



The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Brief communication

Community-acquired methicillin-resistant *Staphylococcus aureus* carrying SCCmec type IV and V isolated from healthy children attending public daycares in northeastern Brazil



Suzi P. de Carvalho^{a,b}, Jéssica B. de Almeida^{a,b}, Yasmin M.F.S. Andrade^b, Lucas S.C. da Silva^b, Arianne C. de Oliveira^b, Flávia S. Nascimento^b, Guilherme B. Campos^c, Márcio V. Oliveira^b, Jorge Timenetsky^c, Lucas M. Marques^{a,b,c,*}

^a Universidade Estadual de Santa Cruz, Departamento de Microbiologia, Ilhéus, BA, Brazil

^b Universidade Federal da Bahia, Instituto Multidisciplinar em Saúde, Núcleo de Tecnologia em Saúde, Vitória da Conquista, BA, Brazil

^c Universidade de São Paulo, Instituto de Ciências Biomédicas, Departamento de Microbiologia, São Paulo, SP, Brazil

ARTICLE INFO

Article history:

Received 24 August 2016

Accepted 17 April 2017

Available online 5 May 2017

Keywords:

CA-MRSA

Antibiotic resistance

SCCmec

MLST

ABSTRACT

Nasal colonization with methicillin-resistant *Staphylococcus aureus* (MRSA) have increasingly been reported in healthy communities. This study aimed to assess the rate of *S. aureus* in general and MRSA in particular from nasal secretion of children in daycare centers in Vitória da Conquista, Brazil. The isolates were identified based on morphology, biochemical tests and by PCR. Detection of virulence genes, biofilm production, and susceptibility test by disk diffusion agar were performed. MRSA isolates were characterized by spa, SCCmec, and multilocus sequence typing (MLST). *S. aureus* were recovered from 70 (47.3%) of 148 children. Among the 11 MRSA strains (15.7%), two SCCmec types (IV and V) were detected. MLST identified four STs related to three clonal complexes (CC): 5, 45, and 398. Four spa types were found circulating in this setting. Resistance of *S. aureus* isolates to ampicillin, erythromycin, ciprofloxacin, clindamycin, and tetracycline was 80%, 32.8%, 7.1%, 7.1% and 4.3%, respectively. One isolate presented intermediate resistance to vancomycin detected by Etest methodology. All strains were biofilm producers. The virulence genes *seb*, *sec*, *spa*, and *pvl* were detected in some isolates. This study revealed a high rate of children carrying MRSA among healthy attendees in daycare centers in Vitória da Conquista, Brazil.

© 2017 Sociedade Brasileira de Infectologia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

* Corresponding author.

E-mail address: lucasm@ufba.br (L.M. Marques).

<http://dx.doi.org/10.1016/j.bjid.2017.04.001>

1413-8670/© 2017 Sociedade Brasileira de Infectologia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Staphylococcus aureus may cause various infections with considerable morbidity and mortality in healthy and immunocompromised hosts.¹ Methicillin-resistant *Staphylococcus aureus* (MRSA) is commonly associated with severe nosocomial infections (HA-MRSA); however, it has been detected in individuals without risk factors for infection, referred to as community-acquired MRSA (CA-MRSA).² Methicillin resistance is carried on a staphylococcal cassette chromosome *mec* (SCCmec). HA-MRSA usually carry SCCmec types I, II, or III, whereas CA-MRSA strains commonly carry SCCmectypes IV and V.³

The molecular characterization is important, since it enables studying the relatedness of MRSA strains, their genetic diversity, and clonal distribution. Various molecular typing techniques have been developed, including the multi-locus sequence type (MLST), as well as the SCCmec typing, spa typing.^{3–5}

Nasal colonization of *S. aureus* is common in children, but the genetic findings suggested a causal relationship between carriers, chromosome cassettes of those harboring MRSA, and invasive staphylococcal disease.⁶ In addition, the incidence of pediatric infections due to CA-MRSA, including children with no identifiable risk factors, has increased worldwide.⁷ Daycare centers (DCCs) are reservoirs of *S. aureus* in general and MRSA in particular, and these children may spread these bacteria to the community and hospitals.⁶

In Brazil little is known about the distribution and characteristics of *S. aureus* and MRSA in children as nasal carriers, particularly in those attending DCCs. The aim of the present study was to assess the rate of *S. aureus* and MRSA nasal carriage in healthy children attending public DCCs in Vitória da Conquista – Bahia State (BA), Brazil, and to identify the resistance profile, genotypic characterization, and pathogenicity.

The study was conducted from October to December 2012, in four public DCCs in Vitória da Conquista, a city in Northeastern Brazil. One hundred and forty-eight samples of nasal swabs were obtained from healthy children ranging from one to six years attending DCCs. Children treated with antibiotics in the last 30 days were not included. The isolates obtained were incubated at 37 °C for 48 h; Gram-positive cocci colonies, positive for catalase and coagulase tests were selected as presumptive *S. aureus* and identified by PCR.

MRSA isolates and reduced vancomycin activity were screened for all *S. aureus* isolates by microdilution method following the recommendations of the CLSI guidelines.⁸ Strains of *S. aureus* ATCC 29213 and ATCC 43300 were used as controls. Vancomycin-resistant isolates were confirmed using the E-test® (bioMérieux's, Brazil) and the bacterial growth at ≥16 µg/mL was indicative of resistance. Susceptibility to erythromycin, tetracycline, clindamycin, ciprofloxacin, and ampicillin were performed by disk diffusion.⁸ Inducible resistance to clindamycin was tested by 'D test'.⁸ Multidrug resistance was considered when the strain was resistant to two or more antibiotics.

The genomic DNA of all strains were extracted using the Genomic DNA Purification Kit (Invitrogen, Carlsbad, CA, USA), and all isolates were characterized as SCCmec types (I–V),³ and for the presence of virulence genes, including *sea*, *seb*, *sec*, *pvl*, *clfA*, and *spa* by PCR.⁹ Moreover, MRSA isolates were subjected to MLST and SPA typing techniques, as described previously.^{4,5}

Biofilm assays were performed in 96-well polystyrene microplates using trypticase soy broth (TSB/Difco) with 1% (w/v) glucose (TSB-1% Glc).¹⁰ The samples were compared with cultures of *Streptococcus pyogenes* ATCC75194 (non-biofilm producer sample). *S. aureus* isolates were classified as non-biofilm producers, weak producers, moderate producers, producers, and strong producers. Statistical analysis was performed by chi-square test and a p-value lower than 0.05 was considered statistically significant.

Out of 148 children analyzed, *S. aureus* was recovered from 70 (47.3%). Eleven (15.7%) isolates were MRSA, the majority ($n=8$; 72.7%) were SCCmec IVA type and three (27.2%) were SCCmec V type.

Resistance rates of *S. aureus* isolates to ampicillin, erythromycin, clindamycin, ciprofloxacin, and tetracycline were 80%, 32.8%, 7.1%, 7.1%, and 4.3%, respectively. One methicillin-susceptible *S. aureus* (MSSA) isolate showed reduced susceptibility to vancomycin by E-test and was confirmed with a MIC of 6 µg/mL. There were no significant differences in resistance between MRSA and MSSA isolates. The rate of resistant isolates to more than two antibiotics was 47.1% (33/70), and only 15.7% (11/70) of the isolates were susceptible to the studied antibiotics. A total of 63.6% of MRSA isolates were multidrug resistant (Table 1).

MLST performed on MRSA isolates identified four STs, related to three clonal complexes (CC): 5 (63.5%), 45 (18.2%), and 398 (18.2%). Among the four STs identified, the ST5 clone (63.6%, 7/11) was the most prevalent (Table 2). Protein A was characterized in isolates of four spa types. The t242 (72.7%, 8/11) was dominant among all isolates (Table 1).

Two MRSA isolates (2.8%) had the *seb* gene, and all had *spa* genes. Two strains of MSSA (2.8%) showed the *sec* gene, three (4.2%) were positive for *pvl* gene, and all had *spa* (Table 1). All *S. aureus* isolates produced biofilm (Table 2) and there was no statistical difference in biofilm production between MRSA and MSSA isolates ($p>0.05$).

The detection of *S. aureus* (47.3%) and MRSA (15.7%) in nasal swabs of the studied children demonstrated a higher rate of these staphylococci than in studies conducted in other DCCs with rates ranging from 17% to 31.1% and 0.8 to 13.2%, respectively.^{7,11,12}

Although the prevalence of MRSA among healthy children was high, no correlation was found between carriage of MRSA and antibiotic use. In addition, the close contact among DCC attendees has also been considered a risk factor for carrying MRSA.⁶ A similar Brazilian study showed a MRSA prevalence of 1.2%¹² confirming the variations between regions. The high incidence of MRSA in healthy children is the most relevant and worrying finding for the susceptible hosts and even for the healthy carrier children if immunosuppressed. This context involves at least the risk for coagulase-positive staphylococcal infections in elderly hosts, hospitalized individuals, neonates, and the community. Furthermore, most CA-MRSA reported worldwide have been shown to carry SCCmec types IV and V, as found in our results.^{11,13} The isolates detected by Ho et al. had SCCmec IV (46.4%) or V (53.6%),¹³ unlike Lamaro-Cardoso et al. who found SCCmec IIIA (57%) in most Brazilian children, which is widespread in hospitals in many countries worldwide, and only three MRSA

Table 1 – Microbiological aspects and genetic characteristics of 11 MRSA isolates carriage from infants attending DCCs in Vitória da Conquista, Brazil.

Isolates	SCCmec typing	Resistance profile ^a	MIC (µg/mL)	Phenotype ^b	Virulence genes profiles	ST/CC	spa type
(MRSA)		OXA	VAN				
C18	V	amp, ery	4	–	Moderate	spa	398/398
C48	IVA	amp, ery	4	2	Strong	spa	5/5
C54	IVA	amp	4	2	Strong	spa	5/5
C60	IVA	amp	4	2	Strong	seb, spa	2228/45
C77	IVA	amp, ery	4	–	Strong	spa	5/5
C80	IVA	–	4	–	Strong	spa	5/5
C83	V	amp, ery	4	2	Producer	seb, spa	398/398
C94	IVA	–	8	2	Strong	spa	5/5
C137	V	amp, cli, ery	4	–	Strong	spa	45/45
C138	IVA	amp, ery, tet	4	–	Strong	spa	5/5
C152	IVA	amp, ery, cip	4	–	Strong	spa	5/5

^a amp, ampicillin; ery, erythromycin; cip, ciprofloxacin; cli, clindamycin.^b Moderate, moderate biofilm producer; producer, biofilm producer; strong, strong biofilm producer.

strains showed SCCmec IV and one SCCmec type V genes.¹² Therefore the distribution of the MRSA clone varies throughout the world.

The SCCmec IV has been strongly associated with strains causing MRSA infections in patients without risk factors in Brazil¹⁴ and elsewhere.¹⁵ Our findings point out the potential of this specific DCCs to be a reservoir of emerging MRSA genotypes and highlight the need to enhance surveillance of these bacteria and control their transmission.

The MLST and spa typing revealed a small diversity between CA-MRSA isolates. ST5 clones have emerged in hospital and community isolates, while ST45 is most often found in the community carrying SCCmecIV.^{16,17} The Clonal complexes – CC5 and CC45 – which represented 81.8% of CA-MRSA isolates in this study, are among the clonal groups known to be involved in global pandemic caused by MRSA. Meanwhile, invasive infection by CC398 rarely occurs.¹⁸

High rates of MRSA and MSSA isolates resistant to ampicillin (80%) and erythromycin (32.8%) were found, as well as low resistance rates to clindamycin, tetracycline, and ciprofloxacin. Our findings agree with previous studies in the literature, except for higher susceptibility to erythromycin elsewhere (14–20%).⁷ Thus, isolates of CA-MRSA had limited resistance to non-β-lactams. The majority of MRSA isolates (63.6%) detected herein were resistant to more than two antibiotics and this is a challenge in controlling these infections. One isolate (1.43%) with reduced susceptibility to vancomycin among MSSA strains is an important finding.

Most cases of reduced vancomycin susceptibility in *S. aureus* (vancomycin-intermediate *S. aureus*, or VISA) reported in the literature are also methicillin-resistant. However, the development of VISA in MSSA isolates have been reported.^{19,20} Thus, reduced vancomycin susceptibility can occur in *S. aureus* irrespective of background methicillin susceptibility. There are no data in the literature of CA-MRSA strains susceptible to or with reduced susceptibility to vancomycin, so this fact is of particular concern in a healthy young population. Therefore, additional studies and continuous monitoring of *S. aureus* and MRSA are recommended.

Herein, all *S. aureus* isolates were able to produce biofilm, and the percentage of MRSA and MSSA strains producing biofilm was similar. There are no studies on biofilm production of *S. aureus* recovered from healthy individuals. There is no direct relationship of the methicillin resistance profile with greater or lesser ability to produce biofilm by the strains. The majority of biofilm producer isolates were classified as strong producers. This feature may act as an additional barrier to control an infection with antimicrobials.²

Protein A encoded by spa gene is a hallmark of *S. aureus*.¹⁵ Other virulence genes such as seb, sec, and pvl were detected in a few isolates. In a study of strains isolated from healthy students in Turkey, the sec gene was found to be the most frequent, followed by sea and none of the *S. aureus* isolates had pvl.²¹ The pvl gene has been shown to occur less frequently in CA-MRSA isolates associated with asymptomatic nasal colonization, consistent with the results of the present

Table 2 – Biofilm production by MRSA and MSSA isolates recovered from nasal secretion of healthy children attending daycare centers in Vitória da Conquista, Bahia, Brazil.

Samples	n	Non-producers		Weak producers		Moderate producers		Producers		Strong producers	
		n	%	n	%	n	%	n	%	n	%
MRSA	11	0	0	0	0	1	9.1	1	9.1	9	81.8
MSSA	59	0	0	4	6.7	7	11.8	12	20.3	36	61
Total	70	0	0	4	5.7	8	11.4	13	18.5	45	64.2

MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin-susceptible *S. aureus*.

study.²² The genes of staphylococcal enterotoxins have been implicated as important determinants of *S. aureus* virulence²; however, they have been poorly studied in healthy children in the community.

In conclusion, this study showed high detection rates of *S. aureus* and MRSA (7.4%) in healthy Brazilian day care children. Moreover, the characterized isolates with reduced sensitivity to vancomycin and biofilm production are of special concern. Thus, continuous surveillance of *S. aureus* and MRSA in Vitória da Conquista should be encouraged to gain a better understanding of the circulating staphylococci in order to establish better control of possible infections.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgments

This study was supported by CAPES. We thank the DCCs, children and parents/guardians for allowing this research to be conducted, and AcademicEnglishSolutions.com for proofreading.

REFERENCES

1. Shopsin B, Mathema B, Martinez J, Ha E, Campo ML, Fierman A. Prevalence of methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* in the community. *J Infect Dis.* 2000;182:359–62.
2. Watkins RR, David MZ, Salata RA. Current concepts on the virulence mechanisms of methicillin-resistant *Staphylococcus aureus*. *J Med Microbiol.* 2012;61:1179–93.
3. Zhang K, McClure JA, Elsayed S, Louie T, Conly JM. Novel multiplex PCR assay for characterization and concomitant subtyping of Staphylococcal cassette chromosome *mec* Types I to V in methicillin-resistant *Staphylococcus aureus*. *J Clin Microbiol.* 2005;43:5026–33.
4. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol.* 2000;38:1008–15.
5. Harmsen D, Claus H, Witte W, et al. Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for spa repeat determination and database management. *J Clin Microbiol.* 2003;41:5442–8.
6. Miller MB, Weber DJ, Goodrich JS, et al. Prevalence and risk factor analysis for methicillin-resistant *Staphylococcus aureus* nasal colonization in children attending childcare centers. *J Clin Microbiol.* 2011;49:1041–7.
7. Tavares DA, Sa-Leao R, Miragaia M, de Lencastre H. Large screening of CA-MRSA among *Staphylococcus aureus* colonizing healthy young children living in two areas (urban and rural) of Portugal. *BMC Infect Dis.* 2010;10:110.
8. CLSI. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Second Informational Supplement. CLSI document M100-S22. Wayne, PA: Clinical and Laboratory Standards Institute; 2012.
9. Proietti PC, Coppola G, Bietta A, Marenzoni ML, Passamonti Hyatt DR. Characterization of genes encoding virulence determinants and toxins in *Staphylococcus aureus* from bovine milk in Central Italy. *J Vet Med Sci.* 2010;72:1443–8.
10. Cassat JE, Lee CY, Smeltzer MS. Investigation of biofilm formation in clinical isolates of *Staphylococcus aureus*. *Methods Mol Bio.* 2007;391:127–44.
11. Lo WT, Lin WJ, Tseng MH, Lu JJ, Chu ML, Wang CC. Nasal carriage of a single clone of community-acquired methicillin resistant *Staphylococcus aureus* among kindergarten attendees in northern Taiwan. *BMC Infect Dis.* 2007;7:51.
12. Lamaro-Cardoso J, Lencastre H, Kipnis A, et al. Molecular epidemiology and risk factors for nasal carriage of *Staphylococcus aureus* and methicillin-resistant *S. aureus* in infants attending daycare centers in Brazil. *J Clin Microbiol.* 2009;47:3991–7.
13. Ho PL, Chiu SS, Chan MY, et al. Molecular epidemiology and nasal carriage of *Staphylococcus aureus* and methicillin-resistant *S. aureus* among young children attending daycare centers and kindergartens in Hong Kong. *J Infect.* 2012;64:500–6.
14. Gelatti LC, Bonamigo RR, Inoue FM, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* carrying SCCmec type IV in southern Brazil. *Rev Soc Bras Med Trop.* 2013;46:34–8.
15. David MV, Daum RS. Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. *Clin Microbiol Rev.* 2010;23:616–87.
16. de Sousa MA, Conceicao T, Simas C, De Lencastre H. Comparison of genetic backgrounds of methicillin-resistant and-susceptible *Staphylococcus aureus* isolates from Portuguese hospitals and the community. *J Clin Microbiol.* 2005;43:5150–7.
17. Xie X, Bao Y, Ouyang N, et al. Molecular epidemiology and characteristic of virulence gene of community-acquired and hospital-acquired methicillin-resistant *Staphylococcus aureus* isolates in Sun Yat-sen Memorial hospital, Guangzhou, Southern China. *BMC Infect Dis.* 2016;16:339.
18. Krziwanek K, Metz-Gercek S, Mittermayer H. Methicillin-resistant *Staphylococcus aureus* ST398 from human patients, upper Austria. *Emerg Infect Dis.* 2009;15:766–9.
19. Pillai SK, Wennersten C, Venkataraman L, Eliopoulos GM, Moellering RC, Karchmer AW. Development of reduced vancomycin susceptibility in methicillin-susceptible *Staphylococcus aureus*. *Clin Infect Dis.* 2009;49:1169–74.
20. Hu J, Ma XX, Tian Y, Pang L, Cui LZ, Shang H. Reduced vancomycin susceptibility found in methicillin-resistant and methicillin-sensitive *Staphylococcus aureus* clinical isolates in Northeast China. *PLoS One.* 2013;8:e73300.
21. Dağı HT, Findik D, Demirel G, Arslan U. Detection of methicillin resistance and various virulence factors in *Staphylococcus aureus* strains isolated from nasal carriers. *Balkan Med J.* 2015;32:171–5.
22. Boyle-Vavra S, Daum RS. Community-acquired methicillin-resistant *Staphylococcus aureus*: the role of Panton–Valentine leukocidin. *Laby Invest.* 2007;87:3–9.