

Molecular detection of cytomegalovirus, herpes simplex virus 2, human papillomavirus 16-18 in Turkish pregnant

ABSTRACT

Objective: Human cytomegalovirus (CMV) is the most common cause of viral intrauterine infections in the world. Herpes simplex virus type 2 (HSV-2) and human papillomavirus (HPV) are the main agents of viral sexually transmitted diseases, which cause genital ulcers and genital warts, respectively. HPV infection has been linked to the majority of the anogenital malignancies. The aim of this study was to detect the existence of CMV, HSV-2 and HPV type 16-18 in Turkish pregnant by using sensitive molecular assays. **Methods:** One hundred thirty-four women (18-41 years old; mean age \pm SD: 27 ± 8) applied to outpatient clinic of Obstetrics and Gynecology, in between 18th - 22nd weeks of their pregnancy and a control group of 99 healthy women (15-39 years old; mean age \pm SD: 24 ± 8) were included in the study. Cervical smear samples were used for DNA extraction. CMV, HSV-2 and HPV 16-18 detections were carried out by real time PCR and in house PCR method, respectively. **Results:** Three patients (3/134; 2.2%) were found to be positive for each HPV and HSV-2. Dual infection with HPV and HSV was found in just one patient. HPV 18 was detected in all positive samples. CMV was found to be positive in two patients (2/134; 1.4 %). **Conclusion:** HPV, HSV and CMV must be screened due to high prevalence of these viruses in pregnant by using sensitive molecular methods.

Keywords: pregnancy; infection; polymerase chain reaction.

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INTRODUCTION

Cytomegalovirus (CMV) is the most common cause of congenital infections affecting 0.5% to 2% of all live births in the developed world.¹ Intrauterine infection occurs in up to 50% of pregnancies following primary maternal infection, and 10% of the infected infants are symptomatic at birth.^{2,3} However nearly 10%-15% of the asymptomatic newborns develop late sequelae, particularly sensorineural hearing loss and neurodevelopmental disorders.⁴

Genital herpes continues to be a public health problem in both developed and developing countries. Prevalence of HSV-2 in different populations has shown various results from less than 10% to more than 40%.⁵ Laboratory confirmation for clinical diagnosis of HSV-2 is important, particularly as there are other conditions which present similarly to genital herpes, while atypical presentations of genital herpes also occur.⁶

Uterine cervix carcinoma is the second most common type of female cancers and also the second highest mortality cause for women. Human papillomavirus (HPV) being detected in close to 99.7% of cervical squamous cell carcinomas is imputed to be a necessary factor that causes cervix carcinoma. Hormones and their metabolism alteration have also been implicated as cofactors in HPV proliferation and the cervical cancer pathogenesis. It is well known that high risk HPV infections are more frequent in pregnant women.⁷⁻⁹ The polymerase chain reaction (PCR) technology for CMV, HSV, HPV detection is now highly sensitive and can detect viral DNA in exfoliated cells irrespective of the sampling method used.¹⁰

The aim of this study was to detect CMV, HSV-2, and HPV type 16-18 from cervical smear samples of pregnant women by PCR based methods and to improve an algorithm for follow-up of positive patients.

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We declare no conflict of interest.

MATERIAL AND METHODS

Study group

This study was performed at the Gynecology and Obstetrics Outpatient Clinic at Gazi University Hospital, Ankara, Turkey. One hundred thirty-four women (18-41 years old; mean age \pm SD: 27 \pm 8) applied to outpatient clinic, in between 18th - 22nd weeks of their pregnancy and a control group of 99 healthy women (15-39 years old; mean age \pm SD: 24 \pm 8) followed for routine prenatal visit, were included in the study. Cervical smear samples were used to detect CMV, HSV-2 and HPV. All the patients and the control group answered a questionnaire including age of first sexual intercourse, number of sexual partners and condom use.

Specimen sampling and preservation

Cervical smear samples collected by sterile swabs were sent to the laboratory in phosphate buffered saline (PBS) (pH 8.6). The samples in PBS were vortexed and stored in -86°C until they were tested. And 10 mL of blood was taken from each patient for serological examination of HSV-2.

DNA extraction method for HPV, HSV and CMV

Following the centrifugation at 13,000 rpm for 3 min, the samples in PBS were treated with lysis solution containing 20 mg/mL proteinase K [20 mM (NH₄)₂SO₄, 75 mM Tris HCl (pH 8.8) 0,1% Tween 20]. After discarding the supernatant, precipitate was extracted by phenol-chloroform isoamyl alcohol method.¹¹ Precipitation of DNA was succeeded by using absolute ethanol including 3M sodium acetate at -20°C overnight. On the following day, samples were mixed with 70% alcohol and dried. After drying, they were incubated on dry block at 55°C for 10 min and then 50 μ L sterile deionised water was added.

Amplification of HPV DNA

The primers used in this study were the consensus primers for all HPV genotypes and were chosen from L1 location of HPV genome which is associated with major viral capsid protein metabolism. MY09/MY11 primers were used for amplification of HPV.¹² Caski cell DNA was used as a positive control.¹³ During all amplification procedure, a negative control including no targeted DNA was used. Amplification procedure was performed by adding 5 μ L extracted DNA to a mixture of 45 μ L including 100 mM from each dNTP (dATP, dCTP, dGTP, dTTP) 100 pmol from each primer and 1 unit of Taq DNA polymerase enzyme (DNA mp ltd., Hants, UK) in 4 mM MgCl₂, 50 mM KCl₂, and 10 mM Tris HCl (pH 9.0).

A thermal cycler programme with 35 cycles including initial denaturation at 94°C for 5 min, denaturation

at 94°C for 20 seconds, annealing at 55°C for 45 seconds and extension at 72°C for 1 min, followed by 7 min of final extension at 72°C was performed. PCR products were analyzed on agarose gel stained with ethidium bromide and bands were detected with UV transluminator encompassing 450 bp for HPV.¹¹

Genotyping

MY09/MY14 (5' - CAT ACA CCT CCA GCA CCT AA - 3') and MY09/WD74 (5'-GGA TGC TGC ACC GGC TG A-3') specific primers were used for amplification of HPV type 16 and HPV type 18, respectively.^{12,14}

Amplification procedure was performed by adding 5 μ L pure DNA to a mixture of 95 μ L including 100 mM from each dNTP (dATP, CTP, dGTP, dTTP) 100 pmol from each primer and 1 unit of Taq polymerase in 4 mM MgCl₂, 50 mM KCl₂, and 10 mM Tris HCl (pH 9.0). We used a thermal cycler programme for HPV genotyping as described above. PCR products were analyzed on agarose gel stained with ethidium bromide and transilluminating band was detected with UV transluminator encompassing 110 bp for HPV 16 and 140 bp for HPV 18.

Real-time PCR for CMV and HSV-2

Real-time PCR reactions were carried out by using Ligthcycler (Roche Diagnostics, Germany). Ligthcycler DNA master hybridization probes kit (Roche) was used for amplification. Taqman probes labeled with FAM at their 5' end and labeled with TAMRA as quencher at their 3' end, and specific primer sets for HSV DNA polymerase gene region and CMV pp65 gene region (Table 1) were used for amplifications (Metis Biotechnology Ankara, Turkey).

Table 1. Primers and probes

Agent Region Primers (5 > 3)
HSV DNA polymerase CAT CAC CGA CCC GGA GAG GGA C
HSV DNA polymerase GGG CCA GGC GCT TGT TGG TGT A
CMV pp65 CTT GCC CTG GAT GCG ATA CT
CMV pp65 ACG GCT TTA CGG TGT TGT GT
Probes
HSV LC (TaqMan Probe) FAM- CCG CCG AAC TGA GCA GAC ACC CGC GC -TAMRA
CMV LC (TaqMan Probe) FAM- CGC GAG ACC GTG GAA CTG CG -TAMRA

In each assay, five CMV AD 169 DNA standards from 2×10^2 to 2×10^6 CMV DNA copy/mL were used as external standards. A capillary tube containing distilled water instead of DNA was used as negative control in each run. The detection limit was established as 200 CMV DNA copy/mL in blood. Serial dilutions containing 2×10^2 - 2×10^6 HSV DNA were prepared for HSV and a limit was detected as 200 HSV DNA copy/mL. A negative control was used in each study.

CMV serology

Anti-CMV IgM antibody: specific IgM responding to CMV in cervicovaginal smear samples was studied with two different methods: microparticle enzyme immunoassay (MEIA) system and its appropriate anti-CMV IgM kit (Imx System, Abbott Laboratories, Abbott Park, IL, USA) and a commercial diagnostic anti-CMV IgM enzyme-linked immunosorbent assay (ELISA) kit (Radim SPA, Pomezia-Rome, Italy).

Anti-CMV IgG antibody: specific IgG responding to CMV in cervicovaginal smear samples was studied with MEIA system and its appropriate anti-CMV IgG kit (Axsym System, Abbott Laboratories, Abbott Park, IL, USA).

CMV IgG avidity test: the cervicovaginal smear sample with 4 U/mL of anti-CMV IgG were studied with a commercial diagnostic CMV IgG avidity Enzyme immunoassay (EIA) kit (Radim SPA, Pomezia-Rome, Italy) according to the manufacturer's instructions.

HSV serology

HSV-2 IgM and IgG antibodies were detected by EIA (Meddens Diagnostics B.V., The Netherlands) in the sera samples. Quantitative EIA was performed for HSV 2 IgG Ab according to the manufacturer's instructions.

RESULTS

Patients and USG findings

All pregnant and the control group were monogamic and had no history of sexual intercourse before getting married. The reported age at first sexual intercourse was less than 20 years for 98 (73.1%) pregnant, and 67 (67.7%) of control group respectively. Of the 134 pregnant, 32 (23.8%) and

99 controls 35 (35.4%) reported ever use of condoms for STDs. Of the 134 pregnant, 22 had abnormal ultrasonographic findings, four oligohydramnios, four ventriculomegaly, three intrauterine growth retardation (IUGR), three IUGR + oligohydramnios, two polyhydramnios, one bilateral choroid plexus cyst greater than 7 mm, one echogenic bowel + choroid plexus cyst, one polihydramnios, one omphalocele, one multiple structural abnormalities and one hydrops fetalis were detected.

Out of three pregnant with IUGR + oligohydramnios, one was found to be positive for CMV. All other pregnant with abnormal ultrasonographic findings were found to be negative for each of the three viruses. No abnormal USG findings were detected in any of the control group.

Serology for CMV and HSV

Anti-CMV IgM was found to be negative in all of the cervicovaginal smear samples. In all of the samples studied, one cervicovaginal smear sample was found to be 4 U/mL, with negative result. Because both tests for anti-CMV IgM, and CMV IgG were designed for only serum evaluation, the cut-off values for smear may be lower than the cut-off values (10 U/mL) accepted for serum. In this respect the cervicovaginal smear sample with 4 U/mL of anti-CMV IgG value was also approved as positive and studied with CMV IgG avidity test.

CMV IgG avidity test: according to the kit that we used, we considered the values below 35% as a low avidity, values between 35% and 45% as a gray zone and above 45% as a high avidity. Low avidity is a finding in favor of a primary acute infection. The cervicovaginal smear sample with 4 U/mL of anti-CMV IgG value was also studied with CMV IgG avidity test and again low avidity (21%) was found. This situation was also suggested a primary acute infection.

HSV-2 Ig M and IgG were positive in the sera samples of 6 (4.4 %) and 9 (9 %) of pregnant, respectively.

In house PCR and RT PCR for CMV, HPV and HSV-2

CMV DNA was found to be positive in two (1.5%) of pregnant women by real-time PCR. In control group there was no CMV DNA positivity. In one of the CMV positive pregnant women, there were abnormal USG findings (oligohydroamnios and *in utero* growth retardation) (Table 2).

Table 2. One of two CMV positive pregnant had abnormal ultrasonographic finding. CMV DNA was negative for HPV type 18 positive 3 pregnant, while one of 3 was detected positive for HSV-2

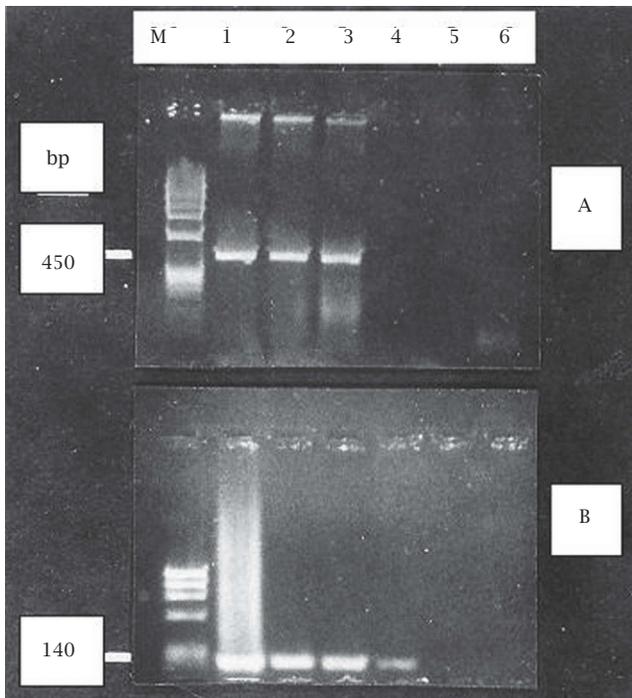
	CMV		HSV-2		Total
	Positive	Negative	Positive	Negative	
HPV Type 18	Positive	-	3 (2.1%)	1 (0.7%)**	2 (1.4%)
	Negative	2 (1.4%)	132 (98.6%)	2(1.4%)	132(98.6%)
Abnormal USG	1*(5%)	20 (95%)	-	21 (100%)	21(100%)
Total	2 (1.4%)	132 (98.6%)	3 (2.1%)	131(97.8%)	134 (100%)

*: abnormal USG findings

**::dual infections (HPV and HSV-2)

Three (3/134; 2.2%) of pregnant women were found to be positive for only HPV 18. No other types of HPV were detected in any of the pregnant (Figure 1). In control group, there was no HPV DNA positivity.

Figure 1: A; (HPV) M: fX174 DNA / *Hae III* marker. Lanes 1, 2, 3: HPV positive patients. Lanes 4, 5: Negative control, B; (HPV type 18), M: fX174 DNA / *Hae III* marker. Lanes 1, 2, 3: HPV type 18 positive patients. Lane 4: Positive control. Lane 5: Negative control.



Three (3/134; 2.2%) of the samples from the pregnant women were found to be positive for HSV-2. Dual infection with HPV and HSV positivity was found in one patient. Any gynecological examination findings were not found out in favor of HPV and HSV in any of the patients (Table 2).

None of the pregnant and the control group had triple infection with CMV, HSV-2 and HPV. While there was no positivity for both CMV and HPV, HSV was found to be positive in 2 (2%) samples in control group.

DISCUSSION

In this study, of the 134 pregnant women, CMV DNA was found to be positive in two (1.5%) patients, while no CMV DNA positivity was found by real-time PCR in control group. In one of the CMV positive pregnant women, there were abnormal USG findings (oligohydroamnios and *in utero* growth retardation). She was followed for IUGR and *in utero* exitus happened during the 20th week of pregnancy.

Autopsy and CMV PCR from these materials could not be performed since the family did not give permission. The other CMV DNA positive pregnant woman gave a healthy birth and no problems occurred with the mother and baby during follow-up.

Previous studies have shown that the incidence of CMV excretion from cervix increases while pregnancy proceeds.¹⁵ In our study group, the patients were at their second trimester. In Turkey, Koksal *et al.*,¹⁶ Kaleli *et al.*¹⁷ and Satilmis *et al.*¹⁸ reported the prevalence of CMV IgG in pregnant women as 79%, 94.7% and 98.5%, respectively. CMV IgM positivity in pregnancy was reported as 0.66% by Kaleli *et al.*¹⁷ It is determined that virus spread with asymptomatic infection is frequent among pregnant women and virus can be isolated from cervical mucus and urine of 2%-28% of pregnant women.¹⁹ There are several potential routes of infection of the placenta during pregnancy, including ascending infection from the genital tract as well as hematogenous transmission. As CMV secretion from cervix increases during pregnancy, a large spectrum of cells of the fetus is infected by CMV. The major target fetal organs for CMV infection are the lungs, pancreas, kidneys and the liver but comparing with these organs, CMV DNA level determined in uterine tissue and cervical smear is higher.²⁰

In our study, 134 pregnant women were screened for HSV-2 DNA. Three (2.2%) pregnant women and 2 (2%) women in control group were found to be positive in favour of HSV-2. In our country, HSV-2 is rarely seen due to the monogamy in the sexual relations. Genital herpes resulting from HSV-2 infection of the urogenital tract is a worldwide public health problem. European investigations of pregnant women have shown overall prevalence of HSV-2 to range between 9% and 33%.²¹ HSV-2 DNA prevalence was reported as 2% in Turkish pregnant recently.²² Transmission of HSV is usually via direct contact with lesion while HSV-2 is classically transmitted sexually.⁶ The women most at risk for mother to child transmission are the ones who are previously uninfected. New onset HSV infection in late pregnancy carries a 30%-50% risk of neonatal infection since there is not an adequate time for antibodies to develop and suppress the viral replication before labor, while infection in early pregnancy carries a risk of less than 1%.^{6,23} Fortunately the risk of fetal infection in recurrent episodes is nearly 2%.²⁴ Risk for baby usually happens during delivery via contact with genital lesions with recurrent episodes of viral shedding; therefore, caesarean section is recommended in these cases.²⁵ In pregnant population, real-time PCR has some advantages, making results available at a much earlier time, thus significantly reducing the need for empirical treatment.^{23,26} Dual infection with HPV and HSV positivity was found in one patient in our study. Any gynecological examination findings were not found out in favor of HPV and HSV in any of the patients. Of the three

HSV-2 DNA positive patients, all were HSV-2 IgM positive with no physical symptoms. These patients did not have a history of condom use before pregnancy and their first sexual intercourse were after the age of 20. Although they were asymptomatic, caesarean section was recommended for HSV positive three patients in our hospital and no problems occurred during the follow up of the mothers and babies.

Studies about HPV show that its transmission mode remains unclear. HPV is positive in the airways of neonates and in maternal genital condylomas, which suggests that it is transmitted in the birth canal, from mothers with genital condylomas.²⁷ Since the serologic diagnosis of HPV is inadequate, molecular diagnosis has been more popular in recent years.²⁸ Epidemiological studies revealed different results in pregnant women. Smith *et al.*,²⁹ Gajewski *et al.*³⁰ and Takakuwa *et al.*³¹ have found out 29%, 26% and 12.5% of HPV positivity respectively in pregnant women taking routine obstetric care. In a study performed in our country, HPV DNA was found as 9.5% by PCR. One patient was found type 16/30, the others were untyped.³² We have found that 2.2% of the patients were positive for HPV 18 in our study where Smith *et al.*²⁹ found that 2.6% of the patients were positive in favor of HPV 18. The association between HSV-2 and HPV 18 was found in one of the pregnant women. This association may be due to the similar transmission route of both viruses. During the neonatal follow-up of baby HPV DNA was not found in the laryngeal swab by Real Time PCR.

CONCLUSION

Since HPV is considered as an infection predisposed to malignancies, HPV detection with molecular methods should be considered as a routine screening test for women applied to obstetrics and gynecology clinics. This screening policy should also contain CMV and HSV infections because of their potential transmission risks for the baby. We suggest that our assays provide useful information about CMV, HSV, HPV infections in the fetus and the neonate; and obstetricians have to make informed decisions about the management of pregnancies complicated by CMV, HSV, and HPV infections.

REFERENCES

- Azam AZ, Yvan V, Fawer CL *et al.* Prenatal diagnosis of congenital cytomegalovirus infection. *Obst and Gynecol* 2001; 97:443-8.
- Yamamoto AY, Mussi-Pinhata MM, Pinto PCG *et al.* Usefulness of blood and urine samples collected on filter paper in detecting cytomegalovirus by the polymerase chain reaction technique. *J Virol Meth* 2001; 97:159-64.
- Schleiss MR, Bourne N, Bravo FJ *et al.* Quantitative-competitive PCR monitoring of viral load following experimental guinea pig cytomegalovirus infection. *J Virol Meth* 2003; 108:103-10.
- Gindes L, Teperberg-Oikawa M, Sherman D *et al.* Congenital cytomegalovirus infection following primary maternal infection in the third trimester. *BJOG* 2008; 115:830-5.
- Eskild A, Jeansson S, Hagen JA *et al.* Herpes simplex virus type-2 antibodies in pregnant women: the impact of the stage of pregnancy. *Epidemiol Infect* 2000; 125:685-92.
- Kriebs JM. Understanding herpes simplex virus: transmission, diagnosis and considerations in pregnancy management. *J Midwifery Womens Health* 2008; 53:202-8.
- Armbruster-Moraes E, Ioshimoto LM, Leao E, Zugaib M. Prevalence of high risk human papillomavirus in the lower genital tract of Brazilian gravidas. *Intern J Gynecol and Obst* 2000; 69:223-7.
- Chan PK, Chang AR, Tam WH *et al.* Prevalence and Genotype Distribution of Cervical Human Papillomavirus Infection: Comparison Between Pregnant Women and Non-Pregnant Controls. *Journal of Medical Virology* 2002; 67:583-8.
- Mayeaux EJ. Reducing the economic burden of HPV-related diseases. *J Am Osteopath Assoc* 2008; 108:52-7.
- De Sanjose S, Bosch XF, Munoz N *et al.* Screening for genital human papillomavirus: results from an international validation study on human papillomavirus sampling techniques. *Diag Mol Path* 1999; 8:26-31.
- Maniatis T, Fritsch EF. Molecular Cloning. In: J Sambrook ed. *A Laboratory Manual*. New York: 1990.
- Baay MFD, Quint WGV, Koudstaal J *et al.* Comprehensive study of several general and type-specific primer pairs for detection of human papillomavirus DNA by PCR in paraffin-embedded cervical carcinomas. *J Clin Microbiol.* 1996; 34:745-7.
- Tuncer S, Ustaçelebi S. Detection of Human Papillomavirus Types 16 and 18 by Polymerase Chain Reaction in Cervical Biopsy Specimens. *Flora* 1996; 1:40-4.
- Poljak M, Seme K. Rapid detection and typing of human papillomaviruses by consensus polymerase chain reaction and enzyme-linked immunosorbent assay. *J Virol Meth* 1996; 56:231-8.
- Chandler SH, Alexander ER, Holmes KK. Epidemiology of cytomegalovirus infection in a heterogeneous population of pregnant women. *J Infec Dis* 1985; 152:249-56.
- Koksal I, Alnaci M, Kardes OB, Aydemir V. Toxoplasma, rubella and cytomegalovirus seropositivity ratios among adults in Eastern Black Sea Region. *Turk Bullet Microb* 1994; 28:58-61.
- Kaleli B, Kaleli I, Yurdakul B, Aksit F. Rubella and Cytomegalovirus Infection in Pregnancy. *Turk J Infect.* 1997; 11:325-7.
- Satilmis A, Gura A, Ongun H *et al.* CMV seroconversion in pregnant and the incidence of congenital CMV infection. *Turk J Pediatr.* 2007; 49(1):30-6.
- Gibbs RS, Sweet RL. Maternal and fetal infectious disorders. In: Creasy RK, Resnik R eds. *Maternal Fetal Medicine*. Philadelphia: W.B. Saunders, 1994.
- Fisher S, Genbacev O, Maidji E, Pereira L. Human cytomegalovirus infection of cytotrophoblasts in vitro and *in utero*: Implications for transmission and pathogenesis. *Journal of Virology* 2000; 74(15):6808-20.
- Gaytant MA, Steegers EAP, Van Laere M *et al.* Seroprevalence of herpes simplex virus type 1 and type 2 among pregnant women in the Netherland. *Sex Transm Dis.* 2002; 29:710-4.
- Rota S, Bozdayi G, Dogan B, Dinc B. Investigation of herpes simplex virus DNA by real-time polymerase chain reaction in the clinical samples. *Bulletin of Microbiology* 2004; 38:233-9.
- Perkins D, Chong H, Irvine B, Domagalski J. Genital co- infection with herpes simplex viruses type 1 and 2; comparison of real time PCR assay and traditional viral isolation methods. *J Cell Mol Med* 2007; 11(3):581-4.

24. Poeran J, Wildshut H, Gaytant M *et al.* The incidence of neonatal herpes in The Netherlands. *J Clin Virol* 2008; 42 (4):321-5.
25. Moss NJ, Harper CC, Ahrens K *et al.* Predictors of incident herpes simplex virus type 2 infections in young women at risk for unintended pregnancy in San Fransisco. *BMC Infect Dis* 2007; 7:113.
26. Gouarin S, Gault E, Vabret A *et al.* Real-Time PCR quantification of human cytomegalovirus DNA in amniotic fluid samples from mothers with primary infection. *J Clin Microbiol* 2002; 40:1767-72.
27. Smith EM, Johnson SR, Pignatari S *et al.* Perinatal vertical transmission of human papillomavirus and subsequent development of respiratory tract papillomatosis. *Ann Otol Rhinol Laryngol* 1991; 100:479-83.
28. Collier L, Oxford J Papovaviridea. In: Collier L. ed. *Human Virology*. New York: Oxford University Press, 2000.
29. Smith EM, Ritchie JM, Yankowitz J *et al.* HPV prevalence and concordance in the cervix and oral cavity of pregnant women. *Infect Dis Obstet Gynecol* 2004; 12:45-56.
30. Gajewski M, Wielgos M, Kami ski P *et al.* The occurrence of genital types of Human papillomavirus in normal pregnancy and in pregnant renal transplant recipients. *Neuro Endocrinol Lett* 2006; 27:529-34.
31. Takakuwa K, Mitsui T, Iwashita M *et al.* Studies on the prevalence of Human papillomavirus in pregnant women in Japan. *J Perinat Med* 2006; 34:77-9.
32. Guney AI, Ince U, Kullu S *et al.* Detection and typing of human papillomavirus in cervical specimens of Turkish women. *Eur J Gynaecol Oncol* 1997; 18(6):546-50.