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Case report

Bacteremia due to *Moraxella osloensis*: a case report and literature review

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ABSTRACT

Herein we report the case of a 10-year-old boy with an autosomal mosaic mutation who developed bacteremia. The causative agent was identified as *Moraxella osloensis* by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry and 16S rRNA gene sequencing. In the pediatric population, there have been 13 case reports of infection attributed to *M. osloensis* and this is the fifth reported case of pediatric bacteremia due to *M. osloensis*. After *Moraxella* species infection was confirmed, the patient recovered with appropriate antimicrobial therapy. It is important to consider that *M. osloensis* can cause serious infections, such as bacteremia, in otherwise healthy children.

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Introduction

Moraxella osloensis is an aerobic, Gram negative coccobacillus. It can be isolated from healthy human respiratory tracts, but has been reported as a rare pathogen in immunocompromised individuals, like patients with cancer, leukemia, and organ transplant recipients.¹ However, it is not well known as a pediatric pathogen. Here, we report a pediatric case of bacteremia due to *M. osloensis*, with a partial review of the literature.

Case report

The patient was 10-year-old boy with an autosomal mosaic mutation [46XY,t(9;15)(p22;p13),46XY,der(9)t(9;15)(p22;p13)]. He had mild intellectual disability and a substantial history of infectious diseases, including pneumonia, otitis media, sinusitis, and urinary tract infection due to *Escherichia coli*. He received prophylactic trimethoprim-sulfamethoxazole until six years of age. His immune function was normal, including number of neutrophils, immunoglobulins, and complement. In recent years, he had been doing well without medication.

The patient visited Kofu Municipal Hospital with a 10-day history of fever, cough, and sore throat, and he had been

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Table 1 – Clinical characteristics of *Moraxella osloensis* bacteremia in children.

Reference	Age(y)	Clinical History	Clinical manifestation(s)			
			Sex	Without bacteremia	Approach for identification	Treatment
Butzler et al. ⁶	2/M	None		Stomatitis, Impetigo	Laboratory culture	Ampicillin
Shah et al. ¹	2/M	None		Reactive airway disease	Laboratory culture	Cefloxiime, ST
Dien Bard et al. ⁷	3/M	Cortical dysplasia and developmental delay		Prolonged hypotension	16S rRNA	Piperacillin/tazobactam
Minami et al. ⁸	9/M	Cerebral palsy		Possible cholecystitis	16S rRNA	Cefmetazole
Present patient	10/M	Mild intellectual disability		Prolonged fever	MALDI-TOF MS, 16S rRNA	Meropenem, ceftriaxone

M, male; ST, trimethoprim-sulfamethoxazole; 16S rRNA, 16S rRNA gene sequencing; MALDI-TOF MS, matrix-assisted laser desorption/ionization time-of-flight mass spectrometry.

treated with oral cephem antibiotics for a few days before presentation. On physical examination, his temperature was 40 °C, and his throat was found to be infected. Laboratory testing revealed the following: white blood cell count 6400/µL (77% neutrophils, 17% lymphocytes, and 15% monocytes), hemoglobin 14.0 g/dL, platelet count $19.9 \times 10^4/\mu\text{L}$, C-reactive protein level 1.8 mg/dL, IgG 1050 mg/dL (IgG1 600 mg/dL, IgG2 326 mg/dL, IgG3 30.6 mg/dL, and IgG4 < 3.0 mg/dL), IgA 101 mg/dL, and IgM 71 mg/dL. Urinalysis was normal and rapid influenza A and B antigen tests were negative. Chest radiography and abdominal ultrasonography findings were normal. He received ambulatory care for acute pharyngitis after blood and a throat swab for culture were obtained. Two days later, a Gram negative coccobacillus was isolated from blood culture. The patient, who still had fever and fatigue, was admitted to the hospital for Gram negative bacteremia. Intravenous meropenem (100 mg/kg/day) was administered as an empirical treatment. The patient became afebrile after treatment, and his other symptoms improved. On day 2, *Moraxella* species were isolated from blood culture, but precise identification was not obtained with our BD Phoenix system (BD Japan, Tokyo, Japan). The isolate was determined to be susceptible to ampicillin, piperacillin, cefazolin, cefotaxime, gentamicin, levofloxacin, meropenem, and trimethoprim-sulfamethoxazole. However, a cefinase test found the isolate positive for β-lactamase. We switched to ceftriaxone (120 mg/kg/day) until discharge on day 10 when a repeated blood culture was sterile. The throat culture did not yield *Moraxella* species.

The organism in this case was finally identified as *M. osloensis* by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS; Bruker Biotype). Its identity was confirmed by 16S rRNA gene sequencing. Namely, 16S rRNA gene fragments were amplified by PCR using universal primers 5'-AGAGTTTGATCCTGGCTCAG-3' and 5'-CTTGTGCGGCCGCCGTCAATT-3' (Y is C or T and H is A, C, or T) and 55 °C annealing temperature.² PCR amplicon was purified with a Wizard SV Gel and PCR Clean Up system (Promega, Madison, WI, USA), and sequenced directly on both strands with a BigDye Terminator v1.1 Cycle Sequencing Kit and an ABI 3730xl genetic analyzer (Applied Biosystems, Foster City, CA, USA). The 1308-bp nucleotide sequence obtained had 100% sequence identity to the corresponding sequence of *M. osloensis* (accession no. AB931117) by analyzing with Ribosomal Database Project II (release 11, update 5) (<http://rdp.cme.msu.edu/index.jsp>).

Discussion

M. osloensis was first described by Bovre in 1947 and has been isolated in various environments, including hospitals. In the pediatric population, there have been 13 case reports of infection attributed to *M. osloensis* including four cases of bacteremia (Table 1). This is the fifth reported case of pediatric bacteremia due to *M. osloensis*, to the best of our knowledge. In contrast to older patients, the majority of these infections were found in patients that did not have underlying medical conditions.

The appropriate treatment for invasive *M. osloensis* infection has not been studied. Most reported isolates have been susceptible to penicillin and cephalosporins, but penicillin-resistant strains of *M. osloensis* (minimum inhibitory concentration of 6.25 µg/mL) have been reported.³ Among *Moraxella* species, *M. catarrhalis*, *M. lacunata*, and *M. nonliquefaciens* are known to produce BRO β-lactamase, which degrades penicillin and a part of first-generation cephalosporin.⁴ It remains to be clarified whether or not *M. osloensis* produces BRO β-lactamase. We chose ceftriaxone as a definitive therapy because of its performance against BRO β-lactamase and effectiveness against other *Moraxella* species.

M. osloensis is difficult to identify because of the presence of several other species with similar phenotypic characteristics.⁵ MALDI-TOF MS is a tool for rapid, accurate, and cost-effective identification of cultured bacteria and fungi based on automated analysis of the mass distribution of bacterial proteins. The organism in this case was identified by this method and confirmed furthermore by 16SrRNA sequence analysis, which is the most reliable method for species-level identification.

Herein we report a rare pediatric case of bacteremia caused by *M. osloensis*. The patient recovered with antimicrobial therapy, but it is important to consider that *M. osloensis* can cause serious infections, such as bacteremia, in otherwise healthy children.

Conflicts of interest

The authors declare no conflicts of interest.

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REFERENCES

1. Shah SS, Ruth A, Coffin SE. Infection due to *Moraxella osloensis*: case report and review of the literature. *Clin Infect Dis*. 2000;30:179-81.
2. Héritier C, Poirel L, Aubert D, Nordmann P. Genetic and functional analysis of the chromosome-encoded carbapenem-hydrolyzing oxacillinase OXA-40 of *Acinetobacter baumannii*. *Antimicrob Agents Chemother*. 2003;47:268-73.
3. Hansen W, Butzler JP, Fuglesang JE, Henriksen SD. Isolation of penicillin and streptomycin resistant strains of *Moraxella osloensis*. *Acta Pathol Microbiol Scand B Microbiol Immunol*. 1974;82:318-22.
4. Wallace RJ Jr, Steingrube VA, Nash DR, et al. Beta-lactamases of *Branhamella catarrhalis* and *Moraxella* subgenus *Moraxella*, including evidence for chromosomal beta-lactamase transfer by conjugation in *B. catarrhalis*, *M. nonliquefaciens*, and *M. lacunata*. *Antimicrob Agents Chemother*. 1989;33:1845-54.
5. Roh KH, Kim CK, Koh E, et al. Three cases of *Moraxella osloensis* meningitis: a difficult experience in species identification and determination of clinical significance. *J Korean Med Sci*. 2010;25:501-4.
6. Butzler JP, Hansen W, Cadanel S, Henriksen SD. Stomatitis with septicemia due to *Moraxella osloensis*. *J Pediatr*. 1974;84:721-2.
7. Dien Bard J, Lewinski M, Summanen PH, Deville JG. Sepsis with prolonged hypotension due to *Moraxella osloensis* in a non-immunocompromised child. *J Med Microbiol*. 2011;60 Pt 1:138-41.
8. Minami K, Higuchi T, Cho Y, et al. A pediatric case of bacteremia and possible cholecystitis due to *Moraxella osloensis*. *Jpn J Infect Dis*. 2015;68:324-5.