4.3
	Tigecycline	0.25	1	95.7	4.3	0
	KPC-positive (364) <sup>d</sup>	Ceftazidime-avibactam	0.5	2	99.5	NA
Ceftazidime		64	>128	9.1	5.8	85.2
Cefepime		>16	>16	7.7	9.3	83.0
Piperacillin-tazobactam		>64	>64	2.5	5.8	91.8
Meropenem		>8	>8	6.3	5.5	88.2
Levofloxacin		>8	>8	16.5	6.6	76.9
Amikacin		4	>32	76.9	12.9	10.2
Colistin		0.5	>8	NA	75.3	24.7
Tigecycline		0.5	2	97.8	1.9	0.3
GES-type carbapenemase-positive (5) <sup>e</sup>		Ceftazidime-avibactam	–	–	100	NA
	Ceftazidime	–	–	0	0	100
	Cefepime	–	–	40.0	60.0	0
	Piperacillin-tazobactam	–	–	20.0	20.0	60.0
	Meropenem	–	–	100	0	0
	Levofloxacin	–	–	60.0	40.0	0

Table 3 (continued)

Organism, phenotype/genotype (no. of isolates)	Antimicrobial agent	MIC ( $\mu\text{g/ml}$ )		Interpretation (CLSI)		
		MIC <sub>50</sub>	MIC <sub>90</sub>	% Susceptible	% Intermediate	% Resistant
OXA-48-like-positive (27) <sup>f</sup>	Amikacin	–	–	40.0	60.0	0
	Colistin	–	–	NA	100	0
	Tigecycline	–	–	100	0	0
	Ceftazidime-avibactam	0.5	1	100	NA	0
	Ceftazidime	64	>128	7.4	0	92.6
	Cefepime	>16	>16	14.8	0	85.2
	Piperacillin-tazobactam	>64	>64	0	7.4	92.6
	Meropenem	2	>8	37.0	18.5	44.4
	Levofloxacin	>8	>8	18.5	7.4	74.1
	Amikacin	2	16	96.3	3.7	0
	Colistin	0.5	1	NA	100	0
MBL-positive (146) <sup>g</sup>	Tigecycline	1	2	100	0	0
	Ceftazidime-avibactam	>128	>128	1.4	NA	98.6
	Ceftazidime	>128	>128	0	0	100
	Cefepime	>16	>16	0	8.2	91.8
	Piperacillin-tazobactam	>64	>64	2.7	8.2	89.0
	Meropenem	>8	>8	4.8	2.1	93.2
	Levofloxacin	8	>8	21.9	15.8	62.3
	Amikacin	32	>32	49.3	6.8	43.8
	Colistin	0.5	>8	NA	82.9	17.1
	Tigecycline	0.5	2	91.8	7.5	0.7

<sup>a</sup> ESBL-positive, isolates in which one or more acquired  $\beta$ -lactamase genes encoding an ESBL was detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases but does not include isolates that co-carry AmpC  $\beta$ -lactamases or KPC, GES-type, OXA-48-like or MBL carbapenemases.

<sup>b</sup> AmpC-positive, isolates in which an acquired  $\beta$ -lactamase gene encoding an AmpC  $\beta$ -lactamase was detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases but does not include isolates that co-carry other serine  $\beta$ -lactamases or MBLs.

<sup>c</sup> ESBL-positive and AmpC-positive, isolates in which acquired  $\beta$ -lactamase genes encoding ESBL and AmpC  $\beta$ -lactamases were detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases but does not include isolates that co-carry serine carbapenemases or MBLs.

<sup>d</sup> KPC-positive, isolates in which a gene encoding a KPC carbapenemase was detected by PCR; includes isolates that co-carry other serine  $\beta$ -lactamases (including one isolate that co-carried KPC-2 and OXA-48) but does not include isolates that co-carry MBLs.

<sup>e</sup> GES-type carbapenemase-positive, isolates in which a gene encoding GES-20 was detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases but does not include isolates that co-carry other serine  $\beta$ -lactamases or MBLs.

<sup>f</sup> OXA-48-like positive, isolates in which a gene encoding an OXA-48-like enzyme (OXA-48, OXA-181, OXA-232, OXA-163, OXA-370) was detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases or ESBLs but does not include isolates that co-carry AmpC, GES-type or KPC serine  $\beta$ -lactamases or MBLs.

<sup>g</sup> MBL-positive, isolates in which a gene encoding an NDM-type, IMP-type or VIM-type MBL was detected by PCR; includes isolates that co-carry serine  $\beta$ -lactamases (original (narrow) spectrum  $\beta$ -lactamases, ESBLs, AmpC  $\beta$ -lactamases, GES-type, KPC, and OXA-48-like  $\beta$ -lactamases) and one isolate that carried two MBLs (IMP-27 and NDM-1).

**Table 4 – In vitro activity of ceftazidime-avibactam and comparator agents against *P. aeruginosa* isolates collected in the Latin American region as part of the ATLAS global surveillance program from 2017 to 2019.**

Organism, phenotype/genotype (no. of isolates)	Antimicrobial agent	MIC ( $\mu\text{g/ml}$ )		Interpretation (CLSI)		
		MIC <sub>50</sub>	MIC <sub>90</sub>	% Susceptible	% Intermediate	% Resistant
Latin America, All <i>P. aeruginosa</i> (2521)	Ceftazidime-avibactam	2	32	86.9	NA	13.1
	Ceftazidime	4	64	71.5	4.3	24.2
	Cefepime	4	>16	72.2	10.4	17.5
	Piperacillin-tazobactam	8	>64	69.1	13.1	17.7
	Meropenem	0.5	>8	66.9	5.9	27.3
	Levofloxacin	0.5	>8	62.7	8.2	29.1
	Amikacin	4	>32	82.1	2.6	15.3
	Colistin	1	2	NA	99.8	0.2
Meropenem-NS (835)	Ceftazidime-avibactam	8	64	61.9	NA	38.1
	Ceftazidime	32	>128	35.3	7.4	57.2
	Cefepime	16	>16	32.9	19.9	47.2
	Piperacillin-tazobactam	64	>64	28.6	28.7	42.6
	Meropenem	>8	>8	0	17.7	82.3
	Levofloxacin	>8	>8	22.0	10.1	67.9
	Amikacin	16	>32	52.3	5.4	42.3
	Colistin	1	2	NA	99.8	0.2
Meropenem-NS, MBL-, carbapenemase+ (80)	Ceftazidime-avibactam	16	64	45.0	NA	55.0
	Ceftazidime	128	>128	2.5	3.8	93.8
	Cefepime	>16	>16	2.5	0	97.5
	Piperacillin-tazobactam	>64	>64	1.3	2.5	96.3
	Meropenem	>8	>8	0	0	100
	Levofloxacin	>8	>8	8.8	1.3	90.0
	Amikacin	>32	>32	35.0	8.8	56.3
	Colistin	1	1	NA	100	0
Meropenem-NS, MBL-, carbapenemase- (541)	Ceftazidime-avibactam	4	16	87.2	NA	12.8
	Ceftazidime	8	128	53.6	8.3	38.1
	Cefepime	16	>16	48.8	20.0	31.2
	Piperacillin-tazobactam	32	>64	42.1	22.4	35.5
	Meropenem	8	>8	0	26.4	73.6
	Levofloxacin	4	>8	31.2	14.6	54.2
	Amikacin	8	>32	70.1	3.7	26.2
	Colistin	1	2	NA	99.6	0.4
Multidrug-resistant (590)	Ceftazidime-avibactam	16	128	49.0	NA	51.0
	Ceftazidime	64	>128	10.5	6.6	82.9
	Cefepime	>16	>16	5.1	22.4	72.5
	Piperacillin-tazobactam	>64	>64	5.9	28.1	65.9
	Meropenem	>8	>8	9.0	5.3	85.8
	Levofloxacin	>8	>8	9.5	4.4	86.1
	Amikacin	>32	>32	34.7	5.9	59.3
	Colistin	1	2	NA	99.7	0.3

Abbreviations: NS, non-susceptible; MBL-, no gene encoding a metallo- $\beta$ -lactamase was detected by PCR; carbapenemase+/-, a gene encoding a serine carbapenemase was (+) or was not (-) detected by PCR; multidrug-resistant, isolates resistant to three or more sentinel agents from different drug classes; NA, no breakpoint available.

other tested agents ranged from 4.8% to 24.2% lower than for ceftazidime-avibactam, with 71.5% of isolates susceptible to ceftazidime alone and 66.9% testing as susceptible to meropenem. Among the subset of all meropenem-nonsusceptible isolates ( $n = 835$ , included 214 MBL-positive isolates), 61.9% of isolates were susceptible to ceftazidime-avibactam. When MBL-positive isolates were removed from the subset, susceptibility to ceftazidime-avibactam increased to 87.2% among meropenem-nonsusceptible and carbapenemase-negative isolates and was 45.0% among serine carbapenemase-positive isolates. Against MDR isolates, which included MBL-positive isolates, 49.0% of isolates were susceptible to ceftazidime-avibactam. Ceftazidime-avibactam displayed the highest percentages of susceptibility of all agents tested among all

meropenem-nonsusceptible isolates, subsets of meropenem-nonsusceptible, carbapenemase-negative and MBL-negative, carbapenemase-positive isolates, and MDR isolates.

Analyzing data by country of origin, percentages of susceptibility to ceftazidime-avibactam were >87% in seven of the 10 countries surveyed, with the lowest values observed for Mexico, Chile and Venezuela (75.9–80.2% susceptible) (Table 5). The overall percentage of *P. aeruginosa* isolates that were meropenem non-susceptible was 33.1% (Table 4) and ranged from 7.8% (Dominican Republic) to 46.7% (Chile) (Table 5). Percentages of susceptibility of meropenem-nonsusceptible isolates to ceftazidime-avibactam were >50% in all countries except Venezuela (26.0% susceptible) and were  $\geq 80\%$  for isolates collected in Argentina (91.8%), Brazil (86.1%), Costa Rica (87.5%),

**Table 5 – Percentages of *P. aeruginosa* isolates collected in ten Latin American countries as part of the ATLAS global surveillance program from 2017 to 2019 that were susceptible to ceftazidime-avibactam and comparator agents.**

Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)					
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-	Multidrug-resistant
Argentina		(308)	(85)	(80) <sup>a</sup>			(71)
	Ceftazidime-avibactam	97.4	91.8	97.5			88.7
	Ceftazidime	75.0	36.5	38.8			16.9
	Cefepime	73.7	30.6	32.5			7.0
	Piperacillin-tazobactam	69.8	20.0	21.3			5.6
	Meropenem	72.4	0	0			15.5
	Levofloxacin	62.0	16.5	17.5			11.3
	Amikacin	85.1	55.3	57.5			43.7
Colistin	NA	NA	NA			NA	
Brazil		(425)	(115)	(109) <sup>b</sup>			(70)
	Ceftazidime-avibactam	95.3	86.1	90.8			72.9
	Ceftazidime	78.6	62.6	66.1			22.9
	Cefepime	78.4	55.7	58.7			4.3
	Piperacillin-tazobactam	73.6	47.8	50.5			5.7
	Meropenem	72.9	0	0			30.0
	Levofloxacin	69.2	34.8	36.7			15.7
	Amikacin	90.8	78.3	80.7			50.0
Colistin	NA	NA	NA			NA	
Chile		(257)	(120)		(16)	(60)	(84)
	Ceftazidime-avibactam	77.4	51.7		75.0	78.3	42.9
	Ceftazidime	58.0	18.3		0	36.7	1.2
	Cefepime	60.3	21.7		0	38.3	1.2
	Piperacillin-tazobactam	57.6	16.7		0	30.0	6.0
	Meropenem	53.3	0		0	0	1.2
	Levofloxacin	49.0	15.0		0	25.0	3.6
	Amikacin	77.0	56.7		87.5	76.7	45.2
Colistin	NA	NA		NA	NA	NA	
Colombia		(384)	(124)		(26)	(68)	(86)
	Ceftazidime-avibactam	87.5	62.1		53.8	92.6	46.5
	Ceftazidime	71.4	35.5		0	63.2	5.8
	Cefepime	71.9	33.9		0	61.8	1.2
	Piperacillin-tazobactam	69.3	29.0		0	50.0	5.8
	Meropenem	67.7	0		0	0	10.5
	Levofloxacin	67.2	26.6		19.2	39.7	19.8
	Amikacin	85.9	60.5		30.8	92.6	45.3
Colistin	NA	NA		NA	NA	NA	

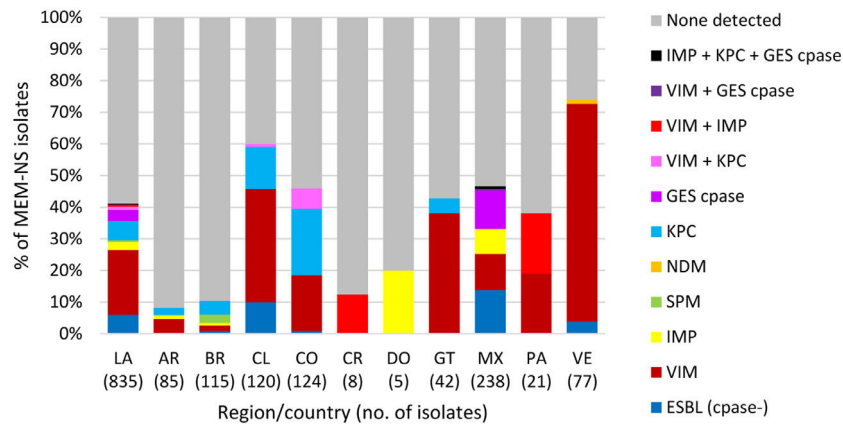
Table 5 (continued)							
Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)					
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-	Multidrug-resistant
Costa Rica		(55)	(8)	(7) <sup>c</sup>			(4)
	Ceftazidime-avibactam	96.4	87.5	100			50.0
	Ceftazidime	81.8	87.5	100			25.0
	Cefepime	94.5	87.5	100			25.0
	Piperacillin-tazobactam	81.8	75.0	85.7			0
	Meropenem	85.5	0	0			50.0
	Levofloxacin	85.5	50.0	57.1			50.0
	Amikacin	98.2	87.5	100			75.0
Colistin	NA	NA	NA			NA	
Dominican Republic		(64)	(5)	(4) <sup>d</sup>			(2)
	Ceftazidime-avibactam	98.4	80.0	100			50.0
	Ceftazidime	93.8	60.0	75.0			0
	Cefepime	96.9	60.0	75.0			0
	Piperacillin-tazobactam	96.9	60.0	75.0			0
	Meropenem	92.2	0	0			0
	Levofloxacin	78.1	60.0	75.0			0
	Amikacin	87.5	80.0	100			50.0
Colistin	NA	NA	NA			NA	
Guatemala		(149)	(42)	(26) <sup>e</sup>			(33)
	Ceftazidime-avibactam	90.6	66.7	100			57.6
	Ceftazidime	75.8	38.1	57.7			18.2
	Cefepime	73.8	23.8	38.5			0
	Piperacillin-tazobactam	71.8	21.4	34.6			0
	Meropenem	71.8	0	0			6.1
	Levofloxacin	70.5	16.7	23.1			3.0
	Amikacin	79.2	33.3	50.0			15.2
Colistin	NA	NA	NA			NA	
Mexico		(562)	(238)		(29)	(160)	(167)
	Ceftazidime-avibactam	80.2	55.0		10.3	79.4	38.9
	Ceftazidime	64.4	32.8		3.4	48.1	9.6
	Cefepime	65.7	31.5		6.9	44.4	9.0
	Piperacillin-tazobactam	64.1	31.1		3.4	43.1	8.4
	Meropenem	57.7	0		0	0	3.6
	Levofloxacin	59.3	23.9		3.4	33.8	7.2
	Amikacin	74.6	42.9		3.4	56.9	25.7
Colistin	NA	NA		NA	NA	NA	

Table 5 (continued)

Country	Antimicrobial agent	Phenotype/genotype (no. of isolates) % Susceptible (CLSI)				
		All isolates	Meropenem-NS	Meropenem-NS, MBL-, All	Meropenem-NS, MBL-, carbapenemase+	Meropenem-NS, MBL-, carbapenemase-
Panama		(80)	(21)	(13) <sup>f</sup>		(11)
	Ceftazidime-avibactam	87.5	52.4	84.6		36.4
	Ceftazidime	82.5	38.1	61.5		18.2
	Cefepime	82.5	38.1	61.5		9.1
	Piperacillin-tazobactam	77.5	28.6	46.2		9.1
	Meropenem	73.8	0	0		9.1
	Levofloxacin	63.7	23.8	38.5		9.1
	Amikacin	88.8	57.1	92.3		45.5
Colistin	NA	NA	NA		NA	
Venezuela		(237)	(77)	(23) <sup>g</sup>		(62)
	Ceftazidime-avibactam	75.9	26.0	73.9		12.9
	Ceftazidime	71.3	18.2	56.5		4.8
	Cefepime	71.7	18.2	43.5		4.8
	Piperacillin-tazobactam	69.6	16.9	47.8		3.2
	Meropenem	67.5	0	0		0
	Levofloxacin	53.2	3.9	8.7		1.6
	Amikacin	74.3	23.4	60.9		8.1
Colistin	NA	NA	NA		NA	

Abbreviations: NS, non-susceptible; MBL-, no gene encoding a metallo- $\beta$ -lactamase was detected by PCR; carbapenemase+/-, a gene encoding a serine carbapenemase was (+) or was not (-) detected by PCR; multidrug-resistant, isolates resistant to three or more sentinel agents from different drug classes; NA, no breakpoint available.

- <sup>a</sup> Composed of 2 carbapenemase-positive and 78 carbapenemase-negative isolates.  
<sup>b</sup> Composed of 5 carbapenemase-positive and 104 carbapenemase-negative isolates.  
<sup>c</sup> Composed of 7 carbapenemase-negative isolates.  
<sup>d</sup> Composed of 4 carbapenemase-negative isolates.  
<sup>e</sup> Composed of 2 carbapenemase-positive and 24 carbapenemase-negative isolates.  
<sup>f</sup> Composed of 13 carbapenemase-negative isolates.  
<sup>g</sup> Composed of 23 carbapenemase-negative isolates.



**Fig. 3 –  $\beta$ -lactamases identified in meropenem-nonsusceptible *P. aeruginosa* collected in 10 Latin American countries as part of the ATLAS global surveillance program from 2017 to 2019**

LA, Latin America; AR, Argentina; BR, Brazil; CL, Chile; CO, Colombia; CR, Costa Rica; DO, Dominican Republic; GT, Guatemala; MX, Mexico; PA, Panama; VE, Venezuela; Cpase, carbapenemase; ESBL, extended-spectrum  $\beta$ -lactamase; None detected, no gene encoding an acquired  $\beta$ -lactamase was detected by PCR. ESBL (cpase-) included isolates carrying ESBL-like GES-type and PER-type  $\beta$ -lactamases.

and Dominican Republic (80.0%). A total of 23.4% of all *P. aeruginosa* displayed an MDR phenotype (Table 4), with percentages of MDR isolates ranging from 3.1% (Dominican Republic) to 32.7% (Chile) across the region (Table 5). Ceftazidime-avibactam was most active against MDR isolates from Argentina (88.7% susceptible; MIC<sub>90</sub>, 16  $\mu$ g/ml) and Brazil (72.9% susceptible; MIC<sub>90</sub>, 64  $\mu$ g/ml) and least active against MDR isolates from Venezuela, Panama, and Mexico (12.9–38.9% susceptible, MIC<sub>90</sub>, 128 - >128  $\mu$ g/ml). Amikacin was the only tested comparator that displayed greater activity than ceftazidime-avibactam in some countries.

No acquired ESBLs or carbapenemases were detected in 58.8% (491/835) of meropenem-nonsusceptible *P. aeruginosa* isolates screened for genes encoding  $\beta$ -lactamases (Fig. 3), implying the role of chromosomally-coded mechanisms in meropenem resistance such as alterations in OprD or efflux pump expression, likely combined with hyper-production of the intrinsic chromosomal AmpC  $\beta$ -lactamase of *P. aeruginosa*. Among  $\beta$ -lactamase-positive isolates, VIM-type MBLs were the most common acquired  $\beta$ -lactamases identified, followed by KPC. VIM-positive isolates were identified in all countries surveyed except the Dominican Republic and accounted for 68.8% of meropenem-nonsusceptible isolates collected in Venezuela and 36.7–38.1% of meropenem-nonsusceptible isolates from Chile, Guatemala, and Panama. KPC, which is rarely found in *P. aeruginosa* collected outside of Latin America, was identified in isolates from Argentina, Brazil, Chile, Colombia, Guatemala, and Mexico and was co-carried with VIM-type ( $n = 9$ ) or IMP-type and GES-type carbapenemases ( $n = 2$ ) in a small number of isolates. The majority (72.4%; 21/29) of *P. aeruginosa* carrying IMP-type MBLs and all isolates carrying GES-type carbapenemases were identified in Mexico, whereas SPM-positive isolates were only identified in Brazil. Carbapenemase-negative, ESBL-positive isolates were found primarily in Chile (10% of isolates, 12/120) and Mexico (13.9%, 33/238). Ceftazidime-avibactam was not active against isolates carrying MBLs (4.2% susceptible), as expected, and it also

demonstrated reduced activity against MBL-negative, ESBL-positive isolates (34.0% susceptible) and GES carbapenemase-positive isolates (10.3%) (Table 6). These isolates may have carried additional  $\beta$ -lactamases that were not included in the molecular testing algorithm and that were not inhibited by avibactam, or may contain non-enzymatic resistance mechanisms. In contrast, 64.7% of KPC-positive isolates (MIC<sub>90</sub>, 32  $\mu$ g/ml) and 92.5% of meropenem non-susceptible isolates in which no acquired  $\beta$ -lactamase was detected were susceptible to ceftazidime-avibactam (MIC<sub>90</sub>, 8  $\mu$ g/ml).

## Discussion

The current study summarizes the *in vitro* antimicrobial susceptibility and molecular  $\beta$ -lactamase carriage of isolates collected as part of the ATLAS global surveillance program from 2017 to 2019 in 10 Latin American countries. Earlier publications only included isolates from six of the 10 countries (Argentina, Brazil, Chile, Colombia, Mexico, and Venezuela) described in the current study.<sup>2,3</sup> In 2018, laboratory sites in Costa Rica, Dominican Republic, Guatemala, and Panama were added to the ATLAS global surveillance program and this is the first publication of data from those four countries as part of ATLAS or any other study.

In 2012–2015, 99.7% of Enterobacterales isolates ( $n = 7665$ ) collected in Latin America as part of the ATLAS surveillance program were susceptible to ceftazidime-avibactam.<sup>3,24</sup> In the 2015–2017 ATLAS report, 99.3% of Enterobacterales isolates ( $n = 7729$ ) were susceptible to ceftazidime-avibactam.<sup>2</sup> In the current study of 2017–2019 isolates, 98.1% of Enterobacterales were susceptible to ceftazidime-avibactam. The lower susceptibility to ceftazidime-avibactam observed in the current study is partly attributable to the inclusion of isolates from Guatemala, which displayed a >10% lower percentage of susceptibility than observed for isolates from all other countries

**Table 6 – In vitro activity of ceftazidime-avibactam and comparator agents against  $\beta$ -lactamase-positive *P. aeruginosa* isolates collected in the Latin American region as part of the ATLAS global surveillance program from 2017 to 2019.**

Organism, phenotype/genotype (no. of isolates)	Antimicrobial agent	MIC ( $\mu$ g/ml)		Interpretation (CLSI)		
		MIC <sub>50</sub>	MIC <sub>90</sub>	% Susceptible	% Intermediate	% Resistant
ESBL-positive (50) <sup>a</sup>	Ceftazidime-avibactam	32	128	34.0	NA	66.0
	Ceftazidime	>128	>128	0	0	100
	Cefepime	>16	>16	0	4.0	96.0
	Piperacillin-tazobactam	>64	>64	8.0	34.0	58.0
	Meropenem	>8	>8	0	16.0	84.0
	Levofloxacin	>8	>8	0	0	100
	Amikacin	>32	>32	14.0	4.0	82.0
	Colistin	1	2	NA	98.0	2.0
KPC-positive (51) <sup>b</sup>	Ceftazidime-avibactam	8	32	64.7	NA	35.3
	Ceftazidime	128	>128	2.0	3.9	94.1
	Cefepime	>16	>16	0	0	100
	Piperacillin-tazobactam	>64	>64	0	0	100
	Meropenem	>8	>8	0	0	100
	Levofloxacin	>8	>8	11.8	2.0	86.3
	Amikacin	16	>32	52.9	13.7	33.3
	Colistin	1	1	NA	100	0
GES-type carbapenemase-positive (29) <sup>c</sup>	Ceftazidime-avibactam	64	64	10.3	NA	89.7
	Ceftazidime	>128	>128	3.4	3.4	93.1
	Cefepime	>16	>16	6.9	0	93.1
	Piperacillin-tazobactam	>64	>64	3.4	6.9	89.7
	Meropenem	>8	>8	0	0	100
	Levofloxacin	>8	>8	3.4	0	96.6
	Amikacin	>32	>32	3.4	0	96.6
	Colistin	1	1	NA	100	0
MBL-positive (214) <sup>d</sup>	Ceftazidime-avibactam	32	>128	4.2	NA	95.8
	Ceftazidime	64	>128	1.4	6.5	92.1
	Cefepime	>16	>16	4.2	27.1	68.7
	Piperacillin-tazobactam	64	>64	4.7	54.7	40.7
	Meropenem	>8	>8	0	2.3	97.7
	Levofloxacin	>8	>8	3.7	1.9	94.4
	Amikacin	>32	>32	14.0	8.4	77.6
	Colistin	1	2	NA	100	0
No acquired $\beta$ -lactamase (479)	Ceftazidime-avibactam	4	8	92.5	NA	7.5
	Ceftazidime	8	64	59.9	8.1	31.9
	Cefepime	8	>16	54.5	21.9	23.6
	Piperacillin-tazobactam	32	>64	46.8	21.1	32.2
	Meropenem	8	>8	0	28.2	71.8
	Levofloxacin	2	>8	35.3	16.5	48.2
	Amikacin	4	>32	77.2	3.8	19.0
	Colistin	1	2	NA	99.8	0.2

<sup>a</sup> ESBL-positive, isolates in which one or more  $\beta$ -lactamase genes encoding a PER-type or GES-type ESBL was detected by PCR; does not include isolates that co-carry serine carbapenemases or MBLs.

<sup>b</sup> KPC-positive, isolates in which a gene encoding a KPC carbapenemase was detected by PCR; includes isolates that co-carry original (narrow) spectrum  $\beta$ -lactamases but does not include isolates that co-carry ESBLs or MBLs.

<sup>c</sup> GES-type carbapenemase-positive, isolates in which a gene encoding GES-20 was detected by PCR; includes isolates that co-carry GES-type ESBLs (GES-1, GES-19, GES-26) but does not include isolates that co-carry other serine  $\beta$ -lactamases or MBLs.

<sup>d</sup> MBL-positive, isolates in which a gene encoding an NDM-type, IMP-type, VIM-type or SPM-type MBL was detected by PCR; includes isolates that co-carry serine  $\beta$ -lactamases (original (narrow) spectrum  $\beta$ -lactamases, PER-type or GES-type ESBLs, KPC or GES-type carbapenemases) and five isolates co-carrying two MBLs (IMP-18 and VIM-2).

(Table 2). Removal of isolates from Guatemala from the current dataset resulted in 98.7% (7946/8052) of Enterobacterales being susceptible to ceftazidime-avibactam (data not shown), a 1.0% decrease from 2012–2015. Further limiting the dataset to only isolates collected in Argentina, Brazil, Chile, Colombia, Mexico, and Venezuela (i.e. the six countries included in the 2012–2015<sup>3</sup> and 2015–2017<sup>2</sup> study reports), 98.6% (7256/7358) of Enterobacterales collected in 2017–2019 were susceptible to ceftazidime-avibactam (data not shown), a 1.1% decrease from 2012 to 2015. The changes in susceptibility to

ceftazidime-avibactam correlated with increases in the incidence of MBLs in Latin American isolates over time: 0.2% of isolates collected in 2012–2015 were MBL-positive<sup>3</sup>; 0.6% of isolates collected in 2015–2017<sup>2</sup>; and 1.3% (95/7358; isolates from the six countries participating since 2012) or 1.7% (146/8416; isolates from all 10 countries) collected in 2017–2019 were MBL-positive (Fig. 2). The proportion of MBL-positive isolates collected in Guatemala in 2017–2019 (12.9% of isolates) was much higher than observed for the nine other countries surveyed ( $\leq 2.2\%$ ). Earlier data from Guatemala are not



published and it is not possible to determine whether the abundance of MBL-positive Enterobacterales is due to recent or distant emergence.

In the current study, KPC was the most common carbapenemase identified, accounting for 61.5% of meropenem-nonsusceptible Enterobacterales from all 10 countries surveyed and 66.5% (339/510; data not shown) of isolates from the six countries included in the 2012–2015 and 2015–2017 ATLAS reports. In the 2012–2015 report, KPC carbapenemases comprised 89.1% of detected carbapenemases and MBLs were only identified in isolates from Colombia, Mexico, and Venezuela.<sup>3</sup> In the current study, MBL-positive isolates were detected in eight of 10 countries surveyed and the proportion of carbapenemase-positive isolates that carried MBLs increased more than two-fold for isolates collected in Colombia and Venezuela compared to the 2012–2015 report.

Previous surveillance studies have reported phenotypic and genotypic ESBL rates in Latin American countries of 20–>40% for both *E. coli* and *K. pneumoniae*, as well as rates of carbapenem-resistant Enterobacterales that often exceed 10%, particularly for *K. pneumoniae* and *Enterobacter* spp.<sup>2,3,21–23</sup> In the current study, 6.0% of isolates were meropenem-resistant and genes encoding ESBLs were identified in 25.7% (2161/8416) of collected Enterobacterales isolates. The distribution of ESBL and carbapenemase types observed in the current study was in general agreement with previous reports for South American countries and Mexico<sup>2,3,29</sup> and the ESBL rate compared well to that reported for isolates tested in 2015–2017 (24.1%), when similar molecular testing criteria were applied.<sup>2</sup> In the current study, we found that ceftazidime-avibactam continues to inhibit ESBL-positive, AmpC-positive, ESBL- and AmpC-positive, GES-type carbapenemase-positive, and OXA-48-like-positive isolates of Enterobacterales ( $\geq 99.8\%$  susceptible), as was found in earlier studies.<sup>2,3</sup>

The current study identified 156 isolates of Enterobacterales (1.9% of all isolates) that were resistant to ceftazidime-avibactam (Table 1); 144 (92.3%) of these 156 isolates were MBL-positive (Table 3). The mechanism(s) of reduced susceptibility for the remaining 12 isolates may reflect the presence of an avibactam-insensitive  $\beta$ -lactamase that was not detected using the current molecular algorithm<sup>17,18</sup> or a combination of mechanisms, such as increased  $\beta$ -lactamase production with porin deficiency and altered efflux and/or penicillin-binding protein alterations.<sup>6–13,30,31</sup> It should be noted that four of these isolates carried PER-2 or PER-4  $\beta$ -lactamases that are inhibited less effectively by avibactam in some cases.<sup>13,15</sup>

In the current study, 86.9% of all isolates of *P. aeruginosa* tested from 10 Latin American countries were susceptible to ceftazidime-avibactam (Table 4), similar to the ATLAS study reports for isolates collected in 2012–2015 (87.4% susceptible)<sup>3</sup> and 2015–2017 (86.6% susceptible).<sup>2</sup> Other studies reported 84.0% (21/25) of *P. aeruginosa* collected in 2014–2015 to be susceptible to ceftazidime-avibactam<sup>19</sup> and a ceftazidime-avibactam MIC<sub>90</sub> of 16  $\mu\text{g/ml}$  for 13 isolates of *P. aeruginosa* tested in 2011.<sup>20</sup> Among clinical isolates of *P. aeruginosa* from Latin American countries, other investigators have reported country-specific percentages of susceptibility to ceftazidime that ranged from 50–80%, while 60–70% of isolates were carbapenem-susceptible,<sup>2,3,23</sup> similar to the findings in the current study.

In the current study, 25.6% of all meropenem-nonsusceptible *P. aeruginosa* isolates collected in Latin America and 24.8% of isolates from the six countries participating in the study since 2012 carried MBLs. In comparison, 110 MBL-positive *P. aeruginosa* isolates were identified among 750 isolates nonsusceptible to meropenem, doripenem or imipenem in the 2012–2015 ATLAS report.<sup>3</sup> VIM-type enzymes continue to predominate among carbapenemase-positive isolates in the region, comprising 60.5% of carbapenemase-positive *P. aeruginosa* from all 10 countries and 63.3% of carbapenemase-positive *P. aeruginosa* from the six countries participating since 2012, compared to  $\sim 50\%$  of carbapenemase-positive isolates reported in the 2012–2015 report.<sup>3</sup>

We conclude that clinical isolates of Enterobacterales collected from 10 Latin America countries in 2017–2019, including MBL-negative meropenem-nonsusceptible isolates and isolates with an MDR phenotype, were highly susceptible to ceftazidime-avibactam, which was comparably active or more active *in vitro* than currently available agents of last resort (e.g., amikacin, colistin, tigecycline) that are associated with well-established toxicities. Similarly, ceftazidime-avibactam was the most potent agent tested against isolates of *P. aeruginosa* collected in these same countries in 2017–2019. Ceftazidime-avibactam has retained its *in vitro* potency against clinical isolates of Enterobacterales and *P. aeruginosa* collected from hospitalized patients in Latin American countries since 2012. Regional and country prevalence of different carbapenem-resistance mechanisms do exist and must be considered when evaluating treatment options.

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## Conflicts of interest

KMK and DFS are employees of IHMA, who were paid consultants to Pfizer in connection with the development of this manuscript. MdFV, ELL, MB, AQ, PI and GGS are employees of Pfizer Inc. JAK is an employee of the University of Manitoba and Shared Health Manitoba and is a consultant to IHMA. The IHMA authors and JAK do not have personal financial interests in the sponsor of this manuscript. QA and MdFV are Pfizer stock holders.

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## Supplementary materials

Supplementary material associated with this article can be found in the online version at doi:10.1016/j.bjid.2021.101647.

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