



Sociedade
Brasileira de
Infectologia

The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Original article

Improvement of *Mycobacterium tuberculosis* detection in sputum using DNA extracted by sonication



Graziele Lima Bello^a, Franciele Costa Leite Morais^a, Jonas Michel Wolf^a, Mirela Gehlen^b, Tainá dos Santos Soares^c, Maria Laura Halon^d, Regina Bones Barcellos^{d,e}, Maria Lucia Rosa Rossetti^{a,e,*}

^a Universidade Luterana do Brasil (ULBRA), Programa de Pós-Graduação em Biologia Celular e Molecular Aplicada à Saúde, Canoas, RS, Brasil

^b Universidade Federal do Rio Grande do Sul (UFRGS), Programa de Pós-Graduação em Pneumologia, Porto Alegre, RS, Brasil

^c Universidade Luterana do Brasil (ULBRA), Curso de Biomedicina, Canoas, RS, Brasil

^d Secretaria do Estado do Rio Grande do Sul (SES/RS), Centro de Desenvolvimento Científico e Tecnológico (CDCT), Porto Alegre, RS, Brasil

^e Universidade Federal do Rio de Janeiro (UFRJ), Programa de Pós-Graduação em Clínica Médica, Rio de Janeiro, RJ, Brasil

ARTICLE INFO

Article history:

Received 10 May 2020

Accepted 16 August 2020

Available online 12 September 2020

Keywords:

Tuberculosis

Sputum

DNA extraction

Sonication

ABSTRACT

Tuberculosis (TB) is one of the infectious diseases with high mortality in the world. DNA amplification techniques have been used to overcome barriers to the diagnosis of this disease. However, the success of these methodologies is highly dependent on the DNA obtained from the sample. This study was carried out to verify whether the DNA extracted by sonication (in house method) could yield suitable DNA for amplification by real-time PCR (qPCR). Sixty sputum samples were submitted to DNA extraction using sonication compared to a commercial method (Detect-TB kit, Labtest/MG-Brazil). All DNA samples were amplified by qPCR for IS6110 region (IS6110-qPCR/SYBR Green assay). Out of 60 samples, 40 were positive for TB; of these, all had positive results when extracted by sonication (100%) and 80% when extracted by the commercial method. The limit of detection (LOD) of *Mycobacterium tuberculosis* (H37Rv strain) by qPCR was 14 CFU/mL when the DNA was extracted by sonication, compared to countless colonies when extracted by commercial kit. In conclusion, the sonication protocol (without purification step) proved to be a simple, fast, and suitable method for obtaining DNA for use in qPCR from sputum samples.

© 2020 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

* Corresponding author.

E-mail addresses: taina.soares@rede.ulbra.br (T.S. Soares), maria.rossetti@ulbra.br (M.L. Rossetti).

<https://doi.org/10.1016/j.bjid.2020.08.006>

1413-8670/© 2020 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Background

Tuberculosis (TB) is one of the main causes of death and the leading cause from a single infectious agent worldwide.¹ Millions of people continue to fall sick with TB each year,² more than 1.7 billion people (about 25 percent of the world population) are estimated to be infected with *M. tuberculosis*. In 2017, 10 million individuals became ill with TB and 1.6 million died.³

Early diagnosis is one of the strategies to combat the disease.⁴ Methodologies based on the amplification of nucleic acids by Polymerase Chain Reaction (PCR) for the detection of *M. tuberculosis* DNA have improved the diagnosis and can overcome problems associated with classical methods.⁵ The use of molecular techniques, besides contributing to detect the increase in the number of TB cases, also allows to quickly characterize resistance and define the existing genotypes in a given community.⁶

However, differences in sensitivity and specificity when using PCR techniques are a reality. There are many reasons for this variability. One is the applied DNA extraction protocol.⁷ The DNA extraction method must provide a good yield and DNA of quality, elimination of PCR inhibitors, implementable as a laboratory technique, resulting in a genetic material of good quality for additional molecular analyses, while still being simple and fast.⁸ Many commercial methods are available (QIAGEN QIAmp DNA mini kit, Nucleic Acid and Protein Purification-Macherey, Nagel, DynaMag™-2 Magnet, Detect-TB kit), whereas DNA purification with silica, magnetic beads, Chelex beads, among others, have been described to improve DNA quality.^{7,9,10} Also, many laboratories use *in-house* methods for this step, mainly to curb costs.^{11–13}

The DNA extraction method that uses the cellular disruption by sonication has also been used in several laboratories,^{14–16} but there is still doubt about the efficacy of DNA extracted directly from sputum clinical samples (which have a wide variety of microorganisms, many impurities, and dependence on good collection) for use in PCR, as the protocol does not include a purification step.^{17,18} Therefore, this study aimed to compare a sonication technique as the method for obtaining DNA from sputum samples with the commercial method (silica resin - Detect-TB kit, marketed by Labtest / MG-Brazil). IS6110 – qPCR/SYBRGreen was used as a confirmatory method for detecting bacterial DNA.¹⁹

Materials and methods

Samples

Sixty sputum samples were collected from patients who attended the Department of Tisiology and Leprosy in Canoas, Southern Brazil. Initially, to evaluate DNA extraction with the two protocols from the same bacterial concentration, 20 sputum samples known to be from TB-negative patients were contaminated with 25 µL (100 CFU/mL) of an aqueous suspension of *Mycobacterium bovis* strain.

qPCR amplification was verified in 20 sputum samples from patients with tuberculosis. To determine the limit of detection,

20 negative samples were contaminated with 25 µL of 10-fold dilutions of *M. tuberculosis* reference strain H37Rv.

The sputum samples used in this study were also assessed on smear slides, culture, and GeneXpert MTB/RIF (Cepheid, EUA). The flowchart of the analysis is presented in Fig. 1.

Sample preparation

The sputum samples were decontaminated and treated with 4% sodium hydroxide (NaOH) and PBS (Phosphate-buffered saline) solution containing sodium chloride, potassium chloride, sodium phosphate, monobasic potassium phosphate, and ultrapure water, at an adjusted pH of 7.4. Briefly, a total of 500 µL of sputum was transferred to a 1.5 mL microtube and added the corresponding volume of NaOH 4% solution, followed by vortex homogenization, and incubation at 37 °C for 15 min. Subsequently, it was centrifuged 3000 *g* for 15 min and the supernatant was discarded. The pellet was resuspended in 500 µL of PBS buffer and homogenized on a vortex.

DNA extraction

Protocol A - silica resin (Detect-TB kit, Labtest / MG-Brazil)

The silica resin method was described by Michelon et al. (2011)²⁰ as an adaptation of the Boom et al. protocol (1990).²¹ Briefly, the protocol consisted of a cell lysis step with a solution containing 8 M guanidine hydrochloride, pH 6.4, 40 mM EDTA and 2 g/dL polyoxyethylene octyl phenyl ether, followed by incubation in a thermoblock at 100 °C for 10 min, aiming at the release and solubilization of the genomic DNA. Purification of the released DNA was carried out through specific uptake by a silica resin in the presence of chaotropic agents. Subsequently, a wash sequence was performed with distinct solutions: the first containing guanidine hydrochloride, the second containing buffered sodium chloride concentrate at pH 7.0 and polyoxyethylene sorbitan monolaurate 0.1%, and the third containing concentrated sodium chloride and buffered at pH 7.5 with polyoxyethylene 0.1% monolaurate. After washing, the purified DNA was eluted in 25 µL of ultrapure water and frozen at –20 °C until use.

Protocol B - sonication (*in-house* method)

A microtube containing 500 µL of sputum sample pretreated as mentioned above, was placed at 95 °C for 20 min. After this step, it was placed in the sonication bath for 15 min, and then it was centrifuged for 5 min at 3000 *g*. Subsequently, 100 µL of the supernatant (containing the DNA) was transferred to a new microtube and frozen at –20 °C until use.

Quantitative assessment of extracted DNA

The DNA quantification was performed by spectrophotometry. The DNA absorbance and concentration were evaluated for nucleic acid purity (ideal ratio: A260/A280 ≥ 1.8 and A260/A230 = 2). Samples were quantified in the SpectraMaxPlus® (Eppendorf) spectrophotometer using 2 µL of DNA according to the manufacturer's instructions.

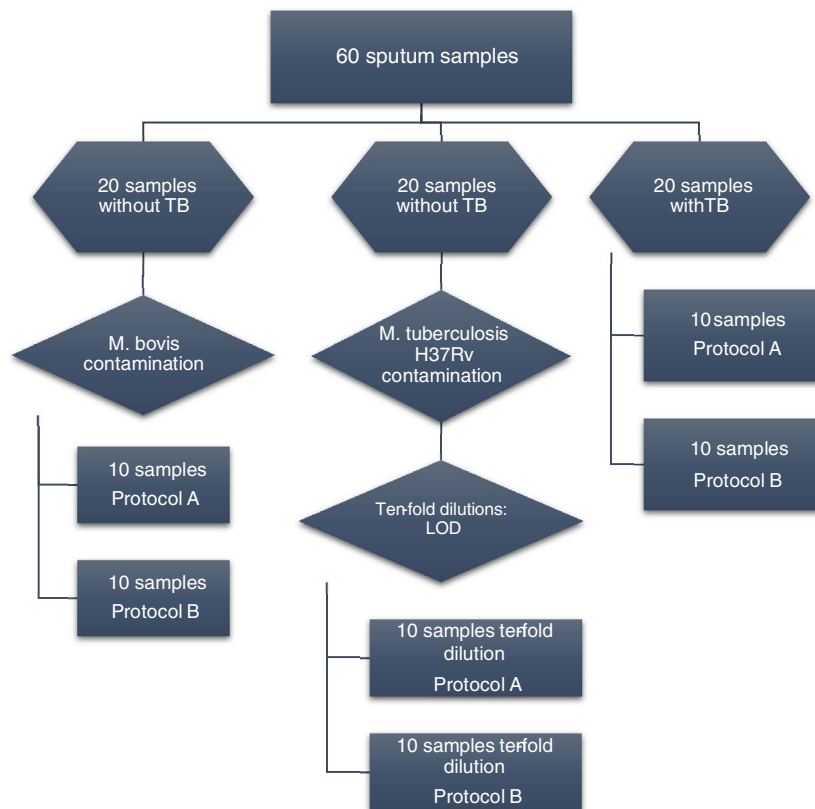


Fig. 1 – Flowchart of the samples used in the study.

Protocol A: Detect-TB kit, Labtest / MG-Brazil; Protocol B: Sonication; LOD: limit of detection.

Limit of detection (LOD)

Colonies of *M. tuberculosis* H37Rv strain grown in Ogawa Kudoh culture medium were scraped and placed in a glass tube containing beads to solubilize and homogenize bacterial colonies, and added ultrapure water (the volume can be stipulated by the operator). The turbidity of the bacterial solution was compared to the turbidity of the Macfarland number 1 scale (widely used in microbiology, especially for conventional antibiograms). Ten-fold dilutions (turbidity scale 1 solution) was performed. From these dilutions, 25 μ L were mixed to 20 TB-negative sputum samples. The same volume of 25 μ L of the dilutions was inoculated in 7H10 culture medium, and the CFUs was assessed for each dilution.

Real-time PCR (IS6110 – qPCR/SYBR Green)

For evaluation of the viability of the extracted DNA, IS6110 real-time PCR was performed with the Step One Real-Time PCR Systems (AB Applied Biosystems) equipment and the amplified products were detected using the SYBR[®] Green (fluorophore). The reaction was standardized in a final volume of 20 μ L containing 10 μ L of Fast SYBR[®] Green Master Mix, 4 pmol of each primer (INS1 5'CGTGAGGGCATGGAGGTGGC 3' and INS2 5'GCGTAGGCGTCGGTGACAAA 3') and 5 μ L of extracted DNA and quantified. The conditions of the amplification were as follows: activation of the enzyme at 95 °C for 20 s, denaturation at 95 °C for 1 s, and finally annealing and extension

at 62 °C for 20 s. For each run, a negative reaction control (PCR mix containing ultrapure water) and a positive control reference (*M. tuberculosis* H37Rv - 80 ng/ μ L) were used. Amplification and dissociation curves were analyzed with StepOne software version 2.3. For the analysis of the amplification curve, the parameters threshold (detectable fluorescence level) and baseline (defined limit of PCR cycles) were used to determine the Ct (cycle threshold) of each sample. The dissociation curve was used to observe the possible amplification of non-specific products or primer dimers. For standardization of the ideal concentration of primers at 4 pmol, it was used the analysis of the dissociation curve of known samples and the analysis of amplicons in 1.5% agarose gel, stained with ethidium bromide.

Standard curve and efficiency

The setup of the standard curve was performed according to the manufacturer's recommendations (AB Applied Biosystems-ThermoFisher Scientific)—five points and in duplicate—for assessing DNA amplification protocol. Ten-fold dilutions were performed from the quantified DNA of *M. tuberculosis* strain H37Rv (using as a genomic marker the element of insertion IS6110 of the bacterium, by qPCR). The PCR mixture contained 5 μ L of DNA from each dilution for each reaction, and 15 μ L of the mix, totaling the final volume of 20 μ L per reaction. Efficiency, calculated using StepOne software version 2.3, was of 96.15% (reference value: 90–100%), R^2 of 0.97 (indicates the closeness of fit between the linear regression line of the stan-

Table 1 – Comparison between the averages of the degrees of purity and DNA concentrations obtained by the two protocols applied to clinical samples contaminated with *M. bovis* strain.

Protocols	A260/280 (≥ 1.8)	A260/230 (< 2.0)	DNA ng/ μ L	Intensity amplicons electrophoresis gel
A	1.67 \pm 0.35	0.14 \pm 0.12	58.83 \pm 76.65	++
B	0.97 \pm 0.07	0.74 \pm 0.33	340.43 \pm 154.34	+++
p-values	<0.01*	<0.01*	<0.01*	

Protocols: A) Detect-TB Kit; B) Sonication.
* Significant statistical difference.

standard curve and the data of individual Ct points), -3.418 of slope (indicates the efficiency of amplification by PCR assay) and Y-inter of 16.87 (indicates the expected Ct for the sample).

Ethical aspects

The study was approved by the Lutheran University of Brazil (ULBRA) research ethics committee (approval number 2011-340H).

Statistical analysis

All data were analyzed using IBM[®] SPSS[®] statistics (version 21.0). The Shapiro-Wilk test was used for the analysis of type of data distribution. Student's T-test for independent samples and Mann-Whitney test were used to identify possible differences of DNA concentrations and Cts obtained between the two protocols evaluated. All tests performed were interpreted using a two-sided significance level of 5% ($p < 0.05$).

Results

The qPCR amplification of the DNA extracted from the 20 contaminated samples with *M. bovis* (10 DNAs extracted per protocol) showed that DNA extracted by Protocol A (silica resin) was detected in 8 of the 10 samples (80.0%) and all 10 DNAs extracted by Protocol B (sonication) were amplified (100%).

The spectrophotometric quantification of the DNAs amplified and purity evaluation by the different protocols are shown in (Table 1). Regarding DNA purity at wavelength 260/280 nm, significant differences were found between the results of two protocols (1.67 ± 0.35 vs. 0.97 ± 0.07 ; $p < 0.01$). At the wavelength of 260/230 nm, a statistical difference was detected between the data evaluated, Protocol A differed from Protocol B (0.14 ± 0.12 vs. 0.74 ± 0.33 ; $p < 0.01$). There was a significant difference in DNA concentration between the two protocols used in the analysis ($58.83 \text{ ng}/\mu\text{L} \pm 76.65 \text{ ng}/\mu\text{L}$ vs. $340.43 \text{ ng}/\mu\text{L} \pm 154.34 \text{ ng}/\mu\text{L}$; $p < 0.01$).

The amplifications Cts by real-time PCR and the means of each DNA extraction protocol are shown in (Table 2). Comparing Cts of protocol 1 vs. 2, there was statistical difference (27.73 ± 2.48 vs. 26.92 ± 0.89 ; $p < 0.01$).

Regarding the LOD (using ten-fold dilutions of the *M. tuberculosis* strain H37Rv), with protocol B the qPCR assay detected up to 14 CFUs (1: 1000 dilution), whereas with Protocol A the LOD was countless colonies.

Table 2 – Cts corresponding to the extraction protocols applied from DNA extracted from clinical samples contaminated with *M. bovis* strain.

Protocol A Detect-TB Kit	Protocol B Sonication
26.48	27.25
26.21	27.16
24.87	27.10
–	27.27
31.49	27.21
25.27	27.23
27.58	27.25
30.56	27.28
–	24.40
29.42	27.14
27.73 \pm 2.48	26.92 \pm 0.89 p-value = 0.01*

* Significant statistical difference.

Among the 20 samples evaluated from patients with TB, DNA extracted with Protocol A was detected TB in seven of the 10 (70.0%) samples and in all 10 (100%) DNA extracted with Protocol B (Table 3).

DNA purity and quantification extracted from *M. tuberculosis* sputum samples followed the same standards as those of *M. bovis* contaminated samples. Table 4 shows the mean purity and quantification of the nucleic acid extracted with the two protocols. Regarding DNA purity at the wavelength of 260/280 nm, there was significant difference between the two protocols (2.19 ± 1.62 vs. 1.09 ± 0.21 ; $p < 0.01$). When analyzing DNA purity at wavelength 260/230 nm, there was no significant difference between the two protocols (0.07 ± 0.04 vs. 0.68 ± 0.50 ; $p < 0.01$). Concerning DNA quantification, there was a significant difference ($20.75 \text{ ng}/\mu\text{L} \pm 11.74 \text{ ng}/\mu\text{L}$ vs. $37.01 \text{ ng}/\mu\text{L} \pm 36.91 \text{ ng}/\mu\text{L}$; $p < 0.01$) between the protocols used in the analysis.

The results of the Cts for the DNA extracted samples from patients with TB using the two protocols are shown in Table 5. The difference observed between Protocol A vs. B was not significant (29.03 ± 4.14 vs. 26.48 ± 4.44 ; $p = 0.23$) when considering the samples with *Mycobacterium* DNA detected.

Discussion

We assessed in this study if extraction DNA from sputum samples (simple and low-cost technique) with sonication would provide DNA suitable for the detection of the *M. tuberculosis* by qPCR, through the identification of the insertion

Table 3 – Real-time PCR detection of *M. tuberculosis* DNA in samples from patients with TB: results after DNA extraction by two protocols.

Clinical sputum sample	Protocol A		Protocol B
	Detect-TB Kit		Sonication
1	+		+
2	+		+
3	+		+
4	+		+
5	-		+
6	+		+
7	-		+
8	+		+
9	+		+
10	-		+

sequence IS6110, being reported as the most specific and widely explored genetic marker (number of IS6110 copies per genome might influence the sensitivity of the test).^{22,23} Sputum is the most common biological sample used for the diagnosis of pulmonary TB disease.²⁴ As compared to urine and blood, sputum is thicker and more viscous but provides better insight into TB and other respiratory-related diseases.²⁵ When used directly for DNA extraction and subsequent detection by PCR, it is known that sputum samples, present barriers related to the presence of internal Taq inhibitors, as well as presence of several microorganisms that are part of the microbiota of the respiratory and oral tracts.²⁶⁻²⁸ Thus, sample preparation and choice of an efficient method for DNA extraction is one of the main challenges for the molecular diagnosis of TB.²⁹

Regarding the use of the available commercial kits, it is known that they are generally preferred because they provide greater reproducibility, quality control, and automation potential.³⁰ Silica matrices (Detect-TB kit, marketed by Labtest/MG-Brazil - Protocol A) provide efficient and selective binding of nucleic acids (NA), having as characteristic the procedure of purification of DNA.³¹ The mechanism involved in this technique is the affinity between negatively charged NA and positively charged silica material, resulting in selective binding of NA.³² In clinical applications, even using kits, to obtain high yields of NA is still challenging, since sputum samples often exhibit a low final concentration of DNA, which is masked against a large background of contaminants.

In our study, there was variability in some standards regarding the quantitative evaluation of the extracted DNA. Nonetheless, Protocol B provided a higher amount of extracted

Table 5 – CTs corresponding to the extraction protocols applied from DNA extracted from clinical samples from patients with TB.

Sample	Protocol A Detect-TB Kit	Protocol B Sonication
1	33.18	28.43
2	33.80	35.28
3	32.10	29.32
4	28.06	27.56
5	-	24.55
6	26.84	27.06
7	-	20.03
8	26.75	27.97
9	22.49	23.88
10	-	20.77
	29.03 ± 4.14	26.48 ± 4.44 p-value = 0.23

DNA, lower Cts in the plot amplification (by qPCR) and profiles of fragments with a more uniform aspect, when visualized in electrophoresis. In line with our findings, the study by Krinitsina et al. (2015)³³ showed that the purity values and some DNA concentrations measured by absorbance, clearly differ from the visible data of real-time PCR and electrophoresis detection (fluorescence intensity of amplicons). Depending on the extraction method applied, it is important to point out that purification of DNA molecules is known to be influenced by their specificities of size, nucleotide composition, topology, and association with proteins.³⁰ We believe that reagent residues with different chemicals, or the need for further washes during the application of the protocols, may have influenced the absorbance reading. Therefore, a quantitative evaluation of the extracted DNA cannot be evaluated as an absolute parameter.

According to the study by Aldous et al. (2005),³⁴ it was possible to conclude that the purification step using silica resin was not necessary for the extraction of DNA from sputum samples before the PCR technique for identification of *M. tuberculosis* since it makes the method more extensive and impairs laboratory routine. Comparing the use of Kit (Detect-TB - Protocol A) with *in-house* method (Sonication - Protocol B), it was concluded that the DNA extracted by Kit presented higher purity, but the amount of extracted DNA was, in many cases, inconsistent for detection.³² In a study by Psifidi et al. (2015),³⁵ 11 different protocols were tested for DNA extraction and further used in genomic and microarray sequencing. Among them, the one that obtained the shortest execution time, lowest cost, and the best results were the *in-house* protocol, as our results also demonstrated.

Table 4 – Comparison between the averages of the degrees of purity and the DNA concentrations obtained by protocols applied to clinical samples of patients with TB.

Protocols	A260/280 (≥1.8)	A260/230 (<2.0)	DNA ng/μL	Intensity amplicons electrophoresis gel
A	2.19 ± 1.62	0.07 ± 0.04	20.75 ± 11.74	++
B	1.09 ± 0.21	0.68 ± 0.50	37.01 ± 36.91	+++
p-values	<0.01*	<0.01*	<0.01*	

Protocols: A) Detect-TB Kit; B) sonication.
* Significant statistical difference.

The analyses by Liu et al. (2013),³⁶ Greenly, and Tester (2015)³⁷ showed that the use of ultrasound allows cell walls to be cleaved by shock waves, created by rapid changes in pressure caused by sonication. The study by Granger et al. (2004)¹⁵ reported that the use of sonication improved mycobacterial DNA by removing background bacteria and other cells. In addition to these experiments, studies by Pchelintsev, Adams, and Nelson (2016)¹⁷ showed that the use of sonication minimizes the risk of cross-contamination between samples. Dai et al. (2016)¹⁶ concluded that the sonication-based technique is considered more promising, efficient, and economical for homogeneous cell decomposition, in a short period.

Regarding the limit of detection (LOD) of qPCR/SYBR Green assay, Protocol B provided a good detection capacity (up to 14 CFU/mL), even from smaller amounts of DNA (1:1000 dilution), consumed fewer reagents, with fewer execution steps, proving to be effective, less laborious, and fast method that presented greater potential for application in sputum samples.⁸ The LOD obtained in our study was similar to that found in the study by Chakravorty et al. (2017),³⁸ which analyzed the performance of Xpert MTB/RIF Ultra as a suitable point-of-care (POC) assay for TB diagnosis, and obtained a LOD of 15.6 CFU/mL.

We recognize potential study limitations. However, our findings are important for guiding the choice of DNA extraction method applicable to clinical sputum samples. Future studies may consolidate our findings, thus allowing greater understanding of the need for NA purification for molecular detection of potential microorganisms that cause infectious diseases.

Conclusions and perspectives

Identification techniques based on molecular biology have been established as potential tools for diagnosis. In light of this, this study showed that the extraction of DNA by sonication, even without presenting purification steps during the execution of the technique, provided suitable genetic material for identification by real-time PCR, presenting higher DNA concentrations, lower costs and complexity of execution. Thus, it is one of the most promising DNA extraction techniques for use in clinical sputum samples from patients, presenting high applicability to the diagnosis of TB, as well as the possible potential for molecular epidemiology analyses, such as Whole Genome Sequencing (WGS).

Funding

Graziele Bello and Jonas Michel Wolf were supported by CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior)/MEC/Brazil-Fellowship. Financing code 001.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgments

The authors wish to thank the Postgraduate Program in Cellular and Molecular Biology Applied to Health - Lutheran University of Brazil (ULBRA), Center for Scientific and Technological Development (CDCT-RS), and the Department of Tisiology and Leprosy of Canoas, Brazil.

REFERENCES

- Petersen E, Rao M, Ippolito G, Gualano G, Chakaya J, Ntoumi F, et al. World Tuberculosis Day March 24th 2019 Theme: "It's TIME" — International Journal of Infectious Diseases Tuberculosis Theme Series. *Int J Infect Dis.* 2019;80S:S1–5.
- WHO. Global Tuberculosis Report 2018 <https://www.who.int/tb/publications/global.report/en/> [accessed March 28, 2019].
- WHO. World Health Organization. Tuberculosis (TB). <http://www.who.int/topics/tuberculosis/en/>; 2016 [accessed 21, february 2016].
- Cheon SA, Cho HH, Kim J, Lee J, Kim HJ, Park TJ. Recent tuberculosis diagnosis toward the end TB strategy. *J Microbiol Methods.* 2016;123:51–61.
- Khosravi AD, Alami A, Meghdadi H, Hosseini AA. Identification of Mycobacterium tuberculosis in clinical specimens of patients suspected of having extrapulmonary tuberculosis by application of nested PCR on five different genes. *Front Cell Infect Microbiol.* 2017;7:1–7.
- Verza M, Schmid KB, Barcellos RB, Linck N, Bello GL, Scapin D, et al. Performance of a molecular assay to detect Mycobacterium tuberculosis complex DNA in clinical specimens: multicenter study in Brazil. *Mem Inst Oswaldo Cruz.* 2017;112:94–9.
- Knudsen BE, Bergmark L, Munk P, Lukjancenko O, Priemé A, Aarestrup FM, et al. Impact of sample type and DNA isolation procedure on genomic inference of microbiome composition. *MSystems.* 2016;1:e00095–16.
- De Almeida IN, Da Silva Carvalho W, Rossetti ML, Costa ERD, De Miranda SS. Evaluation of six different DNA extraction methods for detection of Mycobacterium tuberculosis by means of PCR-IS6110: preliminary study. *BMC Res Notes.* 2013;6:561.
- Walden C, Carbonero F, Zhang W. Assessing impacts of DNA extraction methods on next generation sequencing of water and wastewater samples. *J Microbiol Methods.* 2017;141:10–6.
- He H, Li R, Chen Y, Pan P, Tong W, Dong X, et al. Integrated DNA and RNA extraction using magnetic beads from viral pathogens causing acute respiratory infections. *Sci Rep.* 2017;7:45199.

11. Forsberg C, Jansson L, Ansell R, Hedman J. High-throughput DNA extraction of forensic adhesive tapes. *Forensic Sci Int Genet.* 2016;24:158–63.
12. Breacker C, Barber I, Norton WHJ, McDearmid JR, Tilley CA. A low-cost method of skin swabbing for the collection of DNA samples from small laboratory fish. *Zebrafish.* 2017;14:35–41.
13. Bentaleb EM, Abid M, El Messaoudi MD, Lakssir B, Ressami EM, Amzazi S, et al. Development and evaluation of an in-house single step loop-mediated isothermal amplification (SS-LAMP) assay for the detection of *Mycobacterium tuberculosis* complex in sputum samples from Moroccan patients. *BMC Infect Dis.* 2016;16:517.
14. Rabodoarivelo MS, Aerts M, Vandamme P, Palomino JC, Rasolofo V, Martin A. Optimizing of a protein extraction method for *Mycobacterium tuberculosis* proteome analysis using mass spectrometry. *J Microbiol Methods.* 2016;131:144–7.
15. Granger K, Moore RJ, Davies JK, Vaughan JA, Stiles PL, Stewart DJ, et al. Recovery of *Mycobacterium avium* subspecies paratuberculosis from the natural host for the extraction and analysis in vivo-derived RNA. *J Microbiol Methods.* 2004;57:241–9.
16. Dai X, Chen S, Li N, Yan H. Quantitative and qualitative validations of a sonication-based DNA extraction approach for PCR-based molecular biological analyses. *Anal Biochem.* 2016;501:44–6.
17. Pchelintsev NA, Adams PD, Nelson DM. Critical parameters for efficient sonication and improved chromatin immunoprecipitation of high molecular weight proteins. *PLoS One.* 2016;11:e0148023.
18. Stamper T, Wong ES, Timm A, DeBry RW. Validating sonication as a DNA extraction method for use with carrion flies. *Forensic Sci Int.* 2017;275:171–7.
19. Roychowdhury T, Mandal S, Bhattacharya A. Analysis of IS6110 insertion sites provide a glimpse into genome evolution of *Mycobacterium tuberculosis*. *Sci Rep.* 2015;5:1–10.
20. Michelon CT, Rosso F, Schmid KB, Sperhacker RD, Oliveira MM, Kritski AL, et al. Colorimetric microwell plate reverse-hybridization assay for *Mycobacterium tuberculosis* detection. *Mem Inst Oswaldo Cruz.* 2011;106:194–9.
21. Boom R, Sol CJA, Salimans MMM, Jansen CL, Wertheim-Van Dillen PME, Van Der Noordaa J. Rapid and simple method for purification of nucleic acids. *J Clin Microbiol.* 1990;28:495–503.
22. Sankar S, Kuppanan S, Balakrishnan B, Nandagopal B. Analysis of sequence diversity among IS6110 sequence of *Mycobacterium tuberculosis*: possible implications for PCR based detection. *Bioinformation.* 2011;6:283–5.
23. Sposito FL, Campanerut PA, Ghiraldi LD, Leite CQ, Hirata MH, Hirata RD, et al. Multiplex-PCR for differentiation of *Mycobacterium bovis* from *Mycobacterium tuberculosis* complex. *Braz J Microbiol.* 2014;45:841–3.
24. Allen V, Nicol MP, Tow LA. Sputum processing prior to *Mycobacterium tuberculosis* detection by culture or nucleic acid amplification testing: a narrative review. *Res Rev J Microbiol Biotechnol.* 2016;5:96–108.
25. Altiner A, Wilm S, Däubener W, Bormann C, Pentzek M, Abholz HH, et al. Sputum colour for diagnosis of a bacterial infection in patients with acute cough. *Scand J Prim Health Care.* 2009;27:70–3.
26. Soundararajan L, Kambli P, Priyadarshini S, Let B, Murugan S, Iravatham C, et al. Whole genome enrichment approach for rapid detection of *Mycobacterium tuberculosis* and drug resistance-associated mutations from direct sputum sequencing. *Tuberculosis.* 2020;121:101915.
27. Böddinghaus B, Wichelhaus TA, Brade V, Bittner T. Removal of PCR inhibitors by silica membranes: evaluating the Amplicor *Mycobacterium tuberculosis* kit. *J Clin Microbiol.* 2001;39:3750e2.
28. Amicosante M, Richeldi L, Trenti G, Paone G, Campa M, Bisetti A, et al. Inactivation of polymerase inhibitors for *Mycobacterium tuberculosis* DNA amplification in sputum by using capture resin. *J Clin Microbiol.* 1995;33:629e30.
29. Ali N, Rampazzo RDCP, Costa ADIT, Krieger MA. Current nucleic acid extraction methods and their implications to point-of-care diagnostics. *Biomed Res Int.* 2017:1–13.
30. Becker L, Steglich M, Fuchs S, Werner G, Nübel U. Comparison of six commercial kits to extract bacterial chromosome and plasmid DNA for MiSeq sequencing. *Sci Rep.* 2016;6:28063.
31. Esser KH, Marx WH, Lisowsky T. maxXbond: first regeneration system for DNA binding silica matrices. *Nat Methods.* 2006;3:2005–6.
32. Mutiui AI, Brandl CJ. RNA isolation from yeast using silica matrices. *J Biomol Tech.* 2005;16:316–7.
33. Krinitsina AA, Sizova TV, Zaika MA, Speranskaya AS, Sukhorukov AP. A rapid and cost-effective method for DNA extraction from archival herbarium specimens. *Biochem.* 2015;80:1478–84.
34. Aldous WK, Pounder JI, Cloud JL, Woods GL. Comparison of six methods of extracting *Mycobacterium tuberculosis* DNA from processed sputum for testing by quantitative real-time PCR. *J Clin Microbiol.* 2005;43:2471–3.
35. Psifidi A, Dovas CI, Bramis G, Lazou T, Russel CL, Arsenos G, et al. Comparison of eleven methods for genomic DNA extraction suitable for large-scale whole-genome genotyping and long-term DNA banking using blood samples. *PLoS One.* 2015;10:e0115960.
36. Liu D, Zeng XA, Sun DW, Han Z. Disruption and protein release by ultrasonication of yeast cells. *Innov Food Sci Emerg Technol.* 2013;18:132–7.
37. Greenly JM, Tester JW. Ultrasonic cavitation for disruption of microalgae. *Bioresour Technol.* 2015;184:276–9.
38. Chakravorty S, Simmons AM, Rowneki M, Parmar H, Cao Y, Ryan J, et al. The new Xpert MTB/RIF ultra: improving detection of *Mycobacterium tuberculosis* and resistance to Rifampin in an assay suitable for point-of-care testing. *MBio.* 2017;8.