

The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Original article

Propionibacterium acnes-killed attenuates the inflammatory response and protects mice from sepsis by modulating inflammatory factors

José Bruno Nunes Ferreira da Silva^{a,b}, Samara Kelly Mendonça de Oliveira^a, Ingrid Araújo Campos^{a,b}, Carlson Helder Reis de Carvalho-Júnior^a, Thiago da Cunha Coutinho^a, Teresinha Gonçalves Silva^{a,b,*}

^a Laboratory Bioassays for Research of Drugs, Department of Antibiotics, Biological Science Center, Universidade Federal de Pernambuco, Recife, PE, Brazil

^b Postgraduate Program in Industrial Biotechnology, Department of Antibiotics, Biological Science Center, Universidade Federal de Pernambuco, Recife, PE, Brazil

ARTICLE INFO

Article history: Received 17 July 2012 Accepted 17 August 2012 Available online 3 January 2013

Keywords: Propionibacterium acnes Immunomodulation Sepsis Cytokines

$A \hspace{0.1in} B \hspace{0.1in} S \hspace{0.1in} T \hspace{0.1in} R \hspace{0.1in} A \hspace{0.1in} C \hspace{0.1in} T$

Background: Sepsis is a systemic inflammation associated with infection caused by pathogenic micro-organisms with high mortality rates.

Objective: In this study, we investigated the protective effect of *Propionibacterium acnes*-killed against polymicrobial sepsis induced by cecal ligation and puncture.

Methods: The mice were treated by intramuscular route in 1, 3, 5, and 7 days before the cecal ligation and puncture induction. The control group animals received vehicle (saline solution 0.9%) and the animals of the treated group received the *P. acnes*-killed (0.4 mg/animal). After anesthesia, midline laparotomy was performed with exposure of cecum followed by ligature and one transverse perforation of the same, with a 18 G needle, for induction of lethal sepsis. After surgery, the cecum of the animals was replaced into the peritoneal cavity, and it was closed with a 4.0 nylon suture. The survival of animals subjected to lethal sepsis, neutrophil migration, the number of bacteria, TNF- α , MCP-1, IL-6, and IL-10 were performed in the peritoneal lavage.

Results: Prophylactic treatment with P. acnes-killed increased the survival of the animals, followed by a significant decrease in the TNF- α , IL-10, and MCP-1 levels, 6 h after cecal ligation and puncture. Furthermore, P. acnes-killed administration reduced the number of bacteria in the peritoneal cavity with increased migration of leukocytes, especially neutrophils.

*Conclusion: P. acnes-*killed promoted increased survival rate of animals with sepsis, in part attributed to its immunomodulatory properties against pathogenic microorganisms, as well as better control of infection by reducing bacterial counts.

© 2013 Elsevier Editora Ltda. Este é um artigo Open Access sob a licença de CC BY-NC-ND

E-mail address: teresinha.goncalves@pq.cnpq.br (T.G. Silva).

1413-8670 © 2013 Elsevier Editora Ltda. Este é um artigo Open Access sob a licença de CC BY-NC-ND http://dx.doi.org/10.1016/j.bjid.2012.08.023

^{*} Corresponding author at: Departamento de Antibióticos, Universidade Federal de Pernambuco, Rua Prof. Artur Sá, s/n, Cidade Universitária, Recife, PE, Brazil. Tel.: +55 81 21268347; fax: +55 81 21268346.

Introduction

Sepsis is a systemic inflammation associated with infection caused by pathogenic micro-organisms. Gram-negative and Gram-positive bacteria, viruses, and fungi can be responsible for the excessive stimulation of immune system and deregulate the protective response of the host.¹ Sepsis remains a clinical problem in the XXI century. The increased incidence and mortality has evolved substantially over the past two decades, particularly in intensive care units.^{2,3} Despite the intense treatment to contain the infection in patients with sepsis, the use of drugs has not been satisfactory in clinical applications, resulting in a significant impact on health systems around the world, both economically and socially.⁴

The immunomodulation in sepsis has evolved in recent years, stimulated research centers in the investigation for therapeutic targets capable of containing the hyperactivity of the host immune system.^{5,6} The intervention immune is based on the pathophysiology of sepsis that causes the stimulation of intracellular signaling pathways leading to the gene expression of pro and anti-inflammatory, including TNF- α , IL-6, IL-10, and MCP-1.^{7,8} It has been proposed that the therapeutic management using immunomodulators is effective in infection because of its importance in the activation of cell populations and in the stimulation or suppression of immune mediators.^{9,10}

Propionibacterium acne, formerly known as Corynebacterium parvum is a Gram-positive anaerobic micro-organism belonging to the skin human, being widely used in clinical trials as suitable candidate for therapeutic approach. The bacillus has been used as a tool for studies of cellular immune response due to its immunomodulatory effects.¹¹ The inactivated bacteria promotes Th1¹² and Th2¹³ response in mice, induces tumoricidal activity,¹⁴ increases resistance to viral infections,¹⁵ parasitic,^{16,17} has anti-bacterial activity¹⁸ and has an important relationship with the activation of cell populations.^{19,20} One of the most important adjuvant effects of *P. acnes* is the increase in macrophage phagocytic activity, which can explain, in part, by ability to enhance animal resistance to several pathogens.²⁰

The immunomodulator *P. acnes* has been studied by several researchers, both in human and animal models of infectious diseases.^{21–24} *P. acnes* action was evaluated in mice infected by the rabies virus and researchers observed greater survival rates in animals treated with the immunomodulator and antirabies vaccine. This greater survival was not related to serum neutralizing antibodies, interferon gamma concentration and cellular immunity evaluated by the macrophage inhibition test, but it was correlated with higher Natural Killer activity in infected mice treated using *P. acnes*.¹⁵

Although there are studies describing the immunomodulatory and antibacterial properties of *P. acnes*, we found no reports of its effect on polymicrobial sepsis. Thus, the objective of this study was to investigate the effects of *P. acnes*-Killed in lethal sepsis induced by cecal ligation and puncture (CLP).

Methods

Experimental animals

Male mice (Mus musculus) provided by the animal facilities of Federal University of Pernambuco - UFPE, Recife, Brazil, were used for the evaluation of the P. acnes-killed in sepsis model. The mice used weighed between 20 and 25 g and were kept in a room with controlled temperature $(22 \pm 2 \degree C)$, humidity (50–60%) and a 12 h/12 h light/dark cycle. Water and food were made available to the animals without restriction. Before beginning the experiments, animals were acclimated to the laboratory environment for at least 30 min. The Animal Studies Committee of the Federal University of Pernambuco approved the experimental protocols (number 23076.039440/2011-30). The animals were treated according to the ethical principles of animal experimentation of SBCAL (Brazilian Society of Laboratory Animal Science) and the norms of the National Institute of Health Guide for Care and Use of Laboratory Animals.

Drugs and reagents

P. acnes-killed was produced by Laboratório Farmacêutico do Estado de Pernambuco (LAFEPE), Brazil, constituted of 4 mg/2 mL of P. acnes-killed (marketed by name Imunoparvum[®]), dose used was 0.2 mL (0.4 mg)/animal. Other drugs and reagents used in this study were as follows: Agar Mueller Hinton (Difco Laboratories, USA). TNF- α , IL-6, IL-10, and MCP-1 kits were purchased from eBioscience, San Diego, California, USA.

Experimental design

Polymicrobial sepsis was induced using CLP according to previously described method.²⁵ After deep anesthesia, midline laparotomy was performed with exposure of cecum followed of ligature and one transverse perforation of the same, with a 18 G needle, for induction of sepsis lethal. After surgery, the cecum of the animals was replaced into peritoneal cavity, and it was closed in two layers with a 4.0 nylon suture. Immediately after surgery, each animal received a subcutaneous injection of a 1 mL of saline solution as fluid resuscitation.

The animals were initially divided into 3 groups (n = 16) that were treated by intramuscular route in 1, 3, 5, and 7 days before the CLP induction. The control group animals (G-1) received vehicle (saline solution 0.9%) and the animals of the group 2 (G-2) received the *P. acnes*-killed constituted with 4 mg/2 mL of *P. acnes*-Killed at a dose of 0.2 mL (0.4 mg/animal). In the sham-operated group (G-3), underwent identical laparotomy but without cecum puncture and the mice were not treated. Six hours after CLP, half of the animals was euthanized to assess specific parameters (leukocytes count, determination of cytokines and bacteria counts in peritoneal fluid). The other half of each group was used for evaluation of the survival rate.

Cellular migration

Six hours after CLP, eight animals of each group were euthanized and the cells present in the peritoneal cavity were harvested by introducing 3 mL of phosphate-buffered saline (PBS) containing 1 μ M of EDTA. Total white blood cells (WBC) count was performed by using an automatic counter (ABX Micros 60), and differential cell counts were carried out on cytocentrifuge slides (Cytospin 3; Shandon Southern Products, Astmoore, United Kingdom), stained by the panotic dye and analyzed by optical microscopy. The results were expressed as the number of cells per cavity.

Determination of cytokines in peritoneal lavage

The exudates collected from the peritoneal cavity were centrifuged and the supernatant stored in freezer at -20 °C for the determination of cytokine levels. Quantification of the TNF- α , IL-6, IL-10, and MCP-1 levels in these exudates was performed by sandwich ELISA using kits that were specific for mice according to the manufacturer's instructions (eBioscience, San Diego, California, USA).

Determination of bacterial counts

Bacterial counts were performed on aseptically obtained peritoneal fluid. Six hours after CLP, mice were euthanized and the skin of abdomen was cut open in the midline without injury to the muscle. Samples were serially diluted from 1:100 to 1:1000 and 1:10,000 in PBS and cultured on Agar Mueller Hinton (Difco Laboratories) plates. Colony-forming units were counted after 18–24 h incubating 32 °C and the results were expressed as log¹⁰ of the number of colony-forming units/mL peritoneal fluid.

Statistical analysis

The results were expressed as the mean \pm standard deviation. Statistical analyses were performed using the Graph Pad software version 5.0 (GraphPad Software Inc., San Diego, CA, USA). We used a one-way ANOVA followed by a Bonferroni's test, with a significance level of 0.05%. The survival of mice was evaluated using curve-Mantel Cox and log-rank tests to compare the curves. The t-test was used for the comparison of data obtained for the enumeration of bacteria in the peritoneal cavity, with a significance level of 0.05%.

Results

Effect of P. acnes-killed on the survival of animals subjected to lethal sepsis by CLP

As shown in Fig. 1, CLP induced the death of 100% of control animals 36 h after surgery. The reduction in mortality was significant in the animals receiving treatment with *P. acnes*-killed. The rate of the treated group was 50% at 96 h after CLP.

Effect of treatment with P. acnes-killed in cell migration into the peritoneal cavity after inducing lethal sepsis by CLP

A significant increase in the total number of leukocytes were observed in the peritoneal cavity of the group treated with P.

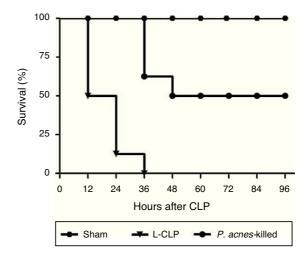


Fig. 1 – Effect of *P. acnes*-killed on survival rate of cecal ligation and puncture (CLP) rats (n = 8). The data are expressed as the cumulative percentage of rats still alive within each interval and the survival each 6 h was recorded. Compared with CLP group, *P. acnes*-killed improved survival within 96 h. All animals survived in the sham group. p < 0.05 vs CLP group.

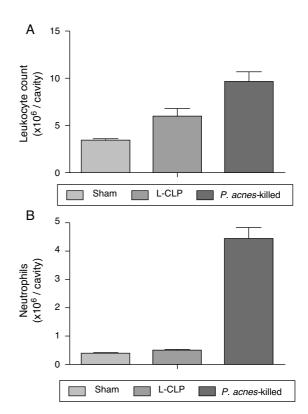


Fig. 2 – Total (A) and differential (B) cell counting in the peritoneal cavity. The total and differential counting of peritoneal cells were performed 6 h after the cecal ligation and puncture (CLP) (A). The results were expressed as mean \pm S.E.M. (7 animals/group). *p < 0.05 when compared to the control group.

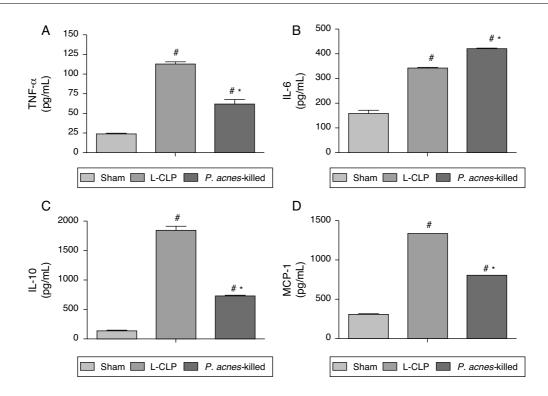


Fig. 3 – Effect of P. acnes-killed treatment in animals (n = 6) in the expression of TNF- α (A), IL-6 (B), IL-10 (C) and MCP-1 (D) levels in the peritoneal cavity of mice subjected to cecal ligation and puncture (CLP). The cytokine levels in peritoneal exudates were determined at 6 h after surgery in sham-operated, L-CLP and P. acnes-killed mice. Results are expressed as mean \pm S.E.M. *p < 0.05 compared with L-CLP.

acnes-killed (9.64 \pm 2.7 \times 10⁶ leukocytes/cavity), in the control group (6.00 \pm 2.0 \times 10⁶ leukocytes/cavity) (Fig. 2). The recruitment of neutrophils into the peritoneal cavity increased in the group of animals that received *P. acnes*-killed compared with the control group.

Effect of P. acnes-killed levels of TNF- α , IL-6, IL-10, and MCP-1

Fig. 3 shows the concentrations of TNF- α , IL-6, IL-10, and MCP-1 in the peritoneal exudate of the groups treated with *P. acnes*-killed, sham, and control group 6 h after surgery. The levels of TNF- α , IL-10, and MCP-1 showed a decreased in animals treated with *P. acnes* when compared with the control group. In contrast, IL-6 increased through treatment with *P. acnes*-killed displayed high concentration 6 h after CLP.

Effect of bacterial count in the peritoneal cavity of animals subjected to CLP and treated with P. acnes-killed

To evaluate the effect of *P. acnes*-killed in control of infection, we investigated the number of bacteria in the peritoneal cavity of animals subjected to lethal sepsis by CLP. We have detected that 6 h after surgery the animals in the control group showed deficiency in control the infection, which was evidenced by the high concentration of colony-forming units (CFU) of the abdominal cavity. The number of bacteria found in the peritoneal cavity of the treated group was lower than the values found in control animals. The sham group had no CFU 24 h after surgery (Fig. 4).

Discussion

This study demonstrated that the protective action of *P. acnes*-killed modulates the inflammatory response in animals subjected to lethal sepsis by CLP. When the animals received

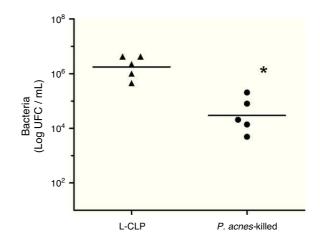


Fig. 4 – Bacterial counts in the peritoneal fluid of L-cecal ligation and puncture (CLP) and *Imunoparvum*[®] groups (n = 5). Quantification of the amount of bacteria in the peritoneal cavity was performed 6 h CLP surgery. The number of bacteria present in the peritoneal cavity is expressed as mean log CFU/mL. **p < 0.05 compared with L-CLP.

an intramuscular injection of *P. acnes*-killed and underwent severe sepsis there was a reduction in mortality, conferring resistance to infection. In this study we have observed a survival rate of 100% in the first 24 h after surgery in the treated group. CLP is a murine model of polymicrobial sepsis similar to the progression and characteristics of human sepsis that has contributed to the knowledge of the involvement of components of innate immunity, including the identification of new potential therapeutic targets.²⁶ The sensitivity of animals treated with *P. acnes* is not restricted to Gram-negative bacteria by LPS, but also to Gram-positive bacteria such as *Listeria monocytogenes*.²⁷ The data found in our study are in agreement with other authors who reported the antibacterial activity of *P. acnes* against Gram-positive and Gram-negative bacteria.^{28,29}

In the sepsis, a failure of neutrophil migration to the infectious site has been found and is associated with a large deficit in the control of infection, leading to increased bacterial dissemination and inducing a high mortality rate.³⁰ The neutrophil migration to the site of infection is extremely important for the control of bacterial growth and consequently for the prevention of bacterial dissemination.³¹ This study demonstrated that the increase in animal survival was accompanied by an increase in migration of cells into the peritoneal cavity and significant decreased in colony forming units of the group treated with P. acnes-killed. Studies have shown that P. acnes has worked in the modulation of macrophages, eosinophils,¹³ neutrophils,³² dendritic cells,³³ and natural killer cells.³⁴ Our data corroborate the findings of other authors that observed the recruitment of neutrophils into the peritoneal cavity when animals were stimulated with P. acnes-killed.³⁵

The expression of inflammatory mediators is the host response to infection. In sepsis, the over expression of proinflammatory cytokines have deleterious effects on the body, disrupt the immune system and is involved in multiple organ dysfunction.³⁶ TNF- α is one of the mediators responsible for organ dysfunction and increased mortality during severe sepsis.³⁷ Some studies have reported that increasing the survival rate is following the decrease of $TNF-\alpha$.^{5,6} Thus, immunotherapy focus of sepsis in blocking TNF- α is promising in animal models. This study assessed the effect of P. acnes-killed on the concentration of TNF- α in the peritoneal cavity 6 h after CLP and there was a significant reduction in levels of this marker. Similar results were found in peritoneal fluid by Pang et al.³⁸ that evaluated the modulating effect of the methylene chloride in animals with sepsis and reported a decrease in TNF- α and consequent increase in survival. Another study found that the decrease in concentration of TNF- α in serum and peritoneal exudate of animals treated with myrrh, an essential oil obtained from species of the genus Commiphora, increased survival in severe sepsis.³⁹ Recently it was shown the immunomodulatory activity of a polysaccharide of P. acnes on reducing TNF- α in animals with focal segmental glomerulosclerosis.40

The monocyte chemotactic protein (MCP-1) is a CC chemokine belonging to the subfamily that stimulates the migration of monocytes cells and is associated with protection in experimental sepsis with activation of neutrophils and macrophages.⁴¹ The coordinated synthesis and release of

MCP-1 is important in acute and chronic inflammatory process by controlling the influx of phagocytic cells and also activation of the release of inflammatory cytokines.⁴² In our study there was a decrease in levels of MCP-1 in animals treated with *P. acnes*-killed and subjected to severe sepsis. Previous study shown that blocking the activity of MCP-1 in animals sensitized with *P. acnes* inactivated decreased of the concentrations of TNF- α and IL- β .⁴³ Recently, anti-MCP-1 strategies have been suggested as a potential therapeutic for the treatment of sepsis and endotoxemia by preventing the interaction between MCP-1 and other inflammatory mediators and oxidative stress in organs.⁴⁴

In this study, the data showed no decrease in the concentration of IL-6 following treatment with P. acnes-killed. The studies that evaluated the effects of P. acnes on the concentration of IL-6 are controversial. The increased IL-6 levels after stimulation with P. acnes has been reported,⁴⁵ moreover, P. acnes also decreases IL-6 concentrations.46 Mice deficient for IL-6 showed no improvement in survival.47 In addition to administration of high doses of anti-IL-6 or lower doses of antibody did not increase survival of animals with sepsis. This suggests that excessive levels of IL-6 are detrimental and that the lock does not help the survival of these animals.48 Although P. acnes modulates the expression of IL-6 increasing its concentration in this experimental model, we believe that the presence of a balanced concentration of IL-6 in acute inflammation may be required for an effective immune response in innate immunity. Therefore, further analyses are needed to clarify this issue.

Notably, pre-treatment of P. acnes-killed suppressed the production of IL-10 in the peritoneal cavity. The cytokine IL-10 appears to be crucial in regulating the production of proinflammatory cytokines.⁴⁹ In a previous study, administration of IL-10 reduced the levels of TNF- α and increased survival of endotoxemic animals.⁵⁰ However, this was not the case of our study, because the treatment with P. acnes-killed reduced IL-10, but also decreased TNF- α and MCP-1. The role of IL-10 in polymicrobial sepsis or its interaction with pro-inflammatory cytokines may therefore be more complex. The decreased expression of IL-10 is associated with increased survival by restoring the host immune response⁵¹ and the decrease of CFUs in animals subjected to CLP.⁵² Recently it was suggested that neutralization of IL-10 decreases the percentage of Treg CD4+ T cells and increases the survival of septic animals.53 Thus, it is suggested that the reduced levels of IL-10 in this study was beneficial in protecting against polymicrobial sepsis.

Finally the data obtained in this study show that the beneficial effect of treatment with *P. acnes*-killed in the survival rate of animals was accompanied by reduced levels of TNF- α , MCP-1, and IL-10 and increased IL-6, contributing to leukocyte recruitment and reduced bacterial growth. These data suggest that the improvement of host defense in the initial focus of infection can weaken the inflammatory response in other places.

Conflict of interest

The authors have no conflict of interest.

Acknowledgments

The authors acknowledge financial support by CAPES and CNPQ.

REFERENCES

- Huttunen R, Aittoniemi J. New concepts in the pathogenesis, diagnosis and treatment of bacteremia and sepsis. J Infect. 2011;63:407–19.
- Angus DC, Linde-Zwirble WT, Lidicker J, et al. Epidemiology of severe sepsis in the United States: analysis of incidence, outcome, and associated costs of care. Crit Care Med. 2001;29:1303–10.
- Silva E, Pedro MA, Sogayar AC, et al. Brazilian Sepsis Epidemilogical Study (BASES study). Crit Care. 2004;8:R251–60.
- 4. Chalupka AN, Talmor D. The economics of sepsis. Crit Care Clin. 2012;28:57–76.
- Ma H, Kou J, Zhu D, et al. Liu-Shen-Wan, a traditional Chinese medicine, imrpoves survival in sepsis induced by cecal ligation and puncture via reducing TNF-alpha level, MDA content and enhancing macrophage phagocytosis. Int Immunopharmacol. 2006;6:1355–62.
- Maciel MCG, Farias JC, Maluf MJ, et al. Syzygium jambolanum treatment improve survival in lethal sepsis induced in mice. BMC Complement Altern Med. 2008;8:57–64.
- 7. Hotchkiss RS, Karl IE. The pathophysiology and treatment of sepsis. N Engl J Med. 2003;348:138–50.
- Giamarellos-Bourboulis EJ. Immunomodulatory therapies for sepsis: unexpected effects with macrolides. Int J Antimicrob Agents. 2008;32S:S39–43.
- 9. O'callaghan A, Redmond HP. Treatment of sepsis: current status of clinical immunotherapy. Surgeon. 2006;4:355–61.
- Opal SM. News perspectives on immunomodulatory therapy for bacteraemia and sepsis. Int J Antimicrob Agents. 2012;36S:S70–3.
- Perry AL, Lambert PA. Propionibacterium acnes. Lett Appl Microbiol. 2006;42:185–8.
- 12. Cervi L, MacDonald AS, Kane C, et al. Cutting edge: dendritic cells copulsed with microbial and helminth antigens undergo modified maturation, segregate the antigens to distinct intracellular compartments, and concurrently induce microbe-specific Th1 and helminth-specific Th2 responses. J Immunol. 2004;172:2016–20.
- Braga EG, Ananias RZ, Mussalem JS, et al. Treatment with Propionibacterium acnes modulates the late phase reaction of immediate hypersensitivity in mice. Immunol Lett. 2003;88:163–9.
- Tsuda K, Yamanaka K, Linan W, et al. Intratumoral injection of Propionibacterium acnes suppresses malignant melanoma by enhancing Th1 immune responses. PLoS ONE. 2011;6:e29020.
- Megid J, Peraçolli MTS, Curi PR, et al. Effect of vaccination and the immunomodulators "bacillus of Calmette" – Guérin, Avridine and Propionibacterium acnes on rabies in mice. Comp Immunol Microbiol Infect Dis. 1998;4:305–18.
- Iseki H, Takabatake N, Ota N, et al. Babesia: the protective effects of killed *Propionibacterium acnes* on the infections of two rodent Babesia parasites in mice. Exp Parasitol. 2008;118:543–8.
- Abel LCJ, Chen S, Ricca LG, et al. Adjuvant effect of LPS and killed Propionibacterium acnes on development of experimental gastrointestinal nematode infestation in sheep. Parasite Immunol. 2009;31:604–12.
- Ha DKK, Lawton JWM, Gardner ID. The effect of in vitro modulation of macrophage activation on Mycobacterium lepraemurinum infection. J Comp Pathol. 1986;96:565–73.

- MacDonald AS, Straw AD, Dalton NM, Pearce EJ. Cutting edge: Th2 response induction by dendritic cells: a role for CD40. J Immunol. 2002;168:537–40.
- 20. Ananias RZ, Rodrigues EG, Braga EG, et al. Modulatory effect of killed Propionibacterium acnes and its purified soluble polysaccharide on peritoneal exudate cells from C57Bl/6 mice: major NKT cell recruitment and increased cytotoxicity. Scand J Immunol. 2007;65:538–48.
- Al-Izzi SA, Maxie MG. Effect of Corynebacterium parvum on bone marrow macrophage colony production, peripheral blood leucocytes, and histologic changes of tissues in calves. Am J Vet Res. 1982;43:2244–7.
- 22. Sodja I. Experimental chemotherapy of rabies: preliminary results. Acta Virol. 1986;30:63–8.
- Lopes CAC, Santana CF. Novas observações sobre o emprego do Corynebacterium parvum em pacientes portadores de doenças infecciosas. Pernambuco: Ministério da Educação e Cultura, Universidade Federal de Pernambuco; 1984, 15 p.
- 24. Cox WI. Examining the immunologic and hematopoietic properties of an immunostimulant. Vet Med. 1988;83:424–8.
- Rittirsch D, Huber-Lang MS, Flierl MA, Ward PA. Immunodesign of experimental sepsis by cecal ligation and puncture. Nat Protoc. 2009;4:31–6.
- Dejager L, Pinheiro I, Dejonckheere E, Libert C. Cecal ligation and puncture: the gold standard model for polymicrobial sepsis? Trends Microbiol. 2011;19:198–208.
- Merlin T, Gumenscheimer M, Galanos C, Freudenberg A. TNF-α hyper-responses to gram-negative and gram-positive bacteria in Propionibacterium acnes-primed or Salmonella typhimurium infected mice. J Endotoxin Res. 2001;7:157–63.
- Collins FM, Scott MT. Effect of Corynebacterium parvum treatment on the growth of Salmonella enteritidis in mice. Infect Immun. 1974;9:863–9.
- 29. Mochizuki H, Nomura T, Kawamura I, Mitsuyama. Enhanced resistance to gram-positive bacterium and increased susceptibility to bacterial endotoxin in mice sensitized with *Propionibacterium acnes*: involvement of Toll-like receptor. FEMS Immunol Med Microbiol. 2005;43:287–93.
- Benjamim CF, Ferreira SH, Cunha FQ. Role of oxide in the failure of neutrophil migration in sepsis. J Infect Dis. 2000;182:214–23.
- Benjamim CF, Silva JS, Fortes ZB, et al. Inhibition of leukocyte rolling by nitric oxide during sepsis leads to reduced migration of active microbicidal neutrophils. Infect Immun. 2002;70:3602–10.
- Lee W-L, Shalita AR, Suntharalingam K, et al. Neutrophil chemotaxis by Propionibacterium acnes lipase and its inhibition. Infect Immun. 1982;35:71–8.
- 33. Zhang Y, Yoneyama H, Wang Y, et al. Mobilization of dendritic cell precursors into the circulation by administration of MIP $1-\alpha$ in mice. J Natl Cancer Inst. 2004;96:201–9.
- Davis EG, Rush BR, Blecha F. Increases in cytokine and antimicrobial peptide gene expression in horses by immunomodulation with Propionibacterium acnes. Vet Ther. 2003;4:5–11.
- Vinson RB, Carroll JL, Pruett SB. Mechanism of suppressed neutrophil mobilization in a mouse model for binge drinking: role of glucorticoids. Am J Physiol Regul Integr Comp Physiol. 1998;275:R1049–57.
- 36. Tavares-Murta BM, Cunha FQ, Ferreira SH. The intravenous administration of tumor necrosis factor, interleukin 8 and macrophage-neutrophil derived chemotact factor inhibits neutrophil migration by stimulating nitric oxide production. Br J Pharmacol. 1998;24:1369.
- Reddy ABM, Srivastava SK, Ramana KV. Anti-inflammatory effect of aldose reductase inhibition in murine polymicrobial sepsis. Cytokine. 2009;48:170–6.

- Pang Q, Dou L, Pan X, et al. Methylene chloride protects against cecal ligation and puncture-induced acute lung injury by modulating inflammatory mediators. Int Immunopharm. 2010;10:929–32.
- 39. Kim MS, Bae GS, Park KC, et al. Myrrh inhibits LPS-induced inflammatory response and protects from cecal ligation and puncture-induced sepsis. eCAM. 2012;2012:278718.
- Reis VO, Silva JC, Soua GT, et al. The polysaccharide fraction of Propionibacterium acnes modulates the development of experimental focal segmental glomerulosclerosis. Immunobiology. doi:10.1016/j.imbio.2011.12.003, in press.
- Matsukawa A, Hogaboam CM, Lukacs NW, et al. Endogenous monocyte chemoattractant protein-1 (MCP-1) protects mice in a model of acute septic peritonitis: cross-talk between MCP-1 and leukotriene B4. J Immunol. 1999;163:6148–54.
- 42. Gomes RN, Figueiredo RT, Bozza FA, et al. Increased susceptibility to septic and endotoxic shock in monocyte chemoattractant protein 1/CC chemokine ligand 2-deficient mice correlates with reduced interleukin 10 and enhanced macrophage migration inhibitory factor production. Shock. 2006;26:457–63.
- Ichiyasu H, Suga M, Matsukawa A, et al. Functional roles of MCP-1 in Propionibacterium acnes-induced, T cell-mediated pulmonary granulomatosis in rabbits. J Leukoc Biol. 1999;65:482–91.
- Ramnath RD, Ng SW, Guglielmotti A, et al. Role of MCP-1 in endotoxemia and sepsis. Int Immunopharmacol. 2008;8:810–8.
- 45. Akaza N, Akamatsu H, Kishi M, et al. Effects of Propionibacterium acnes on various mRNA expression levels in normal human epidermal keratinocytes in vitro. J Dermatol. 2009;36:213–23.

- 46. Megid J, Kaneno R, Nozaki CN, et al. Increased interleukin-10 associated with low IL-6 concentration correlated with greater survival rates in mice infected by rabies virus vaccinated against it and immunomodulated with *P. acnes*. Comp Immunol Microbiol Infect Dis. 2004;27: 393–411.
- Leon LR, White AA, Kluger MJ. Role of IL-6 and TNF in thermoregulation and survival during sepsis in mice. Am J Physiol. 1998;275:R269–77.
- Riedemann NC, Guo RF, Ward PA. Novel strategies for the treatment of sepsis. Nat Med. 2003;9:517–24.
- Moore KW, Malefyt RW, Coffman RL, et al. Interleukin-10 and the Interleukin-10 receptor. Annu Rev Immunol. 2001;19:683–765.
- 50. Smith SR, Terminelli C, Denhardt G, et al. Administration of InterleukiN-10 at the time of priming protects *Corynebacterium parvum*-primed mice against LPS- and TNF-α-induced lethality. Cell Immunol. 1996;173:207–14.
- 51. Hiraki S, Ono S, Kinoshita M, et al. Neutralization of IL-10 restores the downregulation of IL-18 receptor on natural killer cells and interferon- γ production in septic mice, thus leading to an improved survival. Shock. 2012;37:177–82.
- 52. Newsome CT, Flores E, Ayala A, et al. Improved antimicrobial host defense in mice following poly-(1,6)-β-D-glucopyranosyl-(1,3)-β-D-glucopyranose glucan treatment by a gender-dependent immune mechanism. Clin Vaccine Immunol. 2011;18:2043–9.
- 53. Hiraki S, Ono S, Tsujimoto H, et al. Neutralization of interleukin-10 or transforming growth factor-β-decreases the percentages of CD4⁺ CD25⁺ Foxp3⁺ regulatory T cells in septic mice, thereby leading to an improved survival. Surgery. 2012;15:313–22.